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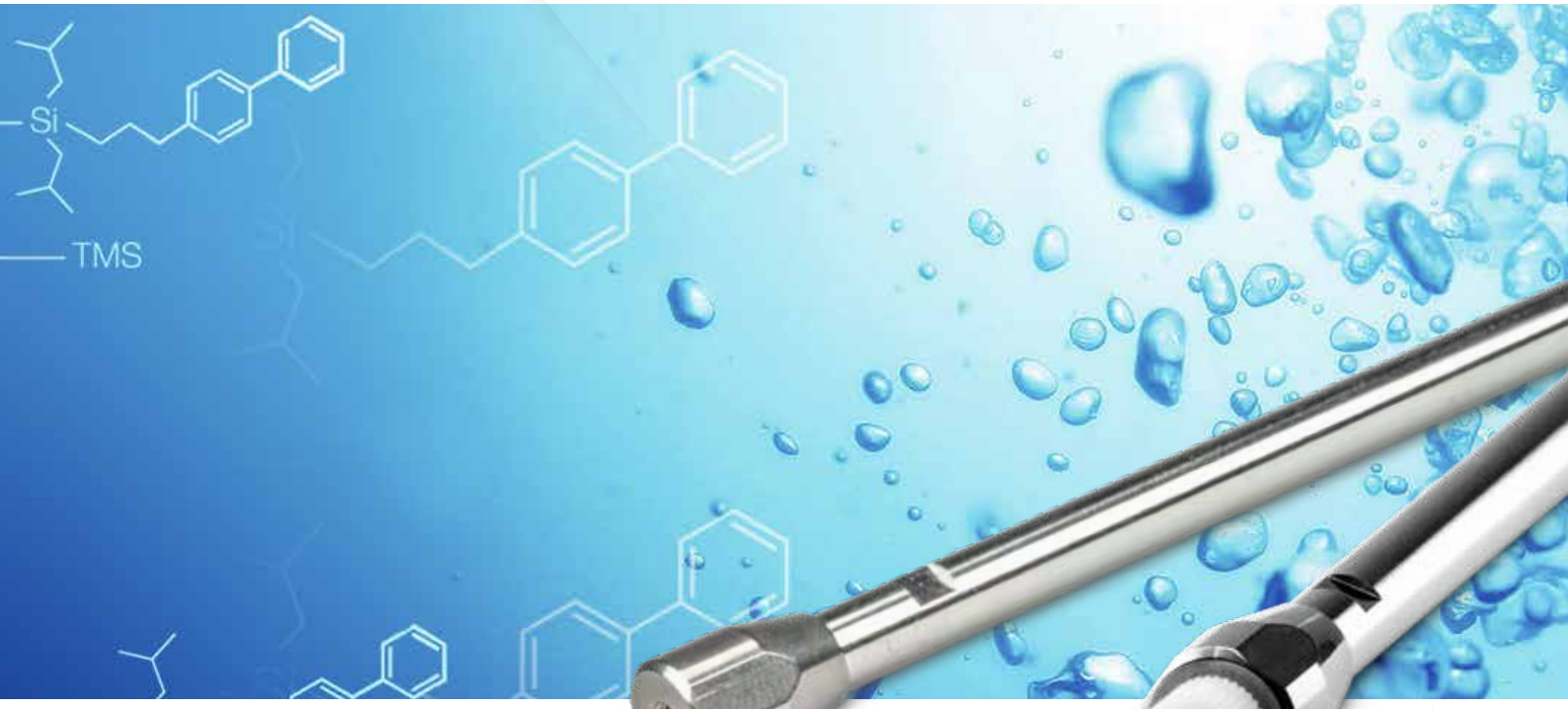
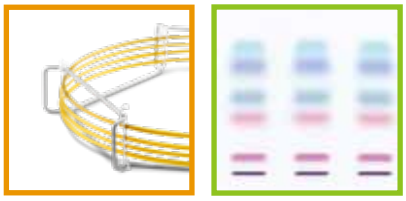
Chromatography Consumables

Lab Equipment Line

HPLC Column | GC Column | TLC





MACHEREY-NAGEL

Columns and supplies

REVODIX
since 1995 레보딕스(주)

Catalog No. **LAB-C13-1**

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Basics

High performance liquid chromatography (HPLC) is part of liquid chromatographic separating processes of substance mixtures and their analysis. At the beginning the technique was also called high pressure liquid chromatography due to the high back pressure of the column. HPLC offers qualitative (identification of substances) and quantitative (concentration determination) analysis by comparison with standard substances. The term HPLC was introduced in the 1970s, for the delineation of the high-performance method to the in the 1930s developed column liquid chromatography (column chromatography). At the beginning of the 21st century the HPLC was complemented by the even more efficient UHPLC (ultra high performance liquid chromatography). Hereby even higher pressures (> 400 bar) result in shorter analysis time and enhanced efficiency enabling a higher sample throughput with smaller sample volumes.

Application

HPLC / UHPLC is used additionally to gas chromatography (GC) for separation and determination of complex substance mixtures composed of low-volatile, polar and ionic, high-molecular or thermal instable substances. Therefore a sufficient solubility of the sample in a solvent or a solvent mixture is required. HPLC / UHPLC is used for purity control of chemicals and industrial products, determination of active agents for drug development, production and testing, environmental analytics, quality and purity control of foods, analysis of ingredients in cosmetics as well as for the isolation of biopolymers.

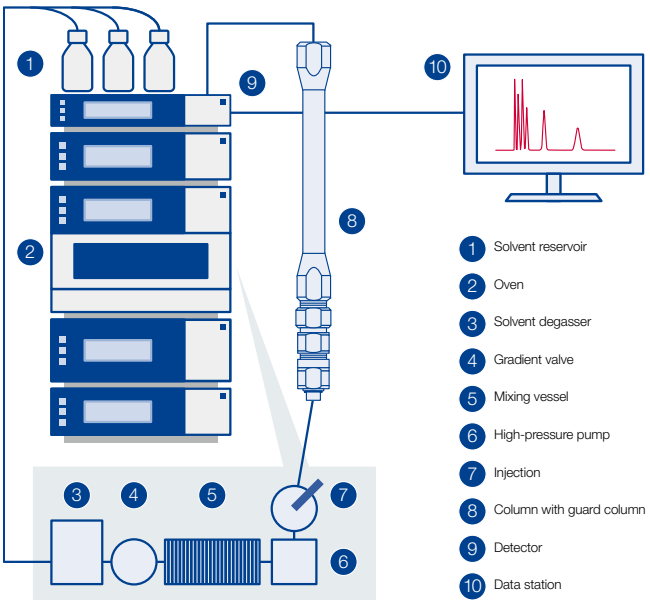
Basic principle

In liquid column chromatography a mobile phase (eluent) flows through a particle filled tube (separation column, stationary phase). In classic column chromatography this tube is a glass column with an inner diameter of several centimeters and a length up to 450 mm or even bigger. The filling material typically consists of coarse-grained particles like silica gel 60. The eluent is transported through the separation column either by hydrostatic pressure or a low-pressure pump with 1.5-2 bar.

In contrast HPLC columns consist of stainless steel with an inner diameter of 2-4.6 mm and a length of 20-300 mm. The column packing, mostly modified porous silica, has generally a particle size of 3, 5, 7 or 10 µm and a pore size of 50, 100, 120 (for low-molecular analytes) or 300-4000 Å (for high-molecular analytes). In UHPLC shorter columns in the range of 20-150 mm length with highly efficient particles of 1.8 µm size (sub-2 µm) are utilized. A guard column of a few millimeters length can be utilized and installed with a specific Column Protection System to increase the column lifetime. HPLC / UHPLC uses a high-pressure pump to transport the eluent from a storage vessel into the system with a column back pressure of up to 600 / 1200 bar.

Instrument

HPLC as well as UHPLC instruments have different building blocks. The storage vessel (eluent reservoir, 1) usually contains a deaerator unit (3) for the solvents. Followed by a gradient valve (4) with mixing chamber (5) in flow direction, which allows the usage of isocratic as well as gradient methods. A high-pressure pump (6) transports the sample into the system. The sample is injected via an injection valve (7). Usually this is operated automatically with a syringe by an autosampler. With the eluent flow the sample is transported to the guard and the separating column (8). For better reproducibility of the separation tempering with a column oven (2) should be performed. The separated substances are determined with a detector (9). In the resulting chromatogram each detector signal of a substance (peak), is related to the retention time of the column. With the data evaluation (10) these peaks can be identified and their concentration can be determined.



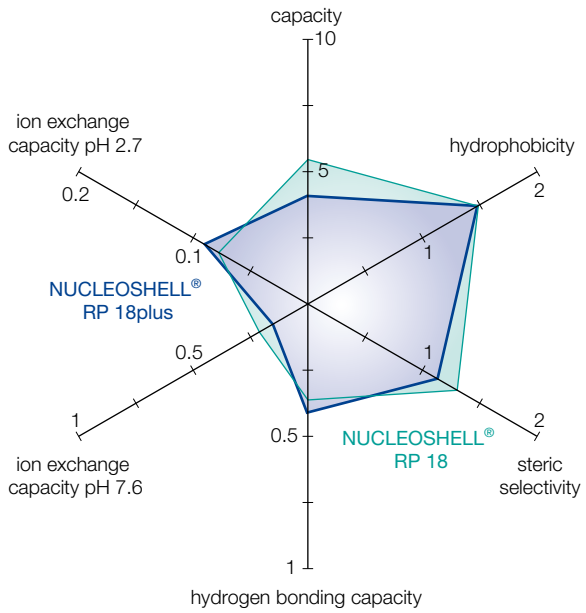
Separation mechanism

While flowing through the column each component of the solved mixture interacts differently with the stationary phase. According to the characteristics of the substance (hydrophobic, polar, ionic, aromatic, sterically hindered etc.) the strength of the interactions vary and thus the compounds are retained by the stationary phase in different ways. Essentially a distinction is drawn between normal phase (NP), reversed phase (RP) and ion exchange chromatography. Depending on the structure of the stationary phase diverse interactions e.g., van der Waals forces or π-π-stacking can occur and different polar mobile phases are required. For polar stationary normal phases (e.g., SiOH, CN, OH, NH_2) non-polar eluents like *n*-heptane, hexane, dichloromethane or 2-propanol are applicable. While for reversed phases (e.g., C_{18}, C_8, C_4, C_2, C_6H_5) typically polar RP eluents (e.g., acetonitrile or methanol with ultrapure water or buffer) and for ion exchange (e.g., SA, SB) aqueous buffers (e.g., phosphate, acetate, citric buffer) come to use.

Selectivity

The characteristic separation behavior of phases under certain conditions is also called selectivity. This is dependent on different parameters like structure and modifications of the base silica gel, nature of the chemical binding or the type of endcapping.

In recent decades several methods have been developed to compare and distinguish the selectivity of various silica gels and their modifications. In this connection defined substances or substance classes are analyzed and the chromatographic parameters are graphically presented. A frequently applied model in specialist literature is e.g., the TANAKA plot, which allows a quick comparison of different HPLC phases. [4]

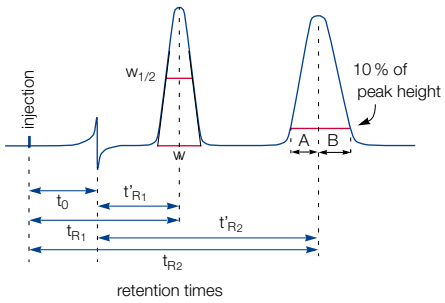


Parameter of the Tanaka diagram:
Capacity = k' (pentylbenzene)
Hydrophobicity = α (pentylbenzene, butylbenzene)
Steric selectivity = α (triphenyl, *o*-terphenyl)
Hydrogen bonding capacity (capacity of silanol) = α (caffeine, phenol)
Ion exchange capacity at pH 2.7 = α (benzylamine, phenol)
Ion exchange capacity at pH 7.6 = α (benzylamine, phenol)

The comparison of NUCLEOSHELL® RP 18 and NUCLEOSHELL RP® 18plus for example shows a lower ion exchange capacity at pH 7.6 for the monomeric NUCLEOSHELL® RP 18plus. The radar chart also reflects a more pronounced steric selectivity of NUCLEOSHELL® RP 18 due to a higher density of modifications with C_{18} chains.

Characteristic parameters

The success of a chromatographic separation depends apart from the stationary and mobile phase also on other characteristics like the quality of the separating column or the linear flow rate. The following schematic chromatogram illustrates the most important parameters which characterize a separation.



Schematic chromatogram

Peak width:	
$w_{1/2}$	peak width at half height
w	peak width of the peak (intersection point of the inflectional tangents with the zero line)
Peak symmetry:	
A	peak front to peak maximum at 10 % of peak height
B	peak maximum to peak end at 10 % of peak height
Retention time::	
t_0	dead time of a column = retention time of a non-retarded substance
t_{R1}, t_{R2}	retention times of components 1 and 2
t'_{R1}, t'_{R2}	net retention times of components 1 and 2

In a chromatographic system the substances differ from each other in their retention time in or on the stationary phase. The time, which is needed by a sample component to migrate from column inlet (sample injection) to the column end (detector) is the retention time t_{R1} or t_{R2}. The dead time t_0 is the time required by an inert compound to migrate from column inlet to column end without any retardation by the stationary phase. Consequently, the dead time is identical with the retention time of the sample component remaining in the stationary phase. The difference of total retention time and dead time yields the net retention time t'_{R1} or t'_{R2}, which is the time a sample component remains in the stationary phase.

$$t'_{R1} = t_{R1} - t_0 \quad \text{bzw.} \quad t'_{R2} = t_{R2} - t_0$$

To compare chromatograms that are recorded with columns of different lengths and internal diameters, as well as different flow rates, the retention time is converted into a dimensionless capacity factor k'.

$$k'_1 = \frac{t_{R1} - t_0}{t_0} \quad \text{bzw.} \quad k'_2 = \frac{t_{R2} - t_0}{t_0}$$

The relative retention α, also known as the separation factor, describes the ability of a chromatographic system (stationary and mobile phase) to distinguish between two compounds. This is calculated from the rate of the capacity factors of the substances, where the figure in the denominator is the reference compound.

$$\alpha = \frac{k'_2}{k'_1}$$

The resolution R is a measure for the efficiency of the column to separate two substances. Besides the retention time t_R the peak width at half height $w_{1/2}$ is also included.

$$R = 1.18 \cdot \frac{t_{R2} - t_{R1}}{(w_{1/2})_2 + (w_{1/2})_1}$$

For practical reasons the peak symmetry is calculated at 10 % of peak height. Ideally symmetry should be 1, i.e. A = B. Values > 1 indicate peak tailing, while values < 1 indicate peak fronting.

$$\text{Peak symmetry} = \frac{B}{A}$$

Instead of the mobile phase volumetric flow rate [mL/min], which is controlled at the HPLC instrument, it is advantageous to use the linear velocity u [cm/sec]. The linear velocity is independent of the column cross section and proportional to the pressure drop in the column. The linear velocity can be calculated by means of the dead time, where L is the column length in cm and t_0 the dead time in sec.

$$u = \frac{L}{t_0}$$

The quality of a column packing is determined through the number of theoretical plates N. High N values indicate a high capability to separate complex sample mixtures.

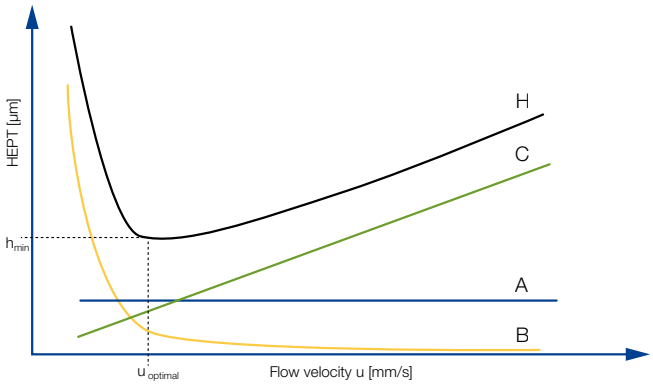
$$N = 5.54 \cdot \left(\frac{t_{R1}}{w_{1/2}} \right)^2$$

The value of the height equivalent to a theoretical plate HEPT is a criterion for the quality of a column. HEPT, is the length, in which the chromatographic equilibrium between mobile and stationary phase has been adjusted once. Its value depends on the particle size, the flow velocity, the mobile phase viscosity and especially on the packing quality. Small HEPT values, meaning a large number of theoretical plates N, facilitate the column to separate complex sample mixtures.

$$H = \frac{L}{N}$$

The Van Deemter equation shows the dependence of the HEPT on the velocity u.

$$H = A + \frac{B}{u} + C \cdot u$$



A term = eddy-diffusion, B term = longitudinal diffusion coefficient, C term = mass transfer coefficient, H = HEPT = height equivalent to a theoretical plate

The A term, also called eddy-diffusion, is a function of the particle size, the B term a function of the diffusion coefficient of the substance in the mobile phase and the C term the retardation of a substance by the interface between stationary and mobile phase. In the point of intersection of h_{min} and u_{opt} the optimal separation efficiency for a column with high peak symmetry for the separated substances is obtained.

Column quality

Each HPLC / UHPLC column of MACHEREY-NAGEL is individually tested according to the most important characteristic parameters in quality control and the results are documented in a certificate of analysis.

Detailed information of the particular properties of the high-purity silica phases NUCLEODUR®, of the established standard silica NUCLEOSIL® and the modern Core-Shell material NUCLEOSHELL® as well as phases for special separations and the equivalent HPLC- and UHPLC-columns can be found on the following pages.

Strict quality specifications for outstanding reliability

- Highest production standard
  our facilities are EN ISO 9001:2008 certified
- Perfect reproducibility from batch to batch and within each lot
- Each column is individually tested and supplied with test chromatogram and test conditions.

Test mixture* for reversed phase columns
in acetonitrile, pack of 1 mL
REF 722394

MACHEREY-NAGEL
MN
EN ISO 9001:2008
CERTIFIED

Furthermore custom-packed columns with different column types, dimensions and particle sizes are available on request.

* This product (REF 722394) contains harmful substances which must be specially labeled as hazardous. For detailed information please see SDS.

USP listing

USP specification of MN HPLC phases

Code	Specification	MN HPLC Phases	Page
USP L1	octadecyl silane chemically bonded to porous silica particles 1.5 to 10 µm diameter, or monolithic silica gel	NUCLEODUR® C_{18} ec	181
		NUCLEODUR® C_{18} Gravity	158
		NUCLEODUR® C_{18} Gravity-SB	162
		NUCLEODUR® C_{18} HTec	178
		NUCLEODUR® C_{18} Isis	164
		NUCLEODUR® C_{18} PAH	227
		NUCLEODUR® C_{18} Pyramid	166
		NUCLEODUR® PolarTec	168
		NUCLEODUR® Sphinx RP	176
		NUCLEOSHELL® RP 18	200
		NUCLEOSHELL® RP 18plus	202
		NUCLEOSIL® C_{18}	214
		NUCLEOSIL® C_{18} AB	214
		NUCLEOSIL® C_{18} HD	214
		NUCLEOSIL® C_{18} MPN	243
		NUCLEOSIL® C_{18} PAH	229
		NUCLEOSIL® C_{18} PPN	244
USP L3	porous silica particles, 1.5 to 10 µm diameter, or monolithic silica gel	NUCLEODUR® SiOH	190
		NUCLEOSIL® SiOH	224
USP L7	octyl silane chemically bonded to totally porous silica particles, 1.8 to 10 µm diameter	NUCLEODUR® C_8 ec	181
		NUCLEODUR® C_8 Gravity	158
		NUCLEOSIL® C_8	217
		NUCLEOSIL® C_8 HD	217
USP L8	an essentially monomolecular layer of aminopropyl silane chemically bonded to totally porous silica gel support, 1.5 to 10 µm diameter	NUCLEODUR® NH_2 / NH_2-RP	188
		NUCLEOSIL® Carbohydrate	246
		NUCLEOSIL® NH_2 / NH_2-RP	221
USP L9	irregular or spherical, totally porous silica gel having a chemically bonded, strongly acidic cation-exchange coating, 3 to 10 µm diameter	NUCLEOSIL® SA	223
USP L10	nitrile groups chemically bonded to porous silica particles, 1.5 to 10 µm diameter	NUCLEODUR® CN / CN-RP	186
		NUCLEOSIL® CN / CN-RP	222

USP specification of MN HPLC phases

Code	Specification	MN HPLC Phases	Page
USP L11	phenyl groups chemically bonded to porous silica particles, 1.5 to 10 µm diameter	NUCLEODUR® Phenyl-Hexyl	170
		NUCLEODUR® π²	172
		NUCLEOSHELL® Phenyl-Hexyl	204
		NUCLEODUR® Sphinx RP	176
		NUCLEOSIL® C_6H_5	220
USP L14	silica gel having a chemically bonded, strongly basic quaternary ammonium anion-exchange coating, 5 to 10 µm diameter	NUCLEOSIL® SB	223
USP L16	dimethylsilane chemically bonded to porous silica particles, 5 to 10 µm diameter	NUCLEOSIL® C_2	219
USP L17	strong cation-exchange resin consisting of sulfonated cross-linked PS/DVB copolymer in the H form, 6 to 12 µm diameter	NUCLEOGEL® ION 300 OA	248
		NUCLEOGEL® SUGAR 810 H	247
USP L19	strong cation-exchange resin consisting of sulfonated cross-linked PS/DVB copolymer in the Ca form, 5 to 15 µm particle size	NUCLEOGEL® SUGAR 810 Ca	247
		NUCLEOGEL® SUGAR Ca	248
USP L20	dihydroxypropane groups chemically bonded to porous silica particles, 5 to 10 µm diameter	NUCLEOSIL® OH (Diol)	220
USP L21	a rigid, spherical styrene-divinylbenzene copolymer, 5 to 10 µm diameter	NUCLEOGEL® RP	245
USP L22	a cation-exchange resin made of porous polystyrene gel with sulfonic acid groups, about 10 µm in size	NUCLEOGEL® SCX	240
USP L23	an anion-exchange resin made of porous polymethacrylate or polyacrylate gel with quaternary ammonium groups, about 10 µm in size	NUCLEOGEL® SAX	240
USP L26	butyl silane chemically bonded to totally porous silica particles, 5 to 10 µm diameter	NUCLEODUR® C_4 ec	241
		NUCLEOSIL® C_4	219
		NUCLEOSIL® C_4 MPN	243
USP L32	a chiral ligand-exchange resin packing · L-proline copper complex covalently bonded to irregular shaped silica particles, 5 to 10 µm diameter	NUCLEOSIL® CHIRAL-1	235
USP L34	strong cation-exchange resin consisting of sulfonated cross-linked PS-DVB copolymer in the Pb form, 5 to 7 µm particle size	NUCLEOGEL® SUGAR Pb	248
USP L36	a 3,5-dinitrobenzoyl derivative of L-phenylglycine covalently bonded to 5 µm aminopropyl silica	NUCLEOSIL® CHIRAL-3	236
USP L40	cellulose tris-(3,5-dimethylphenylcarbamate) coated porous silica particles, 5 to 20 µm diameter	NUCLEOCEL *DELTA*	233
USP L43	pentafluorophenyl groups chemically bonded to silica particles by a propyl spacer, 1.5 to 10 µm diameter	NUCLEODUR® PFP	174
		NUCLEOSHELL® PFP	206
USP L45	beta-cyclodextrin bonded to porous silica particles, R,S-hydroxypropyl ether derivative, 3 to 10 µm diameter	NUCLEODEX β-OH, β-PM	231
USP L58	strong cation-exchange resin consisting of sulfonated cross-linked PS/DVB copolymer in the Na form, 6 to 30 µm diameter	NUCLEOGEL® SUGAR Na	248
USP L60	spherical porous silica gel, particle size of 10 µm diameter or smaller, the surface of which has been covalently modified with alkyl amide groups and endcapped	NUCLEODUR® PolarTec	168
		NUCLEOSIL® C_{18} Nautilus	214
USP L75	A chiral-recognition protein, bovine serum albumin (BSA), chemically bonded to silica particles, about 7 µm in diameter, with a pore size of 300 Angstrom	RESOLVOSIL BSA-7	234

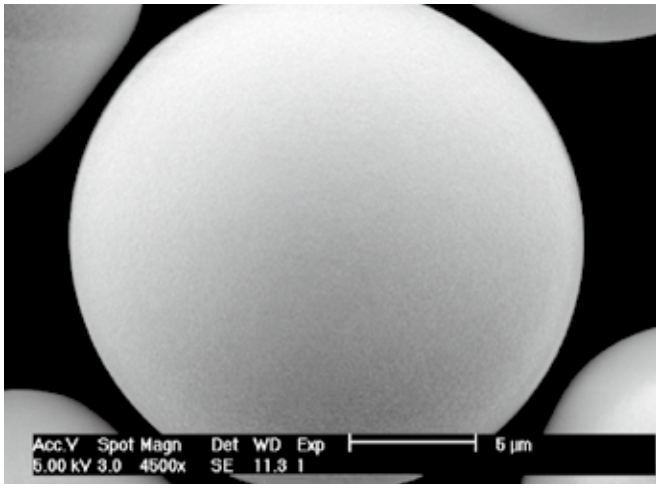
NUCLEODUR® high purity silica for HPLC

NUCLEODUR® is a fully synthetical type B silica (silica of 3rd generation) offering highly advanced physical properties like totally spherical particle shape, outstanding surface microstructure, high pressure stability and low metal content.

NUCLEODUR® as a state-of-the-art silica is the ideal base material for modern HPLC phases. It is the result of MACHEREY-NAGEL's pioneering research in chromatography for more than 40 years.

In RP liquid chromatography the efficiency of the packing is strongly affected by the quality of the base silica itself. Shortcomings in the surface geometry of the particles or metal contaminants are the main reasons for inadequate coverage with the covalently bonded alkylsilanes in the subsequent derivatization steps. It is well known, that poor surface coverage and, in consequence, high activity of residual free silanols often results in peak tailing or adsorption, particularly with basic compounds.

Particle shape and surface symmetry



NUCLEODUR® silicas are synthesized in a unique and carefully controlled manufacturing process which provides silica particles, which are totally spherical. The picture shows the outstanding smoothness of the NUCLEODUR® surface.

Purity

As already mentioned above, a highly pure silica is required for achieving symmetric peak shapes and maximum resolution. Inclusions of, e.g., iron or alkaline earth metal ions on the silica surface are largely responsible for the unwanted interactions with ionizable analytes, e.g., amines or phenolic compounds.

NUCLEODUR® is virtually free of metal impurities and low acidic surface silanols. Elemental analysis data of NUCLEODUR® 5 µm measured by AAS are listed below.

Elementary analysis (metal ions) of NUCLEODUR® 100-5		
Aluminum	< 5	ppm
Iron	< 5	ppm
Sodium	< 5	ppm
Calcium	< 10	ppm
Titanium	< 1	ppm
Zirconium	< 1	ppm
Arsenic	< 0.5	ppm
Mercury	< 0.05	ppm

Pressure stability

The totally spherical and 100 % synthetic silica gel exhibits an outstanding mechanical stability, even at high pressures and elevated eluent flow rates. In addition, after several cycles of repeated packing, no significant drop in pressure can be observed. The latter is of prime importance for preparative and process-scale applications.

NUCLEODUR® silica is available with two pore sizes – 110 Å pore size as standard material and as 300 Å widepore material for the separation of biomolecules, like peptides and proteins.

Physical data of NUCLEODUR®		
	Standard	Widepore
Pore size	110	300 Å
Surface area (BET)	340 m^2/g	100 m^2/g
Pore volume	0.9 mL/g	0.9 mL/g
Density	0.47 g/mL	0.47 g/mL

NUCLEODUR® modifications

Several different surface modifications based on NUCLEODUR® silica have been developed over the last years providing a full range of specified HPLC phases and an ideal tool for every separation.

For a summary of important properties of our NUCLEODUR® phases please see page 152.

1.8 µm particles for increased separation efficiency

Key feature

- Decrease of analysis time (ultra fast HPLC)
- Shorter columns with high separation efficiency and significant improvement of resolution and detection sensitivity
- Suitable for LC/MS due to low bleeding characteristics

Fractionation

- NUCLEODUR® 1.8 µm particles are fractionated to limit the increase in back pressure.

Availability

- The following NUCLEODUR® phases are available in 1.8 µm:
  C_{18} Gravity, C_8 Gravity, C_{18} Gravity-SB, C_{18} Isis, C_{18} Pyramid, PolarTec, Phenyl-Hexyl, PFP, Sphinx RP, C_{18} HTec and HILIC

Advantages of 1.8 µm particle size

Miniaturization started in the early stage of HPLC with the reduction of particle size from 10 µm via 7 µm to standard 5 µm – still the most used particle diameter in analytical HPLC – to 3 µm spherical particles. With the introduction of 1.8 µm NUCLEODUR® particles researchers have turned over a new leaf in HPLC column technology, featuring extraordinary improvements in terms of plate numbers, column efficiency and resolution compared with 3 µm particles.

Increased separation efficiency by higher number of theoretical plates (N):

- 50 x 4.6 mm NUCLEODUR® C_{18} Gravity
- 3 µm: N ≥ 100 000 plates/m (h-value≤ 10)
- 1.8 µm: N ≥ 166 667 plates/m (h-value≤ 6)

Increase of the plate number by ~ 67 % offers the possibility of using shorter columns with equal plate number resulting in a decrease of analysis time.

Significant improvement in resolution

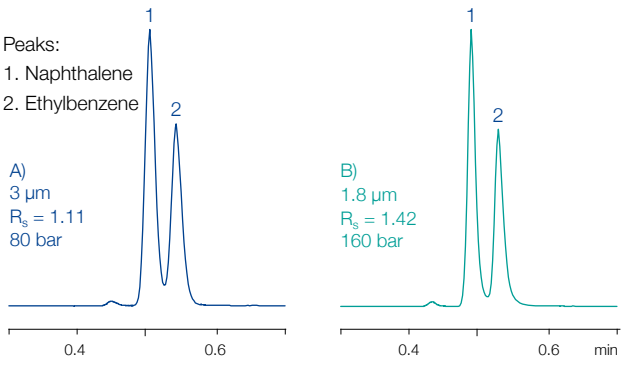
$$R = \frac{\sqrt{N}}{4}\left(\frac{\alpha - 1}{\alpha}\right)\left(\frac{k'_i}{k'_i + 1}\right)$$

R = resolution, α = selectivity (separation factor), k_i' = retention
N = plate number with $N \propto 1/d_P$, d_P = particle diameter

Resolution as a function of particle size

Column: 50 x 4 mm NUCLEODUR® C_{18} Gravity
A) 3 µm, B) 1.8 µm
Eluent: acetonitrile – water (80:20, v/v)
Flow rate: 2 mL/min
Detection: UV, 254 nm

Peaks:
1. Naphthalene
2. Ethylbenzene



Use of 1.8 µm instead of 3 µm particles leads to an increase of resolution by a factor of 1.29 (29 %) since the resolution is inversely proportional to the square root of the particle size.

Column back pressure

Due to the smaller particles the back pressure will increase according to

$$\Delta_P = \frac{\Phi \cdot L_C \cdot \eta \cdot u}{d_P^2}$$

Δ_P = pressure drop, Φ = flow resistance (nondimensional), LC = column length, η = viscosity, u = linear velocity, d_P = particle diameter

The high sphericity of the NUCLEODUR® particles and the very narrow particle size distribution allow to keep the back pressure on a moderate level.

Comparison of back pressures

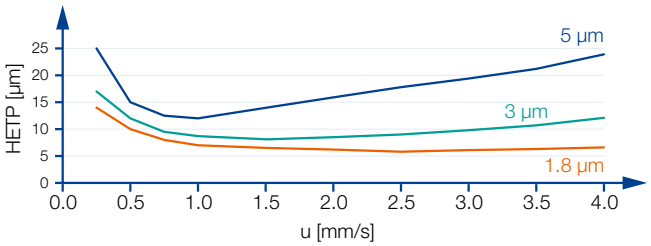
Eluent 100 % methanol, flow rate 1.5 mL/min
temperature 22 °C, column dimensions 50 x 4.6 mm

	NUCLEODUR® C_{18} Gravity	Competitor
3 µm	70 bar	–
1.8 µm	130 bar	170 bar

Higher flow rates and shorter run times

The optimal flow rate for 1.8 µm particles is higher than for 3 and 5 µm particles (see figure – the flow rate should be at the van Deemter minimum).

Van Deemter curves



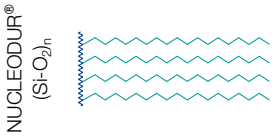
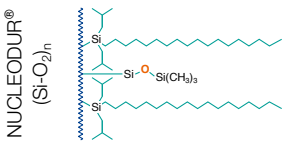
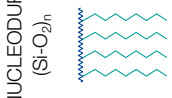
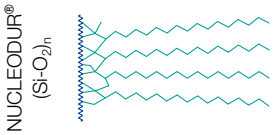
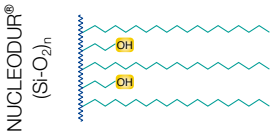
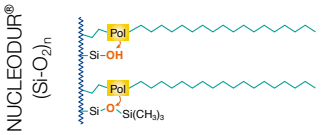
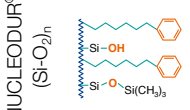
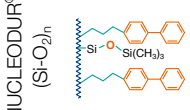
Column 50 x 4.6 mm, acetonitrile – water (50:50, v/v), analyte toluene

Technical requirements

To gain best results with 1.8 µm particles certain technical demands must be met including pumps for flow rates of 2–3 mL with pressures of 250–1000 bar, minimized dead volume, and fast data recording.

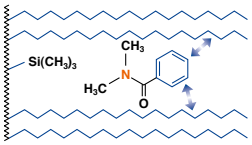
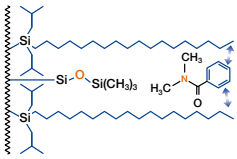
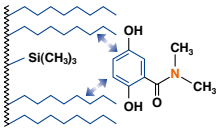
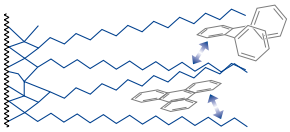
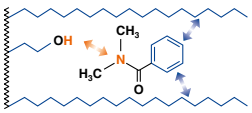
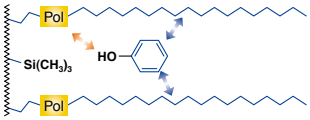
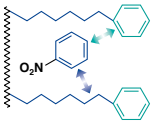
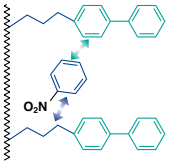
NUCLEODUR® phase overview

Overview of NUCLEODUR® HPLC phases

Phase	Specification	Page	Characteristic*	Stability	Structure
C_{18} Gravity	octadecyl, high density coating, multi-endcapping 18 % C · USP L1	158	A ●●●●● B ◖ C ●●◖	pH 1–11, suitable for LC/MS	
C_{18} Gravity-SB	octadecyl (monomeric), extensive endcapping 13 % C · USP L1	162	A ●●●● B ●●◖ C -	pH 1–9, suitable for LC/MS	
C_8 Gravity	octyl, high density coating, multi-endcapping 11 % C · USP L7	158	A ●●◖ B ● C ●◖	pH 1–11, suitable for LC/MS	
C_{18} Isis	octadecyl phase with specially crosslinked surface modification endcapping 20 % C · USP L1	164	A ●●●●◖ B ●● C ●●●●●	pH 1–10, suitable for LC/MS	
C_{18} Pyramid	octadecyl with polar endcapping 14 % C · USP L1	166	A ●●●● B ●●◖ C ●●	stable in 100 % aqueous eluent, pH 1–9, suitable for LC/MS	
PolarTec	octadecyl with embedded polar group 17 % C · USP L1 and L60	168	A ●●●● B ●●◖ C ●●●●	stable in 100 % aqueous eluent, pH 1–9, suitable for LC/MS	
Phenyl-Hexyl	phenylhexyl, multi-endcapping 10 % C · USP L11	170	A ●● B ●●● C ●	pH 1–10, suitable for LC/MS	
π^2	biphenylpropyl, multi-endcapping 17 % C · USP L11	172	A ●● B ●●●● C ●●●	pH 1.5–10	

* A = ● hydrophobic selectivity, B = ● polar / ionic selectivity, C = ● steric selectivity

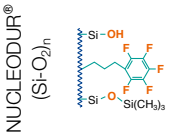
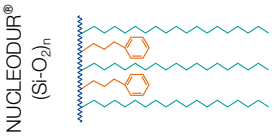
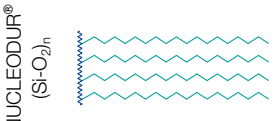
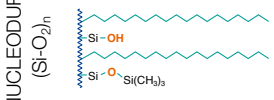
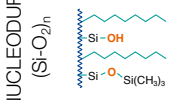
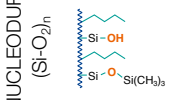
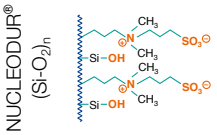
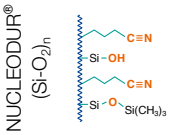
NUCLEODUR® phase overview

Application	Similar phases**	Interactions · retention mechanism	
in general compounds with ionizable functional groups such as basic pharmaceuticals and pesticides	NUCLEOSIL® C_{18} HD Xterra® RP18 / MS C18; Luna® C18(2), Gemini®, Synergi® Max RP; Zorbax® Extend-C18; Inertsil® ODS III; Purospher® STAR RP-18; Hypersil™ BDS	hydrophobic (van der Waals interactions)	
overall sophisticated analytical separations, especially for polar compounds, e.g., antibiotics, water-soluble vitamins, organic acids	–	hydrophobic (van der Waals interactions) with additional polar interactions	
like C_{18} Gravity, however, generally shorter retention times for nonpolar compounds	NUCLEOSIL® C_8 HD Xterra® RP8 / MS C8; Luna® C8; Zorbax® Eclipse XDB-C8	hydrophobic (van der Waals interactions)	
high steric selectivity, thus suited for separation of positional and structural isomers, planar / nonplanar molecules	NUCLEOSIL® C_{18} AB Inertsil® ODS-P; Pro C18 RS	steric and hydrophobic	
basic pharmaceuticals, very polar compounds, organic acids	Aqua, Synergi® Hydro-RP; AQ; Atlantis® dC18; Polaris® C18-A	hydrophobic and polar (H bonds)	
basic pharmaceuticals, organic acids, pesticides, amino acids, water-soluble vitamins	NUCLEOSIL® C_{18} Nautilus ProntoSIL® C18 AQ, Zorbax® Bonus-RP, Polaris® Amide-C18; Ascentis® RP Amide, SymmetryShield™ RP18; SUPELCOSIL™ LC-ABZ+; HyPURITY™ ADVANCE; ACCLAIM Polar AD.II	hydrophobic and polar (H bonds)	
aromatic and unsaturated compounds, polar compounds like pharmaceuticals, antibiotics	Luna® Phenyl-Hexyl; Zorbax® Eclipse Plus Phenyl-Hexyl; Kromasil® Phenyl-Hexyl	π-π and hydrophobic	
aromatic and unsaturated compounds, polar compounds like pharmaceuticals, antibiotics	Pinnacle® DB Biphenyl; Ultra Biphenyl	π-π and hydrophobic	

** phases which provide a similar selectivity based on chemical and physical properties

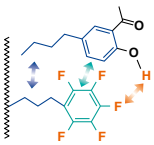
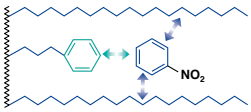
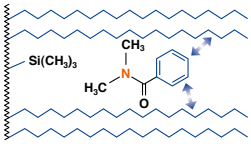
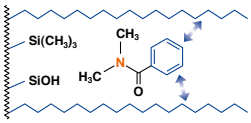
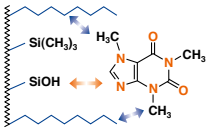
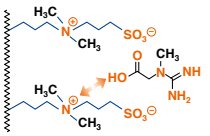
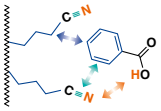
NUCLEODUR® phase overview

Overview of NUCLEODUR® HPLC phases

Phase	Specification	Page	Characteristic*	Stability	Structure
PFP	pentafluorophenylpropyl, multi-endcapping 8 % C · USP L43	174	A ●● B ●●●● C ●●●●	pH 1–9, suitable for LC/MS	
Sphinx RP	bifunctional, balanced ratio of propylphenyl and octadecyl, endcapping 15 % C · USP L1 and L11	176	A ●●● B ●●◖ C ●	pH 1–10, suitable for LC/MS	
C_{18} HTec	octadecyl, high density coating, high capacity, multi-endcapping 18 % C · USP L1	178	A ●●●●● B ◖ C ●●◖	pH 1–11, suitable for LC/MS	
C_{18} ec	octadecyl, medium density, endcapping available in 110 Å and 300 Å pore size 17.5 % / 4 % C · USP L1	181	A ●●●● B ● C ●●●◖	pH 1–9	
C_8 ec	octyl, medium density, endcapping 10.5 % C · USP L7	181	A ●● B ●◖ C ●●◖	pH 1–9	
C_4 ec	butyl, medium density, endcapping, 300 Å pore size 2.5 % C · USP L26	181	A ● B ●● C ●◖	pH 1–9	
HILIC	zwitterionic ammonium – sulfonic acid phase 7 % C	184	A ● B ●●●●● C -	pH 2–8.5	
CN / CN-RP	cyano (nitrile) for NP and RP separations 7 % C · USP L10	186	A ● B ●●●● C -	pH 1–8, stable towards highly aqueous mobile phases	

* A = ● hydrophobic selectivity, B = ● polar / ionic selectivity, C = ● steric selectivity

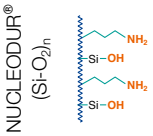
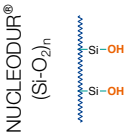
NUCLEODUR® phase overview

Application	Similar phases**	Interactions · retention mechanism	
aromatic and unsaturated compounds, halogen compounds, phenols, isomers, polar pharmaceuticals, antibiotics	ACQUITY® CSH Fluoro-Phenyl; Hypersil™ GOLD PFP; Luna® PFP(2); Discovery® HS F5; Allure® PFP Propyl; Ultra II PFP Propyl	polar (H bond), dipole-dipole, π-π and hydrophobic	
compounds with aromatic and multiple bond systems	no similar phases	π-π and hydrophobic	
robust and well base deactivated C_{18} phase; all separation tasks with preparative potential	Xterra® RP18 / MS C18 / SunFire™ C18; Luna® C18(2), Gemini®, Synergi® Max RP; Zorbax® Extend-C18; Inertsil® ODS III; Purospher® STAR RP-18; Hypersil® BDS	hydrophobic (van der Waals interactions)	
robust C_{18} phase for routine analyses	NUCLEOSIL® C_{18} Spherisorb® ODS II; Symmetry® C18; Hypersil® ODS; Inertsil® ODS II; Kromasil® C18; LiChrospher® RP-18	hydrophobic (van der Waals interactions) some residual silanol interactions	
robust C_8 phase for routine analyses	NUCLEOSIL® C_8 ec / C_8 Spherisorb® C8; Symmetry® C8; Hypersil® MOS; Kromasil® C8; LiChrospher® RP-8	hydrophobic (van der Waals interactions) some residual silanol interactions	
biological macromolecules like proteins or peptides	Jupiter® C4; ACE® C4	hydrophobic (van der Waals interactions) some residual silanol interactions	
hydrophilic compounds such as polar organic acids and bases, polar natural compounds	Sequant™ ZIC®-HILIC; Obelisc™	ionic / hydrophilic and electrostatic	
polar organic compounds (basic drugs), molecules containing π-electron systems	NUCLEOSIL® CN / CN-RP	π-π and polar (H bond), hydrophobic	

** phases which provide a similar selectivity based on chemical and physical properties

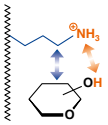
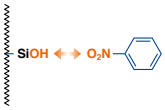
NUCLEODUR® phase overview

Overview of NUCLEODUR® HPLC phases

Phase	Specification	Page	Characteristic*	Stability	Structure
NH_2 / NH_2-RP	aminopropyl for NP and RP separations 2.5 % C · USP L8	188	A ● B ●●●● C -	pH 2–8, stable towards highly aqueous mobile phases	
SiOH	unmodified high purity silica · USP L3	190	A - B - C -	pH 2–8	

* A = ● hydrophobic selectivity, B = ● polar / ionic selectivity, C = ● steric selectivity

NUCLEODUR® phase overview

Application	Similar phases**	Interactions · retention mechanism	
sugars, sugar alcohols and other hydroxy compounds, DNA bases, polar compounds in general	NUCLEOSIL® NH_2 / NH_2-RP	polar / ionic and hydrophobic	
polar compounds in general	NUCLEOSIL® SiOH	polar / ionic	

** phases which provide a similar selectivity based on chemical and physical properties

NUCLEODUR® C_{18} Gravity · C_8 Gravity nonpolar high density phase · USP L1 (C_{18}) · USP L7 (C_8)

Key feature

- Suitable for LC/MS and HPLC at pH extremes (pH 1–11)
- Superior base deactivation
- Ideal for method development

Technical data

- Available as octadecyl (C_{18}) and octyl (C_8), multi-endcapped
- Pore size 110 Å; particle sizes 1.8 µm, 3 µm and 5 µm for C_{18}, 1.8 and 5 µm for C_8; 7, 10, 12 and 16 µm particles for preparative purposes on request
- Carbon content 18 % for C_{18}, 11 % for C_8

Recommended application

- Overall sophisticated analytical separations
- Compound classes separated include pharmaceuticals, e.g., analgesics, anti-inflammatory drugs, antidepressants; herbicides; phytopharmaceuticals; immunosuppressants

Base deactivation

NUCLEODUR® C_{18} Gravity and NUCLEODUR® C_8 Gravity are based on the ultrapure NUCLEODUR® silica. Derivatization generates a homogeneous surface with a high density of bonded silanes (~18 % C for C_{18}, ~11 % C for C_8). Thorough endcapping suppresses any unwanted polar interactions between the silica surface and the sample, which makes “Gravity” particularly suitable for the separation of basic and other ionizable analytes. Even strongly basic pharmaceuticals like amitriptyline are eluted without tailing under isocratic conditions. For a discussion of the different retention behavior of C_{18} phases compared to C_8 phases see page 182.

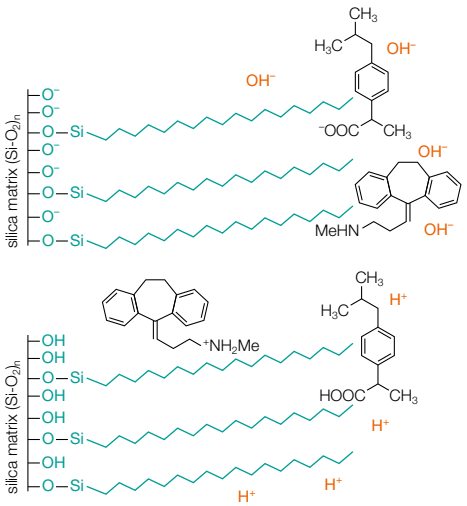
Enhanced pH stability

One major disadvantage of silica stationary phases is limited stability at strongly acidic or basic pH. Cleavage of the siloxane bonding by hydrolysis, or dissolution of the silica will rapidly lead to a considerable loss in column performance. Conventional RP phases are usually not recommended to be run with mobile phases at pH > 8 or pH < 2 for extended periods of time. The special surface bonding technology and the low concentration of trace elements of NUCLEODUR® C_{18} and C_8 Gravity allow for use at an expanded pH range from pH 1 to 11.

Benefits of enhanced pH stability

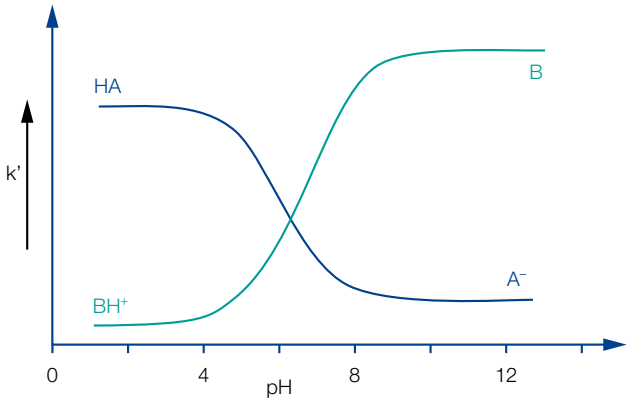
An expanded pH range is often required in method development. Many nitrogen containing compounds like basic drugs are protonated at acidic or neutral pH and exhibit poor retention on a standard C_{18} phase. The retention behavior can be improved by working at a higher pH, where the analyte is no longer protonated, but formally neutrally charged, as a rule between pH 9–10. For acidic analytes it is exactly in inverse proportion, maximum retention can be attained at low pH.

Surface silanols at different pH values



The figure above shows the extent of protonation of surface silanols and of two exemplary analytes at acidic and alkaline pH. The following graph explains the general correlation between retention and pH.

Correlation between retention and pH for basic and acidic compounds



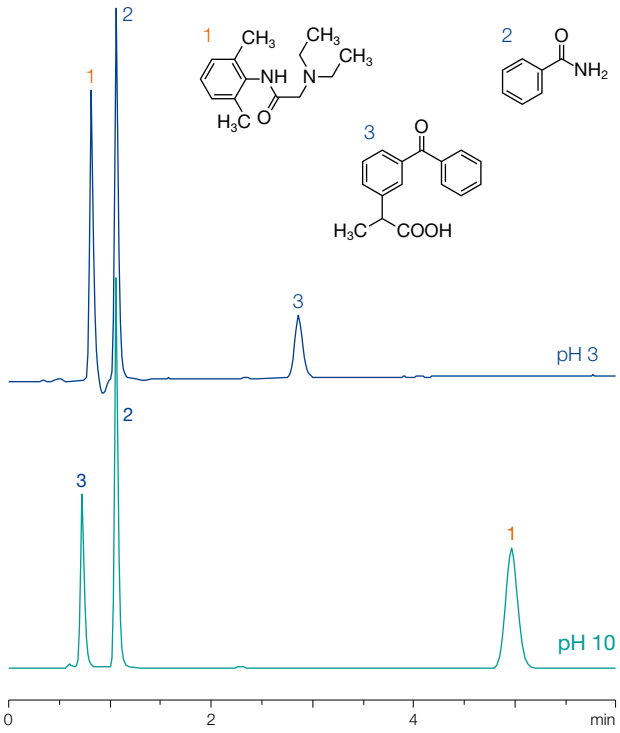
An example how selectivity can be controlled by pH is the separation of the acid ketoprofen, the base lidocaine and benzamide. Under acidic conditions the protonated lidocaine is eluted very fast due to lack of sufficiently strong hydrophobic interactions between analyte and C_{18} chains, while the formally neutral ketoprofen is eluted after about 3 min. However, at pH 10 a reversal of the elution order, with a visibly longer retention time for the basic lidocaine, is observed.

Influence of the pH value on selectivity

MN Appl. No. 120860

Column:	125 x 4 mm NUCLEODUR® C_{18} Gravity, 5 µm
Eluent:	A) acetonitrile – 10 mmol/L ammonium formate, pH 3.0 (50:50, v/v); B) acetonitrile – 10 mmol/L ammonium bicarbonate, pH 10.0 (50:50, v/v)
Flow rate:	1.0 mL/min
Temperature:	30 °C
Detection:	UV, 230 nm
Injection:	2 µL

Peaks:
1. Lidocaine
2. Benzamide
3. Ketoprofen



As mentioned above, pH stability of the stationary phase can be helpful for improving selectivity in method development. The following figure shows the separation of 4 basic drugs under acidic and basic conditions.

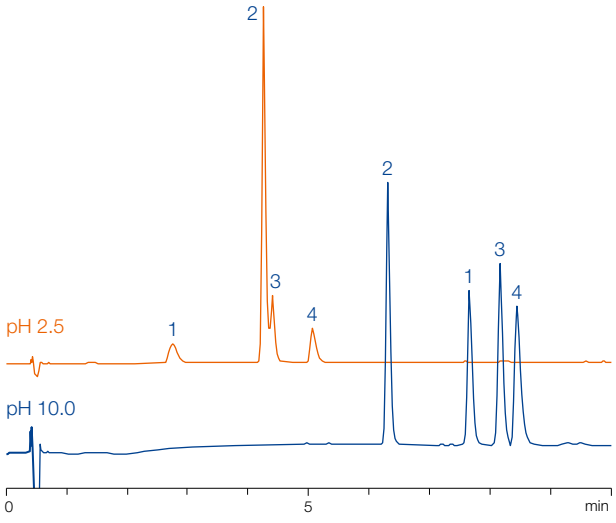
At pH 2.5 the protonated analytes exhibit poor retention (early elution) and in addition an inadequate resolution for papaverine and noscapine, whilst the formally non ionized molecules can be baseline separated due to the better retention pattern at alkaline pH.

Separation of basic alkaloids

MN Appl. No. 118010

Column:	125 x 4 mm NUCLEODUR® C_{18} Gravity, 5 µm
Eluent:	A) acetonitrile B) 20 mmol/L $(NH_4)_2HPO_4$, pH 2.5 / 10.0 10 % A (1 min) → 75 % A in 10 min
Flow rate:	1.0 mL/min; Temperature 25 °C
Detection:	UV, 254 nm; Injection 2 µL

Peaks:
1. Lidocaine
2. Papaverine
3. Noscapine
4. Diphenhydramine



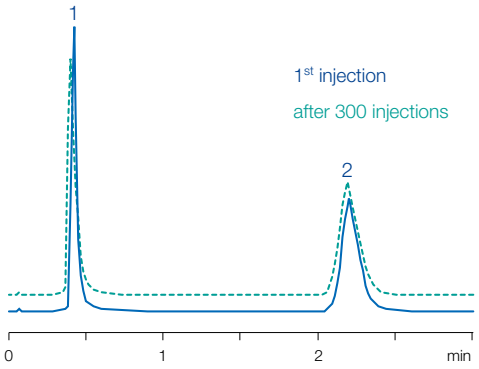
The following chromatogram demonstrates the stability of NUCLEODUR® C_{18} Gravity under alkaline conditions. The ultrapure Gravity with its unique high density surface bonding technology withstands strong alkaline mobile phase conditions.

Stability of NUCLEODUR® C_{18} Gravity at pH 11

MN Appl. No. 120850

Column:	50 x 4.6 mm NUCLEODUR® C_{18} Gravity, 5 µm
Eluent:	methanol – water – ammonia (20:80:0.5, v/v/v), pH 11
Flow rate:	1.3 mL/min
Temperature:	30 °C
Detection:	UV, 254 nm
Injection:	2.0 µL

Peaks:
1. Theophylline
2. Caffeine



Even after 300 injections no loss of column efficiency – identified, e.g., by peak broadening or decrease in retention times – could be observed.

Under alkaline conditions dissolution of the silica support is possible, resulting in dead volume and thus peak broadening. It is worth mentioning, that this phenomenon also depends on type and concentration of buffers, as well as on the temperature. It is well known that the use of phosphate buffers, particularly at elevated temperatures, can reduce column lifetime even at moderate pH. If possible, phosphate buffers should be replaced by less harmful alternatives.

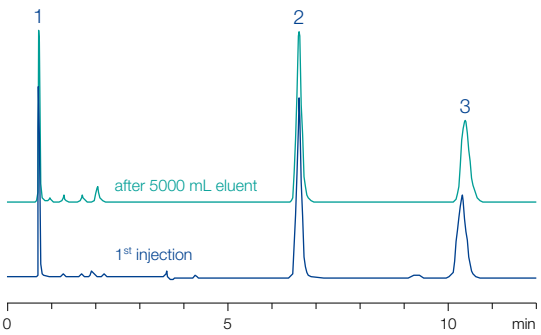
The following chromatograms show the excellent column stability of NUCLEODUR® C_{18} Gravity in acidic conditions. Retention times of all three compounds in the column performance test remain consistent and virtually unchanged, even after the column is run with 5000 mL eluent. Due to the extremely stable surface modification, no cleavage of the Si-O-Si bonding occurs, column deterioration is therefore successfully prevented.

Stability of NUCLEODUR® C_{18} Gravity at pH 1.5

MN Appl. No. 120840

Column:	125 x 4 mm NUCLEODUR® C_{18} Gravity, 5 µm
Eluent:	acetonitrile – 1 % TFA in water (50:50, v/v), pH 1.5
Flow rate:	1.0 mL/min
Temperature:	30 °C
Detection:	UV, 230 nm
Injection:	5 µL

Peaks: 1. Pyridine, 2. Toluene, 3. Ethylbenzene



Ordering information

Eluent in column acetonitrile – water

	ID	Length → 30 mm	50 mm	75 mm	100 mm	125 mm	150 mm	250 mm
NUCLEODUR® C_{18} Gravity, 1.8 µm octadecyl phase, particle size 1.8 µm, 18 % C · UHPLC								
Analytical EC columns								
	2 mm	760078.20	760079.20	760071.20	760076.20		760075.20	
	3 mm	760078.30	760079.30		760076.30			
	4 mm	760078.40	760079.40		760076.40			
	4.6 mm	760078.46	760079.46		760076.46			
EC guard columns*			4 x 2 mm: 761901.20		4 x 3 mm: 761901.30			
NUCLEODUR® C_{18} Gravity, 3 µm octadecyl phase, particle size 3 µm, 18 % C								
Analytical EC columns								
	2 mm		760080.20		760084.20	760081.20	760083.20	760082.20
	3 mm		760080.30		760084.30	760081.30	760083.30	760082.30
	4 mm		760080.40		760084.40	760081.40	760083.40	760082.40
	4.6 mm		760080.46	760086.46	760084.46	760081.46	760083.46	760082.46
EC guard columns*			4 x 2 mm: 761902.20		4 x 3 mm: 761902.30			

Ordering information

Eluent in column acetonitrile – water

ID	Length → 30 mm	50 mm	75 mm	100 mm	125 mm	150 mm	250 mm
NUCLEODUR® C_{18} Gravity, 5 µm octadecyl phase, particle size 5 µm, 18 % C							
Analytical EC columns							
2 mm		760102.20		760104.20	760100.20	760103.20	760101.20
3 mm		760102.30		760104.30	760100.30	760103.30	760101.30
4 mm		760102.40		760104.40	760100.40	760103.40	760101.40
4.6 mm		760102.46	760106.46	760104.46	760100.46	760103.46	760101.46
EC guard columns*		4 x 2 mm: 761903.20		4 x 3 mm: 761903.30			
Preparative VarioPrep columns							
10 mm		762103.100			762109.100		762113.100
21 mm		762103.210			762109.210		762113.210
32 mm							762113.320
40 mm						762100.400	762113.400
VP guard columns ***		10 x 8 mm: 762160.80		10 x 16 mm: 762160.160		15 x 32 mm: 762163.320	
NUCLEODUR® C_{18} Gravity, 10 µm octadecyl phase, particle size 10 µm, 18 % C							
Preparative VarioPrep columns							
21 mm							762250.210
40 mm							762250.400
VP guard columns **				10 x 16 mm: 762160.160		15 x 32 mm: 762163.320	

Ordering information

Eluent in column acetonitrile – water

ID	Length → 30 mm	50 mm	75 mm	100 mm	125 mm	150 mm	250 mm
NUCLEODUR® C_8 Gravity, 1.8 µm octyl phase, particle size 1.8 µm, 11 % C · UHPLC							
Analytical EC columns							
2 mm	760756.20	760755.20	760760.20	760757.20		760759.20	
3 mm	760756.30	760755.30		760757.30			
4 mm	760756.40	760755.40		760757.40			
4.6 mm	760756.46	760755.46		760757.46			
EC guard columns*		4 x 2 mm: 761905.20		4 x 3 mm: 761905.30			
NUCLEODUR® C_8 Gravity, 5 µm octyl phase, particle size 5 µm, 11 % C							
Analytical EC columns							
2 mm		760750.20		760754.20	760751.20	760752.20	760753.20
3 mm		760750.30		760754.30	760751.30	760752.30	760753.30
4 mm		760750.40		760754.40	760751.40	760752.40	760753.40
4.6 mm		760750.46	760749.46	760754.46	760751.46	760752.46	760753.46
EC guard columns*		4 x 2 mm: 761907.20		4 x 3 mm: 761907.30			
Preparative VarioPrep columns							
10 mm		762081.100			762071.100		762070.100
21 mm		762081.210			762071.210	762082.210	762070.210
VP guard columns **		10 x 8 mm: 762097.80		10 x 16 mm: 762097.160			

EC and VarioPrep columns in packs of 1, guard columns see below.

Guard column systems

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966
Guard columns for VarioPrep columns with ID		**8, 10 mm**	**16, 21 mm**	**32, 40 mm**	**≥ 50 mm**	
** VP guard columns (pack of)	VP	10/8 (2)	10/16 (2)	15/32 (1)	15/50 (1)	
VP guard column holder		718251	718256	718253	718255	

For details of our column systems see page 250.

NUCLEODUR® C_{18} Gravity-SB hydrophobic phase with polar selectivity · USP L1

Key feature

- Hydrophobic C_{18} phase with distinct polar selectivity, ideal for method development, better retention of early eluting substances
- Excellent performance under highly aqueous conditions
- Suitable for LC/MS due to low bleeding characteristics

Technical data

- Monomeric octadecyl modification, extensive endcapping
- Pore size 110 Å; available particle sizes 1.8 µm, 3 µm and 5 µm; carbon content 13 %; pH stability 1–9

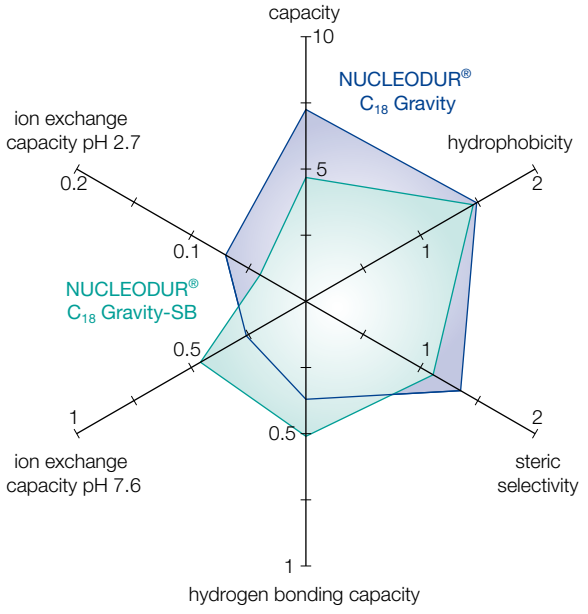
Recommended application

- Overall sophisticated analytical separations, especially for polar compounds, e.g., antibiotics, water-soluble vitamins, organic acids

NUCLEODUR® C_{18} Gravity-SB excels with a relatively high hydrophobicity – similar to C_{18} Gravity – while simultaneously showing distinctive polar selectivity, without having polar embedded groups or polar endcapping. As a result the column displays better retention of early eluting analytes and high performance under strongly aqueous conditions. Additionally the column is suitable for LC/MS due to low bleeding characteristics. These features are achieved through side chains (isobutyl) of the monomeric C_{18} phase.

In the TANAKA plot the NUCLEODUR® Gravity-SB shows similar hydrophobicity than the Gravity, however with a reduced capacity. The ion exchange capacity under basic conditions (pH 7.6) is high, which favors good retention of early eluting, polar substances.

Due to the broad selectivity and stability the base deactivated NUCLEODUR® C_{18} Gravity-SB is versatile applicable, especially for polar analytes like nucleobases or pesticides the column shows good separation efficiency.



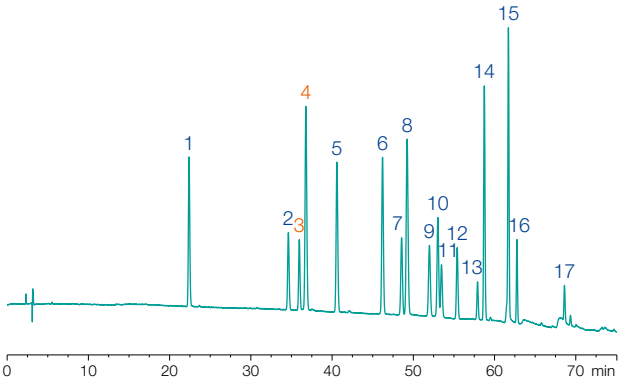
Pesticide mix (Ehrenstorfer, 17 components)

MN Appl. No. 127330

Column:	EC 250/4.6 NUCLEODUR® C_{18} Gravity-SB, 3 µm
Eluent:	A) acetonitrile
	B) 5 mmol/L NH_4Ac;
	10–37.5 % A in 50 min, 37.5–75 % A in 25 min
Flow rate:	1.1 mL/min
Temperature:	35 °C
Detection:	UV, 230 nm
Injection:	3 µL

Peaks:

1. Desethylatrazine	7. Chlortoluron	13. Metazachlor
2. Metoxuron	8. Atrazine	14. Sebuthylazin
3. Hexazinone	9. Monolinuron	15. Terbuthylazine
4. Simazine	10. Isoproturon	16. Linuron
5. Cyanazine	11. Diuron	17. Metolachlor
6. Methabenzthiazuron	12. Metobromuron	



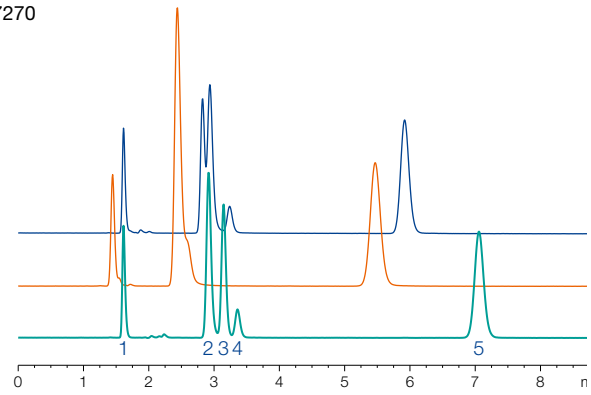
Good separation of the critical pair hexazinone/simazine

Comparing of selectivity for nucleobases

MN Appl. No. 127270

Columns: EC 150/4.6 mm
NUCLEODUR® C_{18} Gravity-SB, 5 µm
NUCLEODUR® C_{18} Gravity, 5 µm
NUCLEODUR® C_{18} Pyramid, 5 µm
Eluent: 25 mmol/L KH_2PO_4, pH 3 – methanol (95:5, v/v)
Flow rate: 1.0 mL/min, Temperature: 20 °C
Detection: UV, 220 nm, Injection: 2.5 µL (1 mg/mL)

Peaks:
1. Cytosine
2. Adenine
3. Uracil
4. Guanine
5. Thymine



Better resolution of early eluting analyte

Ordering information

Eluent in column acetonitrile – water

ID	Length → 30 mm	50 mm	75 mm	100 mm	125 mm	150 mm	250 mm
NUCLEODUR® C_{18} Gravity-SB, 1.8 µm particle size 1.8 µm · UHPLC							
Analytical EC columns							
2 mm	760591.20	760593.20	760595.20	760596.20		760598.20	
3 mm	760591.30	760593.30		760596.30			
4 mm	760591.40	760593.40		760596.40			
4.6 mm	760591.46	760593.46		760596.46			
EC guard columns*		4 x 2 mm: 761990.20		4 x 3 mm: 761990.30			
NUCLEODUR® C_{18} Gravity-SB, 3 µm particle size 3 µm							
Analytical EC columns							
2 mm		760603.20		760606.20	760607.20	760608.20	760609.20
3 mm		760603.30		760606.30	760607.30	760608.30	760609.30
4 mm		760603.40		760606.40	760607.40	760608.40	760609.40
4.6 mm		760603.46	760605.46	760606.46	760607.46	760608.46	760609.46
EC guard columns*		4 x 2 mm: 761991.20		4 x 3 mm: 761991.30			
NUCLEODUR® C_{18} Gravity-SB, 5 µm particle size 5 µm							
Analytical EC columns							
2 mm		760613.20		760616.20	760617.20	760618.20	760619.20
3 mm		760613.30		760616.30	760617.30	760618.30	760619.30
4 mm		760613.40		760616.40	760617.40	760618.40	760619.40
4.6 mm		760613.46	760615.46	760616.46	760617.46	760618.46	760619.46
EC guard columns*		4 x 2 mm: 761992.20		4 x 3 mm: 761992.30			
Preparative VarioPrep columns							
10 mm		762350.100			762351.100		762353.100
21 mm		762350.210			762351.210		762353.210
32 mm							762353.320
40 mm						762352.400	762353.400
VP guard columns **		10 x 8 mm: 762354.80		10 x 16 mm: 762354.160		15 x 32 mm: 762355.320	

EC and VarioPrep columns in packs of 1, guard columns see below.

Guard column systems

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column hold
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966
Guard columns for VarioPrep columns with ID		**8, 10 mm**	**16, 21 mm**	**32, 40 mm**	**≥ 50 mm**	
** VP guard columns (pack of)	VP	10/8 (2)	10/16 (2)	15/32 (1)	15/50 (1)	
VP guard column holder		718251	718256	718253	718255	

For details of our column systems see page 250.

NUCLEODUR® columns

NUCLEODUR® C_{18} Isis phase with high steric selectivity · USP L1

Key feature

- Exceptional steric selectivity
- Outstanding surface deactivation
- Suitable for LC/MS and HPLC at pH 1–10

Technical data

- C_{18} phase with special polymeric, crosslinked surface modification; pore size 110 Å; particle sizes 1.8 µm, 3 µm and 5 µm; carbon content 20 %

Recommended application

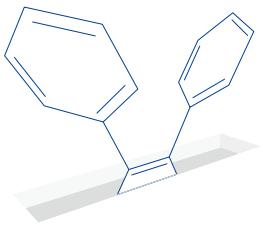
- Steroids, (*o*,*p*,*m*-)substituted aromatics, fat-soluble vitamins

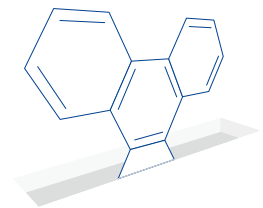
Surface modification

By use of specific C_{18} silanes and polymeric bonding technologies a dense shield of alkyl chains protects the subjacent silica matrix. Elemental analysis of NUCLEODUR® C_{18} Isis shows a carbon load of 20 %. The target crosslinking of the C_{18} chains on the surface enables the separation of compounds with similar molecular structure but different stereochemical properties. The technical term for this feature is steric selectivity.

Slot Model

Sander and Wise [5] proposed a model for the retention of aromatic compounds based on molecular shape, which is referred to as "Slot Model". This model pictures the bonded C_{18} phase on the silica surface with slots which the analytes have to penetrate during retention. Planar molecules are able to penetrate these slots deeper than non-planar molecules of similar molecular weight and length-to-width ratio. Thus triphenylene (lower structure) is longer retained than o-terphenyl (upper structure).





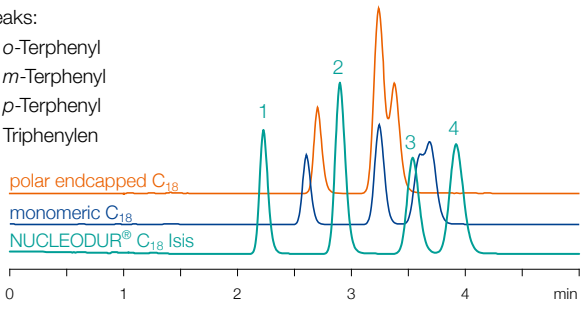
Steric selectivity

The following chromatograms reveal the improved resolution for positional isomers in a test mixture of aromatic compounds on NUCLEODUR® C_{18} Isis (green) in direct comparison with monomerically coated (blue) and polar endcapped (orange) C_{18} columns.

Steric selectivity of NUCLEODUR® C_{18} Isis

Columns: 125 x 4 mm
NUCLEODUR® C_{18} Isis
monomerically coated C_{18} phase
polar endcapped phase C_{18} phase
Eluent: methanol – water (90:10, v/v)
Flow rate: 1 mL/min, temperature: 35 °C
Detection: UV, 254 nm
Injection: 5 µL

Peaks:
1. *o*-Terphenyl
2. *m*-Terphenyl
3. *p*-Terphenyl
4. Triphenylen

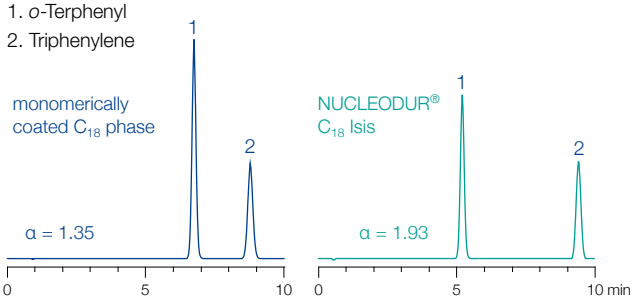


The separation of o-terphenyl and triphenylene is a good example to evaluate the selectivity of a RP column in terms of the shape of two molecules. The phenyl rings of o-terphenyl are twisted out of plane while triphenylene has a planar geometry. The separation factor α is a measure for the steric selectivity. As is shown below the α value is considerable larger on NUCLEODUR® C_{18} Isis compared to a conventional C_{18} column.

Steric selectivity of NUCLEODUR® C_{18} Isis

Columns: 125 x 4 mm
Eluent: methanol – water (80:20, v/v)
Flow rate: 1 mL/min
Temperature: 40 °C
Detection: UV, 254 nm
Injection: 1 µL

Peaks:
1. *o*-Terphenyl
2. Triphenylene



The surface bonding technology also provides improved stability features for the NUCLEODUR® C_{18} Isis phase.

Surface deactivation

The chromatography of basic analytes requires a high density of surface-bonded C_{18} silanes combined with a thorough endcapping procedure to keep silanol activity at a minimum. This ensures tailing-free elution of even strongly basic amino-containing compounds (see application 121210 at *www.mn-net.com/apps*).

Ordering information

Eluent in column acetonitrile – water

ID	Length → 30 mm	50 mm	75 mm	100 mm	125 mm	150 mm	250 mm
NUCLEODUR® C_{18} Isis, 1.8 µm particle size 1.8 µm · UHPLC							
Analytical EC columns							
2 mm	760406.20	760405.20	760396.20	760407.20		760409.20	
3 mm	760406.30	760405.30		760407.30			
4 mm	760406.40	760405.40		760407.40			
4.6 mm	760406.46	760405.46		760407.46			
EC guard columns*		4 x 2 mm: 761910.20		4 x 3 mm: 761910.30			
NUCLEODUR® C_{18} Isis, 3 µm particle size 3 µm							
Analytical EC columns							
2 mm		760400.20		760401.20	760402.20	760403.20	760404.20
3 mm		760400.30		760401.30	760402.30	760403.30	760404.30
4 mm		760400.40		760401.40	760402.40	760403.40	760404.40
4.6 mm		760400.46	760397.46	760401.46	760402.46	760403.46	760404.46
EC guard columns*		4 x 2 mm: 761911.20		4 x 3 mm: 761911.30			
NUCLEODUR® C_{18} Isis, 5 µm particle size 5 µm							
Analytical EC columns							
2 mm		760410.20		760415.20	760412.20	760413.20	760414.20
3 mm		760410.30		760415.30	760412.30	760413.30	760414.30
4 mm		760410.40		760415.40	760412.40	760413.40	760414.40
4.6 mm		760410.46	760416.46	760415.46	760412.46	760413.46	760414.46
EC guard columns*		4 x 2 mm: 761912.20		4 x 3 mm: 761912.30			
Preparative VarioPrep columns							
10 mm		762404.100			762405.100		762403.100
21 mm		762404.210			762405.210		762403.210
32 mm							762403.320
40 mm						762406.400	762403.400
VP guard columns **		10 x 8 mm: 762420.80		10 x 16 mm: 762420.160		15 x 32 mm: 762422.320	
EC and VarioPrep columns in packs of 1, guard columns see below.							

Guard column systems

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966
Guard columns for VarioPrep columns with ID		**8, 10 mm**	**16, 21 mm**	**32, 40 mm**	**≥ 50 mm**	
** VP guard columns (pack of)	VP	10/8 (2)	10/16 (2)	15/32 (1)	15/50 (1)	
VP guard column holder		718251	718256	718253	718255	

For details of our column systems see page 250.

NUCLEODUR® C_{18} Pyramid phase for highly aqueous eluents · USP L1

Key feature

- Stable in 100 % aqueous mobile phase systems
- Interesting polar selectivity features
- Excellent base deactivation; suitable for LC/MS due to low bleeding characteristics

Technical data

- Special phase with polar endcapping; pore size 110 Å; particle sizes 1.8 µm, 3 µm and 5 µm (7 and 10 µm particles for preparative purposes on request); carbon content 14 %; pH stability 1–9

Recommended application

- Analgesics, penicillin antibiotics, nucleic acid bases, water-soluble vitamins, complexing agents, organic acids

RP-HPLC with highly aqueous mobile phases

The efforts to neutralize unwanted silanol activity often results in well base-deactivated RP phases with high carbon load, but a limited scope of selectivity beyond non-polar interactions. Polar compounds like carboxylic acids or drug metabolites show only weak retention on densely bonded RP columns due to distinct hydrophobic properties but low polar interactions. Very polar analytes require highly aqueous mobile phases for solubility and retention. Conventional reversed phase columns often display stability problems in eluent systems with high percentage of water (> 95 %) as evidenced by a sudden decrease of retention time and overall poor reproducibility. This phenomenon is described as phase collapse caused by the mobile phase expelled from the pores due to the fact, that hydrophobic RP phases are incompletely wetted with the mobile phase [6].

Different approaches can be used to increase column stability with highly aqueous mobile phase systems. The most promising concepts are incorporating a polar group in the hydrophobic alkyl chain, or using hydrophilic endcapping procedures to improve the wettability of the reversed phase modification. NUCLEODUR® PolarTec may be taken as an example for the embedded polar group strategy, in which a C_{18} silane with a polar function is successfully linked to the silica surface.

Stability features

NUCLEODUR® C_{18} Pyramid is a silica phase with hydrophilic endcapping, designed especially for use in eluent systems of up to 100 % water. The upper figure shows the retention behavior of tartaric, acetic and maleic acid under purely aqueous conditions on NUCLEODUR® C_{18} Pyramid in comparison with a conventionally bonded C_{18} phase.

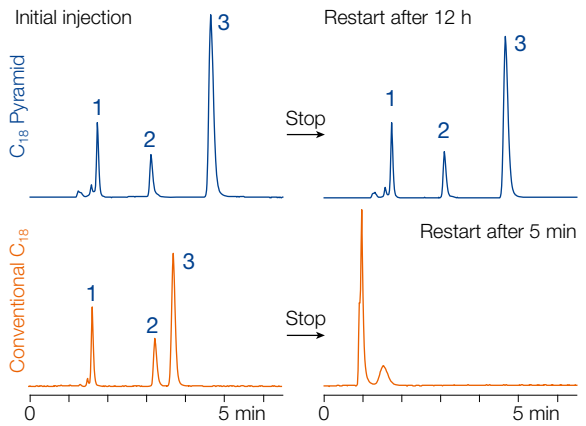
It can be shown that the retention times for NUCLEODUR® C_{18} Pyramid remain nearly unchanged between initial injection and restart after the flow has been stopped for 12 h, whilst the performance of the conventional RP column already collapsed totally after 5 min.

Stability test

MN Appl. No. 120870

Both columns:	125 x 4 mm
Eluent:	50 mmol/L KH_2PO_4, pH 2.5
Flow rate:	0.7 mL/min
Temperature:	25 °C
Detection:	UV, 210 nm
Injection:	1 µL

Peaks:
1. Tartaric acid
2. Acetic acid
3. Maleic acid



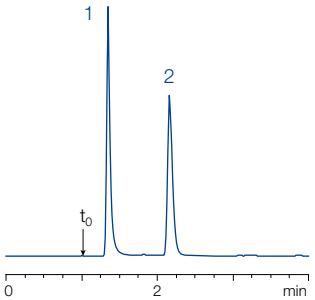
Retention characteristics

Separation of very polar compounds

MN Appl. No. 119170

Column:	125 x 4 mm NUCLEODUR® C_{18} Pyramid, 5 µm
Eluent:	0.2 % H_3PO_4
Flow rate:	1.0 mL/min
Temperature:	22 °C
Detection:	UV, 202 nm
Injection:	2 µL

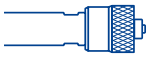
Peaks:
1. Formic acid
2. Acetic acid



The polar surface exhibits retention characteristics different from conventional C_{18} phases. Application 119170 shows the improved retention behavior of the very polar short chain organic acids, which are insufficiently retained on RP columns with predominantly hydrophobic surface properties. In addition to the exceptional polar selectivity NUCLEODUR® C_{18} Pyramid also provides adequate hydrophobic retention (see application No. 19190 at *www.mn-net.com*). The perceptible increase in polarity has no impact on the retention behavior of ionizable analytes. Even with the strongly basic compounds of the tricyclic antidepressant drug test mixture, no unwanted interactions or a so-called lack in base deactivation are observed (see application 119200 at *www.mn-net.com/apps*).

Ordering information

Eluent in column acetonitrile – water

	ID	Length → 30 mm	50 mm	75 mm	100 mm	125 mm	150 mm	250 mm
NUCLEODUR® C_{18} Pyramid, 1.8 µm particle size 1.8 µm · UHPLC								
Analytical EC columns								
	2 mm	760271.20	760272.20	760275.20	760273.20		760274.20	
	3 mm	760271.30	760272.30		760273.30			
	4 mm	760271.40	760272.40		760273.40			
	4.6 mm	760271.46	760272.46		760273.46			
EC guard columns*			4 x 2 mm: 761915.20		4 x 3 mm: 761915.30			
NUCLEODUR® C_{18} Pyramid, 3 µm particle size 3 µm								
Analytical EC columns								
	2 mm		760263.20		760264.20	760260.20	760261.20	760262.20
	3 mm		760263.30		760264.30	760260.30	760261.30	760262.30
	4 mm		760263.40		760264.40	760260.40	760261.40	760262.40
	4.6 mm		760263.46	760259.46	760264.46	760260.46	760261.46	760262.46
EC guard columns*			4 x 2 mm: 761916.20		4 x 3 mm: 761916.30			
NUCLEODUR® C_{18} Pyramid, 5 µm particle size 5 µm								
Analytical EC columns								
	2 mm		760200.20		760204.20	760201.20	760203.20	760202.20
	3 mm		760200.30		760204.30	760201.30	760203.30	760202.30
	4 mm		760200.40		760204.40	760201.40	760203.40	760202.40
	4.6 mm		760200.46	760205.46	760204.46	760201.46	760203.46	760202.46
EC guard columns*			4 x 2 mm: 761917.20		4 x 3 mm: 761917.30			
Preparative VarioPrep columns								
	10 mm		762271.100			762273.100		762272.100
	21 mm		762271.210			762273.210		762272.210
	32 mm							762272.320
	40 mm						762269.400	762272.400
VP guard columns **			10 x 8 mm: 762291.80		10 x 16 mm: 762291.160		15 x 32 mm: 762293.320	
EC and VarioPrep columns in packs of 1, guard columns see below.								

Guard column systems

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966
Guard columns for VarioPrep columns with ID		**8, 10 mm**	**16, 21 mm**	**32, 40 mm**	**≥ 50 mm**	
** VP guard columns (pack of)	VP	10/8 (2)	10/16 (2)	15/32 (1)	15/50 (1)	
VP guard column holder		718251	718256	718253	718255	

For details of our column systems see page 250.

NUCLEODUR® PolarTec RP phase with embedded polar group · USP L1 and L60

Key feature
- Excellent base deactivation
- Suitable for LC/MS and 100 % aqueous eluents
- Pronounced steric selectivity

Technical data
- Phase with embedded polar group; pore size 110 Å; particle sizes 1.8 µm, 3 µm and 5 µm; carbon content 17 %; pH stability 1–9

Recommended application
- Exceptional selectivity for phenols and nitrogen containing compounds, polar compounds like basic pharmaceuticals, organic acids, pesticides, amino acids, water-soluble vitamins, etc.

RP-HPLC under 100 % aqueous conditions

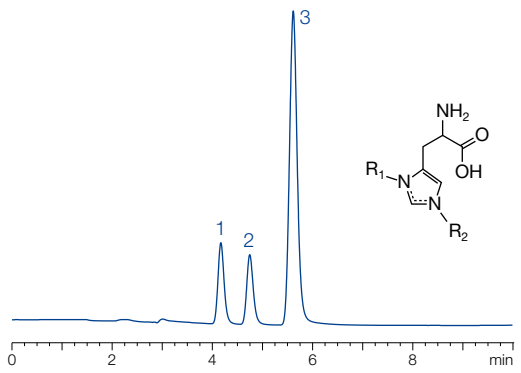
The dominant form of interactions of conventional C_{18} phases are nonpolar London dispersion forces. Besides nonpolar interactions phases with embedded polar groups possess the ability to show polar interactions (dipole-dipole, hydrogen bonds, π-π, etc.). These interactions enhance retention and selectivity for polar compounds like carboxylic acids, phenols and nitrogen containing compounds.

Separation of histidines

MN Appl. No. 125140

Column:	150 x 3 mm NUCLEODUR® PolarTec, 3 µm
Eluent:	1.0 mmol/L perfluoropentanoic acid in water – 0.5 mmol/L perfluoropentanoic acid in acetonitrile (99.5:0.5, v/v)
Flow rate:	0.4 mL/min
Temperature:	20 °C
Detection:	UV, 230 nm

Peaks:
1. 3-Methylhistidine R_1 = H, R_2 = CH_3
2. Histidine R_1 = R_2 = H
3. 1-Methylhistidine R_1 = CH_3, R_2 = H



In order to increase retention for polar compounds it is often necessary to decrease the organic ratio of the mobile phase to zero. Under these conditions many conventional C_{18} phases display the so-called dewetting effect which means that the mobile phase is expelled from the pores. This phenomenon leads to a dramatic loss in retention. NUCLEODUR® PolarTec is stable in 100 % aqueous mobile phases and therefore especially suited for the separation of polar compounds like organic acids.

Due to the shielding effect of the embedded group NUCLEODUR® PolarTec shows an excellent base deactivation, which is at the top-notch of embedded polar group phases on the market. The pronounced steric selectivity (see Tanaka plot) is an additional tool for the separation of complex mixtures.

Due to low bleeding characteristics NUCLEODUR® PolarTec is also suitable for LC/MS.

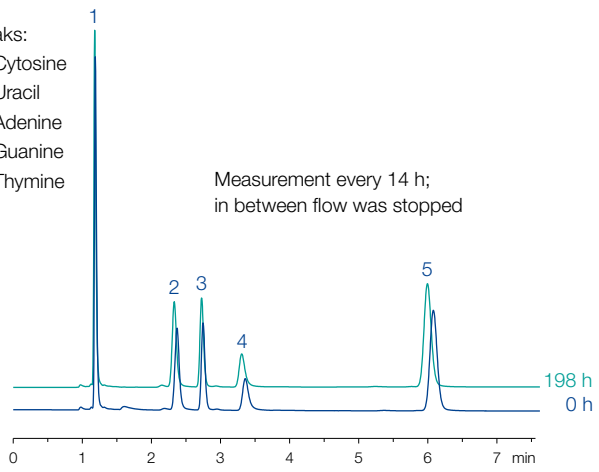
Even after days or weeks of operation in purely aqueous eluents the C_{18} chains of NUCLEODUR® PolarTec are neither folded nor show any collapsing. A significant reduction of retention time cannot be observed.

Stability of NUCLEODUR® PolarTec

MN Appl. No. 124610

Column:	150 x 3 mm NUCLEODUR® PolarTec, 3 µm
Eluent:	30 mmol/L KH_2PO_4, pH 3.0
Flow rate:	0.5 mL/min
Temperature:	30 °C
Detection:	UV, 220 nm

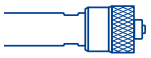
Peaks:
1. Cytosine
2. Uracil
3. Adenine
4. Guanine
5. Thymine



In spite of the polar character of the embedded functional group NUCLEODUR® PolarTec exhibits sufficient hydrophobic properties and is very well suited for analyzing basic compounds.

Ordering information

Eluent in column acetonitrile – water

	ID	Length → 30 mm	50 mm	75 mm	100 mm	125 mm	150 mm	250 mm
NUCLEODUR® PolarTec, 1.8 µm particle size 1.8 µm · UHPLC								
Analytical EC columns								
	2 mm	760461.20	760463.20	760465.20	760466.20		760468.20	
	3 mm	760461.30	760463.30		760466.30			
	4 mm	760461.40	760463.40		760466.40			
	4.6 mm	760461.46	760463.46		760466.46			
EC guard columns*			4 x 2 mm: 761980.20		4 x 3 mm: 761980.30			
NUCLEODUR® PolarTec, 3 µm particle size 3 µm								
Analytical EC columns								
	2 mm		760473.20		760476.20	760477.20	760478.20	760479.20
	3 mm		760473.30		760476.30	760477.30	760478.30	760479.30
	4 mm		760473.40		760476.40	760477.40	760478.40	760479.40
	4.6 mm		760473.46	760475.46	760476.46	760477.46	760478.46	760479.46
EC guard columns*			4 x 2 mm: 761981.20		4 x 3 mm: 761981.30			
NUCLEODUR® PolarTec, 5 µm particle size 5 µm								
Analytical EC columns								
	2 mm		760483.20		760486.20	760487.20	760488.20	760489.20
	3 mm		760483.30		760486.30	760487.30	760488.30	760489.30
	4 mm		760483.40		760486.40	760487.40	760488.40	760489.40
	4.6 mm		760483.46	760485.46	760486.46	760487.46	760488.46	760489.46
EC guard columns*			4 x 2 mm: 761982.20		4 x 3 mm: 761982.30			
Preparative VarioPrep columns								
	10 mm		762220.100			762221.100		762223.100
	21 mm		762220.210			762221.210		762223.210
	32 mm							762223.320
	40 mm						762222.400	762223.400
VP guard columns **			10 x 8 mm: 762224.80		10 x 16 mm: 762224.160		15 x 32 mm: 762226.320	

EC and VarioPrep columns in packs of 1, guard columns see below.

Guard column systems

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966
Guard columns for VarioPrep columns with ID		**8, 10 mm**	**16, 21 mm**	**32, 40 mm**	**≥ 50 mm**	
** VP guard columns (pack of)	VP	10/8 (2)	10/16 (2)	15/32 (1)	15/50 (1)	
VP guard column holder		718251	718256	718253	718255	

For details of our column systems see page 250.

NUCLEODUR® Phenyl-Hexyl productive for polar / aromatic compunds · USP L11

Key feature

- Hydrophobic phase with alternative selectivity compared to classical C_{18} modifications
- Separation principle based on 2 retention mechanisms: π-π interactions and hydrophobic interactions
- Suitable for LC/MS due to low bleeding characteristics

Technical data

- Phase with phenyl-hexyl modification and multi-endcapping; pore size 110 Å; particle sizes 1.8 µm, 3 µm and 5 µm; carbon content 10 %; pH stability 1–10

Recommended application

- Aromatic and unsaturated compounds, polar compounds like pharmaceuticals, antibiotics

Phenylhexyl modified phases are an interesting alternative to classical C_{18} phases due to an excellent separation of aromatic and unsaturated compounds especially with electron withdrawing groups.

The combination of hydrophobic and polar π-π interactions result in an interesting and alternate selectivity in comparison to C_{18} and C_8 modified phases.

Through short phenylhexyl chains the NUCLEODUR® Phenyl-Hexyl is more polar than the bifunctional modified NUCLEODUR® Sphinx RP. Therefore shorter analysis times can be achieved with mixtures of structural similar aromatic and aliphatic unsaturated compounds.

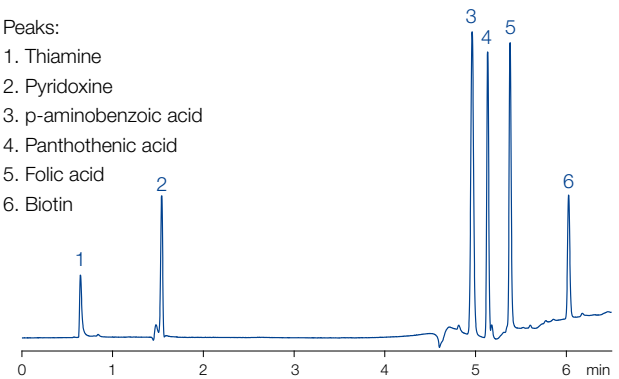
With NUCLEODUR® Phenyl-Hexyl e.g., tricyclic antidepressants or water soluble vitamins can be separated in good resolution.

Separation of water-soluble vitamins on NUCLEODUR® Phenyl-Hexyl

MN Appl. No. 125920

Column:	100 x 3 mm NUCLEODUR® Phenyl-Hexyl, 3 µm
Eluent:	A) 0.1 % phosphoric acid in water B) 0.1 % phosphoric acid in acetonitrile 0 % B for 2 min, then to 60 % B in 7 min
Flow rate:	0.56 mL/min
Temperature:	35 °C
Detection:	UV, 215 nm
Injection:	0.8 µL, 1.0 mg/mL each compound 1 mg/mL in eluent

Peaks:
1. Thiamine
2. Pyridoxine
3. p-aminobenzoic acid
4. Panthothenic acid
5. Folic acid
6. Biotin

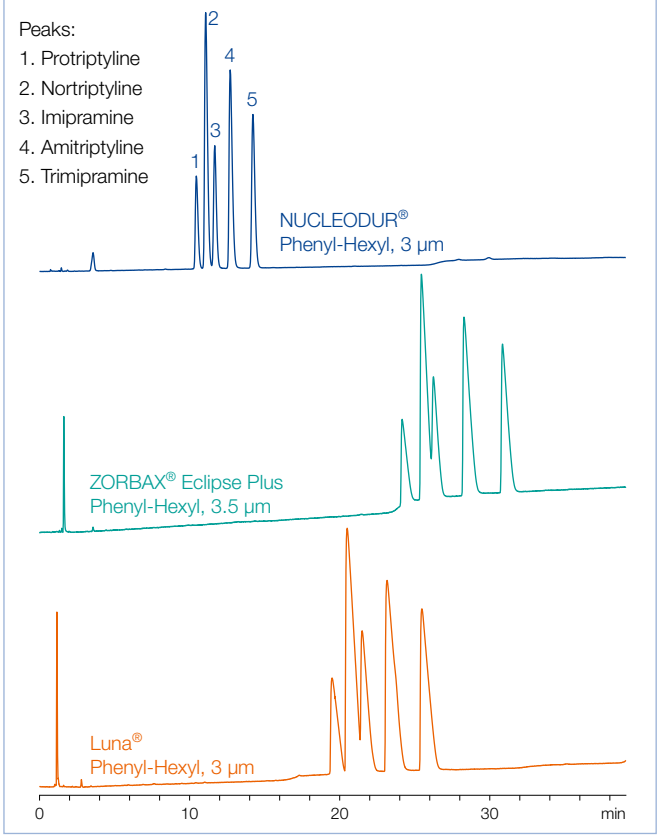


Tricyclic antidepressant (TCA)

MN Appl. No. 126020

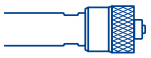
Columns:	150 x 3 mm NUCLEODUR® Phenyl-Hexyl, 3 µm Agilent ZORBAX® Eclipse Phenyl-Hexyl, 3.5 µm Phenomenex Luna® Phenyl-Hexyl, 3 µm
Eluent:	A) 0.1 % formic acid in acetonitrile B) 0.1 % formic acid in water 20–32.5 % A in 40 min
Flow rate:	0.56 mL/min
Temperature:	40 °C
Detection:	UV, 254 nm
Injection:	0.2 µL, each compound 1 mg/mL in eluent

Peaks:
1. Protriptyline
2. Nortriptyline
3. Imipramine
4. Amitriptyline
5. Trimipramine



Ordering information

Eluent in column acetonitrile – water

	ID	Length → 30 mm	50 mm	75 mm	100 mm	125 mm	150 mm	250 mm
NUCLEODUR® Phenyl-Hexyl, 1.8 µm particle size 1.8 µm · UHPLC								
Analytical EC columns								
	2 mm	760561.20	760563.20	760565.20	760566.20		760568.20	
	3 mm	760561.30	760563.30		760566.30			
	4 mm	760561.40	760563.40		760566.40			
	4.6 mm	760561.46	760563.46		760566.46			
EC guard columns*			4 x 2 mm: 761985.20		4 x 3 mm: 761985.30			
NUCLEODUR® Phenyl-Hexyl, 3 µm particle size 3 µm								
Analytical EC columns								
	2 mm		760573.20		760576.20	760577.20	760578.20	760579.20
	3 mm		760573.30		760576.30	760577.30	760578.30	760579.30
	4 mm		760573.40		760576.40	760577.40	760578.40	760579.40
	4.6 mm		760573.46	760575.46	760576.46	760577.46	760578.46	760579.46
EC guard columns*			4 x 2 mm: 761986.20		4 x 3 mm: 761986.30			
NUCLEODUR® Phenyl-Hexyl, 5 µm particle size 5 µm								
Analytical EC columns								
	2 mm		760583.20		760586.20	760587.20	760588.20	760589.20
	3 mm		760583.30		760586.30	760587.30	760588.30	760589.30
	4 mm		760583.40		760586.40	760587.40	760588.40	760589.40
	4.6 mm		760583.46	760585.46	760586.46	760587.46	760588.46	760589.46
EC guard columns*			4 x 2 mm: 761987.20		4 x 3 mm: 761987.30			
Preparative VarioPrep columns								
	10 mm		762210.100			762211.100		762213.100
	21 mm		762210.210			762211.210		762213.210
	32 mm							762213.320
	40 mm						762212.400	762213.400
VP guard columns **			10 x 8 mm: 762234.80		10 x 16 mm: 762234.160		15 x 32 mm: 762236.320	

EC and VarioPrep columns in packs of 1, guard columns see below.

Guard column systems

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966
Guard columns for VarioPrep columns with ID		**8, 10 mm**	**16, 21 mm**	**32, 40 mm**	**≥ 50 mm**	
** VP guard columns (pack of)	VP	10/8 (2)	10/16 (2)	15/32 (1)	15/50 (1)	
VP guard column holder		718251	718256	718253	718255	

For details of our column systems see page 250.

NUCLEODUR® π² hydrophobic biphenylpropyl phase · USP L11

Key feature

- Hydrophobic phase with alternative selectivity compared to classical C_{18} modifications
- Separation principle based on 2 retention mechanisms (π-π interactions and hydrophobic interactions)
- Better retention of aromatic and unsaturated substances
- Excellent performance under highly aqueous conditions

Technical data

- Phase with biphenylpropyl modification and multi-endcapping; pore size 110 Å; particle size 5 µm; carbon content 17 %; pH stability 1.5–10

Recommended application

- Overall sophisticated analytical separations, especially aromatic and unsaturated compounds, polar compounds like pharmaceuticals, antibiotics, steroids

Stationary HPLC phases with biphenyl ligands like NUCLEODUR® π² provide an interesting alternative to classical alkyl modified C_{18} and C_8 HPLC phases due to their remarkable orthogonal selectivity.

Furthermore the NUCLEODUR® π² provides an excellent separation performance for aromatic and unsaturated analytes by combination of hydrophobic and π-π interactions.

A unique feature is the predominant separation mechanism (π-π or hydrophobic interactions) and thus the selectivity can be controlled by selection of the eluent. In acetonitrile / water NUCLEODUR® π² shows similar retention strength then C_{18} modified phases and thereby displays a significantly stronger retention than phenyl phases. These interactions are even further enhanced in a methanol / water eluent.

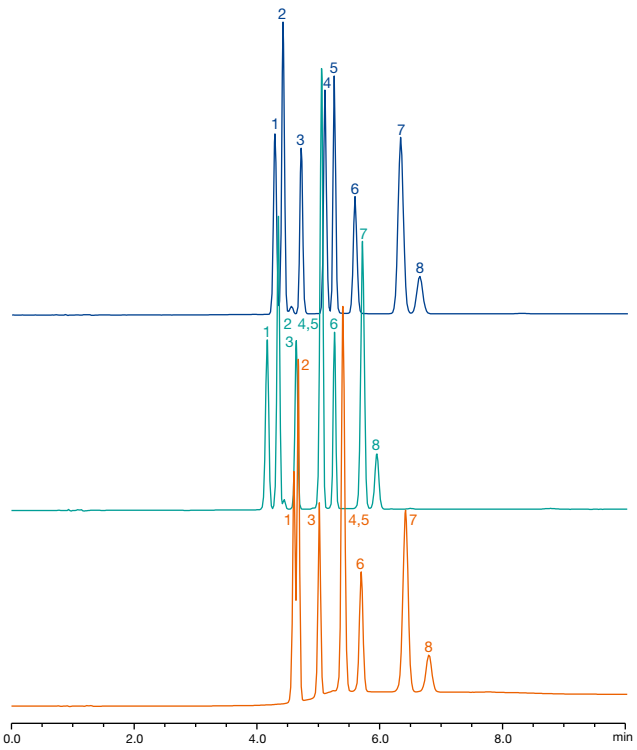
NUCLEODUR® π² exceeds other aryl phases in terms of stability under strongly aqueous conditions. Therefore i.a. steroids, sulfonamides and acidic pharmaceuticals are separated in good resolution with NUCLEODUR® π². NUCLEODUR® π² is the stationary phase with the highest aromatic analyte selectivity.

Sulfonamide antibiotics

MN Appl. No. 127920

Columns:	100 x 3 mm each NUCLEODUR® π², 5 µm Pinnacle® DB Biphenyl, 5 µm Ultra Biphenyl, 5 µm
Eluent:	A) 0.1 % TFA in water B) 0.1 % TFA in methanol 20 % B for 2 min, 20–60 % B in 2 min, 60 % B for 10 min
Flow rate:	0.56 mL/min
Temperature:	30 °C
Detection:	UV, 280 nm
Injection:	1 µL

Peaks:
1. Sulfathiazole
2. Sulfadiazine
3. Sulfachloropyridazine
4. Sulfamerazine
5. Sulfadimidine
6. Sulfamethoxazole
7. Sulfadimethoxine
8. Sulfaquinoxaline

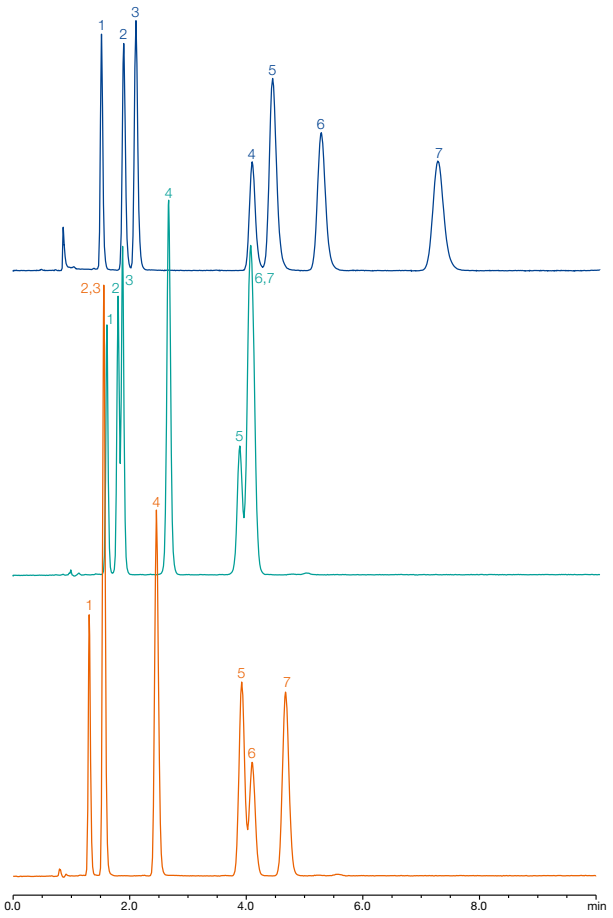


Steroids

MN Appl. No. 127910

Columns: 125 x 4 mm each
NUCLEODUR® π², 5 µm
NUCLEODUR® Phenyl-Hexyl, 5 µm
NUCLEODUR® C_{18} Gravity, 5 µm
Eluent: acetonitrile – water (45:55, v/v)
Injection: 1 µL
Flow rate: 1 mL/min
Temperature: 25 °C
Detection: UV, 230 nm

Peaks:
1. Estriol
2. Hydrocortisone
3. Prednisone
4. β-Estradiol
5. Corticosterone
6. Cortisonacetate
7. Testosterone



Ordering information

Eluent in column acetonitrile – water

ID	Length → 50 mm	75 mm	100 mm	125 mm	150 mm	250 mm
NUCLEODUR® π², 5 µm particle size 5 µm						
Analytical EC columns						
2 mm	760620.20	760621.20	760622.20	760623.20	760624.20	760625.20
3 mm	760620.30	760621.30	760622.30	760623.30	760624.30	760625.30
4 mm	760620.40	760621.40	760622.40	760623.40	760624.40	760625.40
4.6 mm	760620.46	760621.46	760622.46	760623.46	760624.46	760625.46
EC guard columns*	4 x 2 mm: 761810.20		4 x 3 mm: 761810.30			
EC columns in packs of 1, guard columns in packs of 3.						

Guard column systems

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966

For details of our column systems see page 250.

NUCLEODUR® PFP hydrophobic pentafluorophenyl phase · USP L43

Key feature

- Hydrophobic phase with alternative selectivity in comparison to classical C_{18} modifications
- Separation principle based on 4 retention mechanisms (polar interactions (H bonds), dipole-dipole, π-π, and hydrophobic interactions)
- Suitable for LC/MS due to low bleeding characteristics

Technical data

- Phase with pentafluorophenyl-propyl modification and multi-endcapping; pore size 110 Å; particle sizes 1.8 µm, 3 µm and 5 µm; carbon content 8 %; pH stability 1–9

Recommended application

- Aromatic and unsaturated compounds, phenols, halogen compounds, isomers, polar compounds like pharmaceuticals, antibiotics; strong retention of basic compounds

Orthogonality in selectivity

Fluorinated stationary phases in HPLC have gained increasing interest over the last years. Most common representative of fluorinated silica phases is the pentafluorophenyl modification (PFP or F_5). Especially the orthogonal selectivity compared to traditional alkyl phases widens the scope in analytical HPLC.

Thus NUCLEODUR® PFP offers an excellent selectivity especially for highly polar analytes like aromatic and unsaturated compounds, phenols or halogenated hydrocarbons.

While a typical C_{18} phase just provides hydrophobic interactions between stationary phase and analyte NUCLEODUR® PFP offers four different retention mechanisms: polar interactions (H bonds), dipole-dipole, π-π, and hydrophobic interactions. Especially the pronounced ion exchange capacity and distinct steric selectivity are typical for fluorinated phases.

Due to low bleeding characteristics NUCLEODUR® PFP is also suitable for LC/MS. Based on a special surface modification procedure NUCLEODUR® PFP offers highest stability also at low pH values.

NUCLEODUR® PFP offers a completely different retention behavior compared to alkyl modified silica and is often used for separations which provide insufficient results on traditional C_{18} phases.

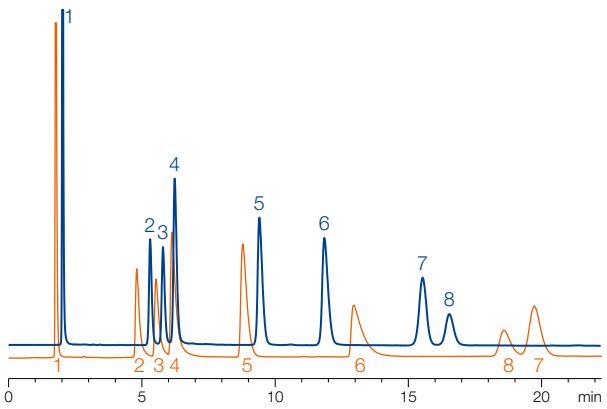
Applications in the areas of (bio-)pharma, natural compounds and environment show the broad applicability of this phase.

Separation of antihistamines

MN Appl. No. 124861

Columns: 250 x 3 mm NUCLEODUR® PFP, 5 µm
250 x 3 mm NUCLEODUR® C_{18} Gravity, 5 µm
Eluent: acetonitrile – 20 mmol/L KH_2PO_4 (30:70, v/v)
Flow rate: 1.3 mL/min
Temperature: 30 °C
Detection: UV, 210 nm

Peaks:
1. Maleic acid
2. Chlorpheniramine
3. Brompheniramine
4. Triprolidine
5. Diphenhydramine
6. Promethazine
7. Cetirizine
8. Hydroxyzine



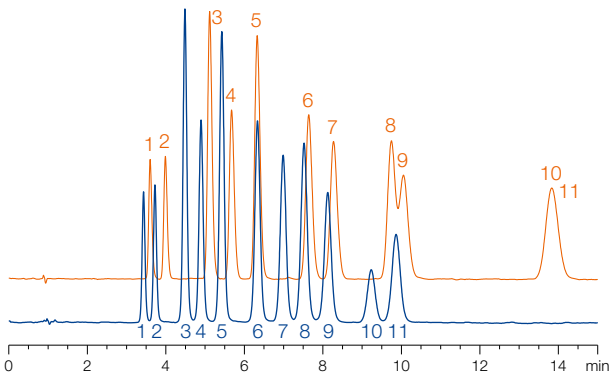
Separation of phenol isomers

MN Appl. No. 124531

Column: 125 x 4 mm NUCLEODUR® PFP, 5 µm
125 x 4 mm NUCLEODUR® C_{18} HTec, 5 µm
Eluent: acetonitrile, 0.1 % formic acid – water, 0.1 % formic acid (35:65, v/v)
Flow rate: 1 mL/min
Temperature: 35 °C
Detection: UV, 280 nm

Peaks:

1. *o*-Kresol
2. *m*-Kresol
3. 3,4-Dimethylphenol
4. 3,5-Dimethylphenol
5. 2,5-Dimethylphenol
6. 2,6-Dichlorophenol
7. 2,3-Dichlorophenol
8. 2,4-Dichlorophenol
9. 3,4-Dichlorophenol
10. 2,4-Dibromophenol
11. 3,5-Dibromophenol



Ordering information

Eluent in column acetonitrile – water

ID	Length → 30 mm	50 mm	75 mm	100 mm	125 mm	150 mm	250 mm
NUCLEODUR® PFP, 1.8 µm particle size 1.8 µm · UHPLC							
Analytical EC columns							
2 mm	760431.20	760433.20	760435.20	760436.20		760438.20	
3 mm	760431.30	760433.30		760436.30			
4 mm	760431.40	760433.40		760436.40			
4.6 mm	760431.46	760433.46		760436.46			
EC guard columns*		4 x 2 mm: 761975.20		4 x 3 mm: 761975.30			
NUCLEODUR® PFP, 3 µm particle size 3 µm							
Analytical EC columns							
2 mm		760443.20		760446.20	760447.20	760448.20	760449.20
3 mm		760443.30		760446.30	760447.30	760448.30	760449.30
4 mm		760443.40		760446.40	760447.40	760448.40	760449.40
4.6 mm		760443.46	760445.46	760446.46	760447.46	760448.46	760449.46
EC guard columns*		4 x 2 mm: 761976.20		4 x 3 mm: 761976.30			
NUCLEODUR® PFP, 5 µm particle size 5 µm							
Analytical EC columns							
2 mm		760453.20		760456.20	760457.20	760458.20	760459.20
3 mm		760453.30		760456.30	760457.30	760458.30	760459.30
4 mm		760453.40		760456.40	760457.40	760458.40	760459.40
4.6 mm		760453.46	760455.46	760456.46	760457.46	760458.46	760459.46
EC guard columns*		4 x 2 mm: 761977.20		4 x 3 mm: 761977.30			
Preparative VarioPrep columns							
10 mm		762210.100			762211.100		762213.100
21 mm		762210.210			762211.210		762213.210
32 mm							762213.320
40 mm						762212.400	762213.400
VP guard columns **		10 x 8 mm: 762214.80		10 x 16 mm: 762214.160		15 x 32 mm: 762216.320	

EC and VarioPrep columns in packs of 1, guard columns see below.

Guard column systems

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966
Guard columns for VarioPrep columns with ID		**8, 10 mm**	**16, 21 mm**	**32, 40 mm**	**≥ 50 mm**	
** VP guard columns (pack of)	VP	10/8 (2)	10/16 (2)	15/32 (1)	15/50 (1)	
VP guard column holder		718251	718256	718253	718255	

For details of our column systems see page 250.

NUCLEODUR® columns

NUCLEODUR® Sphinx RP bifunctional RP phase · USP L1 and L11

Key feature

- Distinct selectivity based on well-balanced bifunctional surface coverage
- Widens the scope for method development based on additional π-π interactions
- Suitable for LC/MS due to low bleeding characteristics

Technical data

- Octadecyl and propylphenyl modified silica; pore size 110 Å; particle sizes 1.8 µm, 3 µm and 5 µm; carbon content 15 %; pH stability 1–10; high reproducibility and consistent quality

Recommended application

- Quinolone antibiotics, sulfonamides, xanthines, substituted aromatics

Alternative RP selectivity

NUCLEODUR® Sphinx RP is characterized by exceptional selectivity features generated by a well-balanced ratio of covalently bonded octadecyl and phenyl groups. The combination of classical hydrophobic with π-π interactions (aromatic ring system) expands the scope of selectivity in comparison with conventional reversed phase packings. NUCLEODUR® Sphinx RP is particularly suited for the separation of molecules containing aromatic and multiple bonds.

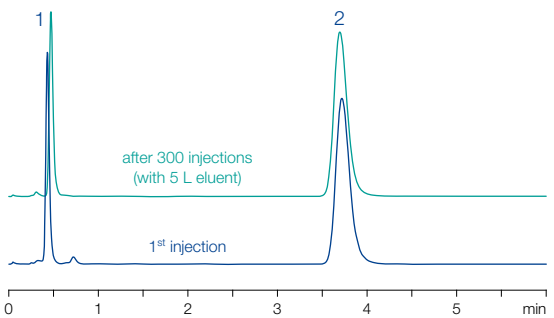
For the separation of polar compounds NUCLEODUR® Sphinx RP can be especially recommended and can also outperform many customary C_{18} phases. In addition, exhaustive endcapping steps minimize unwanted surface silanol activity and guarantee excellent peak shapes even for strong basic analytes.

Stability of NUCLEODUR® Sphinx RP at pH 10

MN Appl. No. 120900

Column:	50 x 4.6 mm NUCLEODUR® Sphinx RP, 5 µm
Eluent:	methanol – dil. NH_3, pH 10 (20:80, v/v)
Flow rate:	1.0 mL/min, temperature 30 °C
Detection:	UV, 275 nm
Injection:	3 µL

Peaks:
1. Theophylline
2. Caffeine

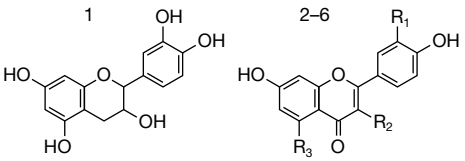


Different from standard phenyl phases, NUCLEODUR® Sphinx RP is far more stable towards hydrolysis and is also suggested for LC/MS applications. Due to the additional intermolecular interactions NUCLEODUR® Sphinx RP is an interesting replenishment to the high density bonded phases NUCLEODUR® C_8/C_{18} Gravity and the polar endcapped NUCLEODUR® C_{18} Pyramid.

Separation of flavonoids on three different NUCLEODUR® phases

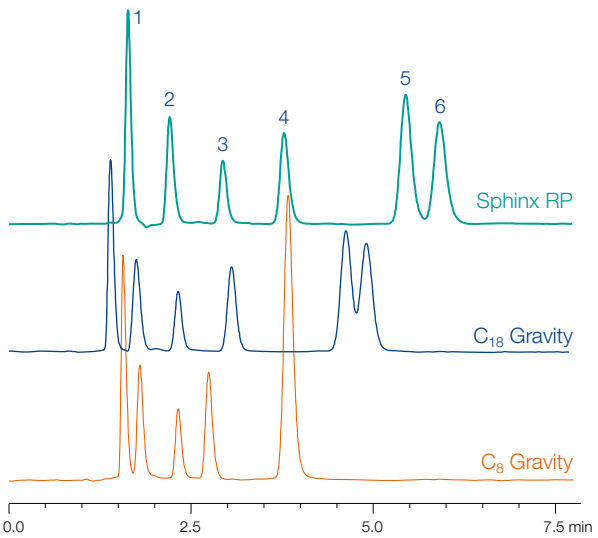
MN Appl. No. 119830

Columns:	150 x 4.6 mm NUCLEODUR® Sphinx RP, 5 µm NUCLEODUR® C_{18} Gravity, 5 µm NUCLEODUR® C_8 Gravity, 5 µm
Eluent:	water – methanol (40:60, v/v)
Flow rate:	1 mL/min
Temperature:	30 °C
Detection:	UV, 270 nm
Injection:	3 µL



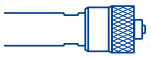
Peaks:

1. Catechin	
2. Rutin	$R_1 = R_3 = OH$, R_2 = O-Rutinose
3. Fisetin	$R_1 = R_2 = OH$, $R_3 = H$
4. Quercetin	$R_1 = R_2 = R_3 = OH$
5. Kaempferol	$R_1 = H$, $R_2 = R_3 = OH$
6. Isorhamnetin	$R_1 = OCH_3$, $R_2 = R_3 = OH$



Ordering information

Eluent in column acetonitrile – water

	ID	Length → 30 mm	50 mm	75 mm	100 mm	125 mm	150 mm	250 mm
NUCLEODUR® Sphinx RP, 1.8 µm particle size 1.8 µm · UHPLC								
Analytical EC columns								
	2 mm	760821.20	760822.20	760825.20	760823.20		760824.20	
	3 mm	760821.30	760822.30		760823.30			
	4 mm	760821.40	760822.40		760823.40			
	4.6 mm	760821.46	760822.46		760823.46			
EC guard columns*			4 x 2 mm: 761920.20		4 x 3 mm: 761920.30			
NUCLEODUR® Sphinx RP, 3 µm particle size 3 µm								
Analytical EC columns								
	2 mm		760806.20		760812.20	760807.20	760805.20	760808.20
	3 mm		760806.30		760812.30	760807.30	760805.30	760808.30
	4 mm		760806.40		760812.40	760807.40	760805.40	760808.40
	4.6 mm		760806.46	760813.46	760812.46	760807.46	760805.46	760808.46
EC guard columns*			4 x 2 mm: 761921.20		4 x 3 mm: 761921.30			
NUCLEODUR® Sphinx RP, 5 µm particle size 5 µm								
Analytical EC columns								
	2 mm		760800.20		760809.20	760801.20	760802.20	760803.20
	3 mm		760800.30		760809.30	760801.30	760802.30	760803.30
	4 mm		760800.40		760809.40	760801.40	760802.40	760803.40
	4.6 mm		760800.46	760815.46	760809.46	760801.46	760802.46	760803.46
EC guard columns*			4 x 2 mm: 761922.20		4 x 3 mm: 761922.30			
Preparative VarioPrep columns								
	10 mm		762372.100			762375.100		762373.100
	21 mm		762372.210			762375.210		762373.210
	32 mm							762373.320
	40 mm						762371.400	762373.400
VP guard columns **			10 x 8 mm: 762390.80		10 x 16 mm: 762390.160		15 x 32 mm: 762392.320	

EC and VarioPrep columns in packs of 1, guard columns see below.

Guard column systems

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966
Guard columns for VarioPrep columns with ID		**8, 10 mm**	**16, 21 mm**	**32, 40 mm**	**≥ 50 mm**	
** VP guard columns (pack of)	VP	10/8 (2)	10/16 (2)	15/32 (1)	15/50 (1)	
VP guard column holder		718251	718256	718253	718255	

For details of our column systems see page 250.

NUCLEODUR® C_{18} HTec base-deactivated preparative octadecyl phase · USP L1

Key feature

- Reliable and durable standard RP phase for up-scaling to preparative scale, suited for LC/MS
- High loading capacity and excellent stability
- Outstanding base deactivation

Technical data

- High density octadecyl modification (C_{18}); pore size 110 Å; particle sizes 1.8 µm, 3 µm, 5 µm, 7 µm and 10 µm for analytical and preparative separations; carbon content 18 %, pH stability 1–11

Recommended application

- Sophisticated analytical and preparative separations of basic, neutral and acidic pharmaceuticals, derivatized amino acids, pesticides, fat-soluble vitamins, aldehydes, ketones and phenolic compounds

Preparative separations place high demands on silica based HPLC materials. Apart from excellent selectivity and base deactivation, robustness (pH, pressure stability, …) and capacity are vital criteria for optimal and efficient separation at the preparative scale.

Selectivity and base deactivation

The innovative endcapping procedure leads to exceptionally good base deactivation – the Engelhardt test demonstrates superb selectivity, peak symmetry and peak shape over the entire polarity range. In addition NUCLEODUR® C_{18} HTec scores in low bleed characteristics and is therefore highly suitable for LC/MS.

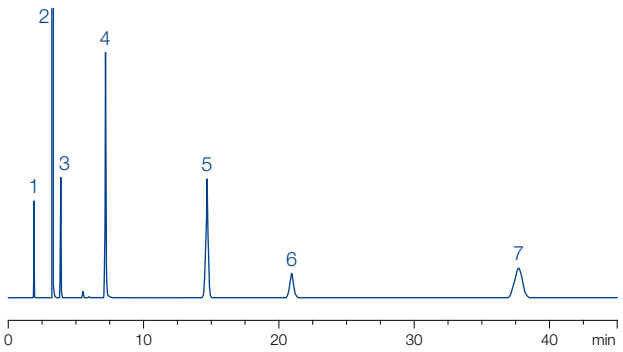
Engelhardt test

MN Appl. No. 123580

Column:	250 x 4 mm NUCLEODUR® C_{18} HTec, 5 µm
Eluent:	methanol – water (49:51, v/v)
Flow rate:	1 mL/min
Temperature:	40 °C
Detection:	UV, 254 nm
Injection:	5 µL

Peaks:

1. Uracil
2. Aniline
3. Phenol
4. *p*-Ethylaniline
5. *N*,*N*-Dimethylaniline
6. Toluene
7. Ethylbenzene



Stability and lifetime

Based on fully synthetic and extremely robust totally spherical NUCLEODUR® silica, NUCLEODUR® C_{18} HTec offers outstanding mechanical rigidity and is thus the perfect choice also for self-packing of prep-columns. The special surface modification and endcapping procedure results in high chemical stability even at extreme chromatographic conditions like high flow rates, temperature or critical solvents (DMSO). Furthermore, NUCLEODUR® C_{18} HTec columns show a remarkably long lifetime in acidic (pH 1) as well as basic (pH 10) mobile phases.

pH stability test

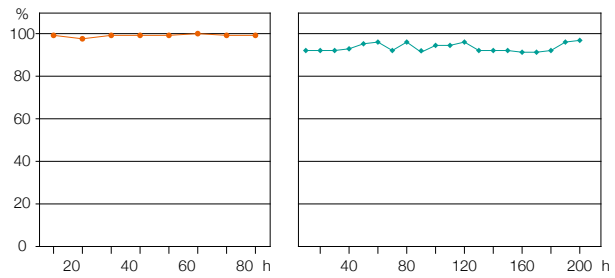
Column:	150 x 4 mm NUCLEODUR® C_{18} HTec, 5 µm
Flow rate:	1 mL/min
Detection:	UV, 254 nm
Injection:	5 µL

• pH 1:

Eluent: acetonitrile – 1 % TFA in water (50:50, v/v);
80 °C
% initial retention of ethylbenzene
693 injections

• pH 10:

Eluent: methanol – 50 mmol/L triethylamine (25:85, v/v);
50 °C
% initial N of theophylline
1034 injections



Due to innovative surface coating procedures NUCLEODUR® C_{18} HTec offers excellent analytical separation properties and is the first choice for up-scaling to preparative column dimensions.

Up-scaling

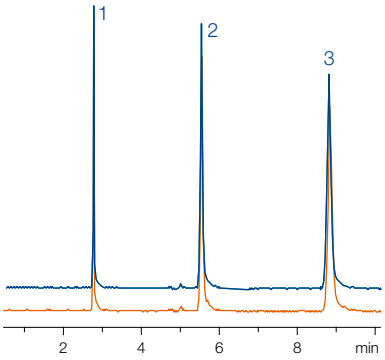
Due to highest quality standards in silica production and phase chemistry combined with optimized packing technology, NUCLEODUR® C_{18} HTec allows exceptional transferability from analytical to preparative scale with respect to different particle sizes (e.g., 5, 7 or 10 µm) as well as column dimensions (e.g., ID 4.6 to 21 mm).

Up-scaling with NUCLEODUR® C_{18} HTec

MN Appl. No. 123780

Columns:	EC 250 x 4,6 mm NUCLEODUR® C_{18} HTec, 5 µm VP 250 x 21 mm NUCLEODUR® C_{18} HTec, 5 µm
Eluent:	acetonitrile – water (80:20, v/v)
Flow rate:	1.3 mL/min / 27 mL/min
Temperature:	22 °C
Pressure:	84 bar / 109 bar
Detection:	UV, 254 nm
Injection:	3 µL / 60 µL

Peaks: (1 mg/mL each)
1. Phenol
2. Naphthalene
3. Anthracene



Capacity

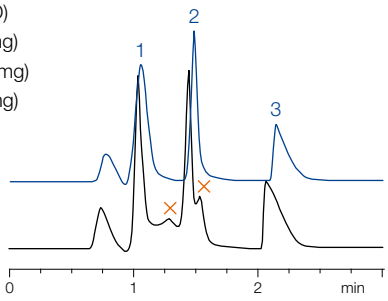
A vital criterion for efficiency in preparative HPLC is the capacity of the separation medium. NUCLEODUR® C_{18} HTec is characterized by a notably high loading capacity under both basic and acidic conditions, while competitor columns show overload effects even at lower loads (x).

Loading capacity under acidic conditions

MN Appl. No. 123890

Columns:	VP 100 x 21 mm NUCLEODUR® C_{18} HTec, 5 µm 100 x 21.2 mm AXIA™ Gemini® 5 µm C18 110 Å
Eluent:	acetonitrile – formic acid in H_2O pH 3.0 (30:70, v/v)
Flow rate:	28 mL/min
Temperature:	22 °C
Pressure:	124 bar
Detection:	UV, 254 nm

Peaks:
total load 40 mg
(sample dissolved in DMSO)
1. 4-Acetamidophenol (5 mg)
2. 2-Acetamidophenol (10 mg)
3. Acetylsalicylic acid (25 mg)



Ordering information

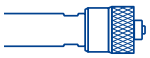
Eluent in column acetonitrile – water

ID	Length → 30 mm	50 mm	75 mm	100 mm	125 mm	150 mm	250 mm
NUCLEODUR® C_{18} HTec, 1.8 µm particle size 1.8 µm · UHPLC							
Analytical EC columns							
2 mm	760301.20	760305.20	760304.20	760306.20		760308.20	
3 mm	760301.30	760305.30		760306.30			
4 mm	760301.40	760305.40		760306.40			
4.6 mm	760301.46	760305.46		760306.46			
EC guard columns*		4 x 2 mm: 761925.20		4 x 3 mm: 761925.30			
NUCLEODUR® C_{18} HTec, 3 µm particle size 3 µm							
Analytical EC columns							
2 mm		760321.20		760323.20	760324.20	760325.20	760326.20
3 mm		760321.30		760323.30	760324.30	760325.30	760326.30
4 mm		760321.40		760323.40	760324.40	760325.40	760326.40
4.6 mm		760321.46	760322.46	760323.46	760324.46	760325.46	760326.46
EC guard columns*		4 x 2 mm: 761926.20		4 x 3 mm: 761926.30			

NUCLEODUR® columns

Ordering information

Eluent in column acetonitrile – water

	ID	Length → 30 mm	50 mm	75 mm	100 mm	125 mm	150 mm	250 mm
NUCLEODUR® C_{18} HTec, 5 µm particle size 5 µm								
Analytical EC columns								
	2 mm		760311.20		760313.20	760314.20	760315.20	760316.20
	3 mm		760311.30		760313.30	760314.30	760315.30	760316.30
	4 mm		760311.40		760313.40	760314.40	760315.40	760316.40
	4.6 mm		760311.46	760312.46	760313.46	760314.46	760315.46	760316.46
EC guard columns*			4 x 2 mm: 761927.20		4 x 3 mm: 761927.30			
Preparative VarioPrep columns								
	10 mm		762551.100			762554.100		762556.100
	21 mm		762551.210		762553.210	762554.210		762556.210
	32 mm				762553.320		762555.320	762556.320
	40 mm						762555.400	762556.400
	50 mm				762553.500		762555.500	762556.500
VP guard columns **			10 x 8 mm: 762591.80		10 x 16 mm: 762591.160			
			15 x 32 mm: 762592.320		15 x 50 mm: 762592.500			
NUCLEODUR® C_{18} HTec, 7 µm particle size 7 µm								
Preparative VarioPrep columns								
	10 mm		762561.100			762564.100		762566.100
	21 mm		762561.210		762563.210	762564.210		762566.210
	32 mm				762563.320		762565.320	762566.320
	40 mm						762565.400	762566.400
	50 mm				762563.500		762565.500	762566.500
VP guard columns **			10 x 8 mm: 762591.80		10 x 16 mm: 762591.160			
			15 x 32 mm: 762592.320		15 x 50 mm: 762592.500			
NUCLEODUR® C_{18} HTec, 10 µm particle size 10 µm								
Preparative VarioPrep columns								
	10 mm		762571.100			762574.100		762576.100
	21 mm		762571.210		762573.210	762574.210		762576.210
	32 mm				762573.320		762575.320	762576.320
	40 mm						762575.400	762576.400
	50 mm				762573.500		762575.500	762576.500
VP guard columns **			10 x 8 mm: 762591.80		10 x 16 mm: 762591.160			
			15 x 32 mm: 762592.320		15 x 50 mm: 762592.500			

EC and VarioPrep columns in packs of 1, guard columns see below.

Guard column systems

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966
Guard columns for VarioPrep columns with ID		**8, 10 mm**	**16, 21 mm**	**32, 40 mm**	**≥ 50 mm**	
** VP guard columns (pack of)	VP	10/8 (2)	10/16 (2)	15/32 (1)	15/50 (1)	
VP guard column holder		718251	718256	718253	718255	

For details of our column systems see page 250.
NUCLEODUR® C_{18} HTec bulk material in 7 and 10 µm for self-packing of preparative columns see page 256.

NUCLEODUR® C_{18} ec · C_8 ec · C_4 ec nonpolar phases for routine analysis · USP L1 (C_{18}) · L7 (C_8) · L26 (C_4)

Key feature

- Ideal and reliable standard RP phase for daily routine analysis and up-scaling for preparative HPLC
- Medium density Octadecyl (C_{18}) and octyl (C_8) with pore size of 110 Å with exhaustive endcapping for a wide range of applications
- Octadecyl (C_{18}) and butyl (C_4) with pore size of 300 Å for the separation of biomolecules

Technical data

- Pore size 110 Å:
  particle sizes 3 µm and 5 µm, 7 µm, 10 µm, 12 µm, 16 µm, 20 µm, 30 µm and 50 µm for preparative separations; carbon content 17.5 % for C_{18}, 10.5 % for C_8; pH stability 1–9; high reproducibility from lot to lot
- Pore size 300 Å:
  technical data and applications in chapter “HPLC column for biochemical separations” (see page 241)

Recommended application

- 110 Å:
  basic, neutral or acidic drugs; derivatized amino acids; pesticides; fat-soluble vitamins; aldehydes and ketones; phenolic compounds
- 300 Å:
  biomolecular macromolecules, like proteins and peptides

NUCLEODUR® C_{18} ec for daily routine analysis

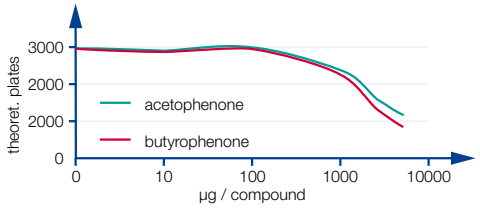
The efficiency of a separation is controlled by particle size and selectivity of the stationary phase. The exceptional surface coverage of monomeric bonded alkylsilanes, combined with an exhaustive endcapping, results in a surface with lowest silanol activity. This allows the tailing-free elution of polar compounds such as basic drugs. NUCLEODUR® C_{18} ec is available in 9 different particle sizes (3, 5, 7, 10, 12, 16, 20, 30 and 50 µm) which cover the whole range from high speed analytical HPLC up to medium and low pressure prep LC. NUCLEODUR® C_{18} ec is also an ideal tool for scale-up purposes.

Loading capacity

Loading capacity, probably the most important feature for preparative LC applications, is determined by pore size, pore volume and surface area of the packing. However, it can also be influenced by the molecular weight of the analytes. In the figure below the mass loading curve for acetophenone and butyrophenone on a NUCLEODUR® 100-20 C_{18} ec column describes the correlation between the increase of column loading and the decrease of separation efficiency.

Loading curve

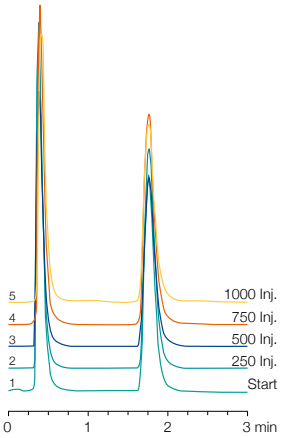
Column: 250 x 4.6 mm NUCLEODUR® 100-20 C_{18} ec
Eluent: acetonitrile – H_2O 80:20 (v/v)
Flow rate: 1.0 mL/min
Temperature: 25 °C
Detection: UV, 280 – 370 nm



pH stability of NUCLEODUR® C_{18} ec

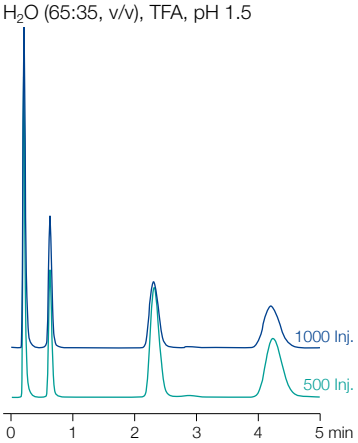
Separation of theophylline and caffeine at pH 10

Column: 30 x 3 mm NUCLEODUR® 100-5 C_{18} ec
Eluent: methanol – aq. NH_3 (20:80, v/v), pH 10
Flow rate: 0.5 mL/min
Temperature: 25 °C
Detection: UV, 254 nm



Separation of uracil, veratrol, toluene and ethylbenzene at pH 1.5

Column: 30 x 3 mm NUCLEODUR® 100-5 C_{18} ec
Eluent: acetonitrile – H_2O (65:35, v/v), TFA, pH 1.5
Flow rate: 1.0 mL/min
Temperature: 25 °C
Detection: UV, 254 nm



Chemical stability

The utmost purity of the base silica and the exceptional silane bonding chemistry minimize the risk of dissolution, or hydrolysis at pH extremes.

The chromatograms show the retention behavior at pH values of 1.5 and 10.0 for NUCLEODUR® 100-5 C_{18} ec.

NUCLEODUR® octyl phases

In addition to NUCLEODUR® C_{18} phases MACHEREY-NAGEL offers octyl modified NUCLEODUR® C_8 Gravity and NUCLEODUR® C_8 ec columns to expand the RP tool box. Based on the same spherical high purity silica the C_8 phases exhibit the same chemical and mechanical stability as the C_{18} counterparts. Indeed NUCLEODUR® C_8 Gravity can also be run at pH extremes (pH 1–11) by choosing appropriate elution parameters. Due to the shorter chain and less hydrophobic properties of the stationary phase the retention of non-polar compounds is decreased, and in consequence a reduction in time of analysis can be achieved. Moreover a stronger polar selectivity, particularly with the separation of ionizable analytes is frequently observed (as distinct from the C_{18} phases). NUCLEODUR® C_8 ec and NUCLEODUR® C_8 Gravity are most suitable for the development of new methods but also for robust routine analyses.

There are no general guidelines which could make the choice between C_8 and C_{18} phases easier but it will always be beneficial to add both phases to the existing pool of RP columns in the laboratory. Comparative studies reveal some different selectivity patterns of NUCLEODUR® C_8 ec and C_{18} ec. The separation of phenols at right shows baseline separation for 2-ethoxyphenol and dimethoxybenzene (veratrol) and in addition a reversal of the elution order of phenol and 4-methoxyphenol can be shown on the octyl phase.

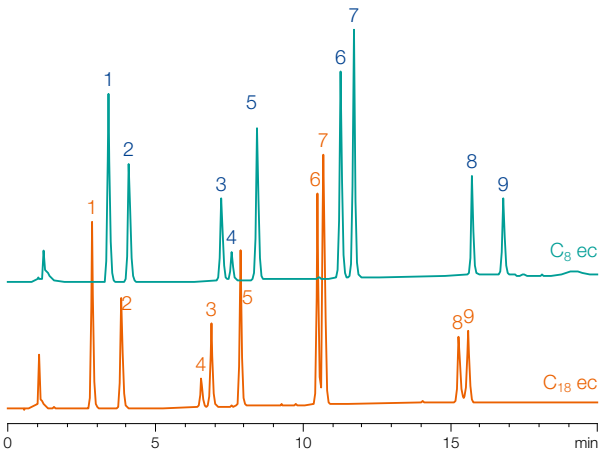
Separation of phenols

MN Appl. Nos. 120890 / 120891

Columns:	250 x 4 mm NUCLEODUR® 100-5 C_8 ec / C_{18} ec
Eluent:	A) water, B) methanol
	C_8: 20 % B (2 min) → 60 % B in 12 min
	C_{18}: 25 % B (2 min) → 65 % B in 12 min
Flow rate:	1.0 mL/min, temperature 25 °C
Detection:	UV, 275 nm, injection 10 µL

Peaks:

1. Resorcinol
2. Pyrocatechol
3. 4-Methoxyphenol
4. Phenol
5. 2-Methoxyphenol
6. 2-Ethoxyphenol
7. Veratrol
8. Biphenyl-2-ol
9. Phenetole



NUCLEODUR® phases for biochromatography

A description and applications for C_{18} and C_4 modified 300 Å NUCLEODUR® widepore materials for the separation of biopolymers, like peptids and proteins can be found in chapter “HPLC column for biochemical separations” (see page 241).

C_{18} or C_8 · the best of both worlds

· Octyl phases (C_8) show superior polar selectivity.
· Octadecyl phases (C_{18}) show superior hydrophobic selectivity.
· Hydrophobic compounds show shorter retention times on C_8 phases.

Ordering information

Eluent in column acetonitrile – water

	ID	Length → 50 mm	75 mm	100 mm	125 mm	150 mm	250 mm
NUCLEODUR® 100-3 C_{18} ec octadecyl phase, particle size 3 µm, 17.5 % C							
Analytical EC columns							
	2 mm	760050.20		760054.20	760051.20	760053.20	760052.20
	3 mm	760050.30		760054.30	760051.30	760053.30	760052.30
	4 mm	760050.40		760054.40	760051.40	760053.40	760052.40
	4.6 mm	760050.46	760046.46	760054.46	760051.46	760053.46	760052.46
EC guard columns*			4 x 2 mm: 761931.20		4 x 3 mm: 761931.30		

Ordering information

Eluent in column acetonitrile – water

	ID	Length → 50 mm	75 mm	100 mm	125 mm	150 mm	250 mm
NUCLEODUR® 100-5 C_{18} ec octadecyl phase, particle size 5 µm, 17.5 % C							
Analytical EC columns							
	2 mm	760004.20		760013.20	760001.20	760008.20	760002.20
	3 mm	760004.30		760013.30	760001.30	760008.30	760002.30
	4 mm	760004.40		760013.40	760001.40	760008.40	760002.40
	4.6 mm	760004.46	760035.46	760013.46	760001.46	760008.46	760002.46
EC guard columns*			4 x 2 mm: 761932.20		4 x 3 mm: 761932.30		
Preparative VarioPrep columns							
	10 mm	762003.100			762029.100		762022.100
	21 mm	762003.210			762029.210		762022.210
	32 mm						762022.320
	40 mm					762027.400	762022.400
VP guard columns **			10 x 8 mm: 762090.80		10 x 16 mm: 762090.160		
			15 x 32 mm: 762311.320		15 x 50 mm: 762311.500		
NUCLEODUR® 100-10 C_{18} ec octadecyl phase, particle size 10 µm, 17.5 % C							
Preparative VarioPrep columns							
	10 mm	762011.100			762302.100		762010.100
	21 mm	762011.210			762302.210		762010.210
	32 mm						762010.320
	40 mm					762303.400	762010.400
	50 mm						762010.500
VP guard columns **			10 x 8 mm: 762090.80		10 x 16 mm: 762090.160		
			15 x 32 mm: 762311.320		15 x 50 mm: 762311.500		

Ordering information

Eluent in column acetonitrile – water

	ID	Length → 50 mm	75 mm	100 mm	125 mm	150 mm	250 mm
NUCLEODUR® 100-3 C_8 ec octyl phase, particle size 3 µm, 10.5 % C							
Analytical EC columns							
	2 mm	760063.20		760059.20	760060.20		760062.20
	3 mm	760063.30		760059.30	760060.30		760062.30
	4 mm	760063.40		760059.40	760060.40		760062.40
	4.6 mm	760063.46	760064.46	760059.46	760060.46	760061.46	760062.46
EC guard columns*			4 x 2 mm: 761936.20		4 x 3 mm: 761936.30		
NUCLEODUR® 100-5 C_8 ec octyl phase, particle size 5 µm, 10.5 % C							
Analytical EC columns							
	2 mm	760700.20		760704.20	760701.20		760703.20
	3 mm	760700.30		760704.30	760701.30		760703.30
	4 mm	760700.40		760704.40	760701.40		760703.40
	4.6 mm	760700.46	760706.46	760704.46	760701.46	760702.46	760703.46
EC guard columns*			4 x 2 mm: 761937.20		4 x 3 mm: 761937.30		
Preparative VarioPrep columns							
	10 mm	762072.100			762061.100		762062.100
	21 mm	762072.210			762061.210		762062.210
	32 mm						762062.320
	40 mm					762079.400	762062.400
VP guard columns **		10 x 8 mm: 762092.80		10 x 16 mm: 762092.160		15 x 32 mm: 762321.320	
EC and VarioPrep columns in packs of 1, guard columns see previous NUCLEODUR® phases.							

Guard column systems see previous NUCLEODUR® phases. For details of our column systems see page 250.
NUCLEODUR® C_{18} ec bulk material with 10–50 µm for self-packing of preparative columns see page 256.
The ordering information for C_{18} and C_4 modified 300 Å NUCLEODUR® widepore materials for the separation of biopolymers can be found in the chapter "HPLC column for biochemical separations" (see page 241).
* and ** for corresponding guard column systems see page 180.

NUCLEODUR® HILIC zwitterionic phase

Key feature

- Ideal for reproducible and stable chromatography of highly polar analytes
- Suitable for analytical and preparative applications
- Very short column conditioning period

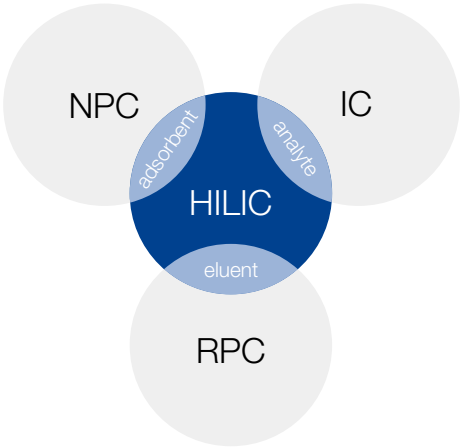
Technical data

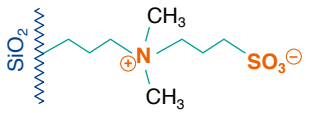
- Ammonium - sulfonic acid modified silica; pore size 110 Å; particle sizes 1.8, 3 and 5 µm; carbon content 7 %; pH stability 2–8.5

Recommended application

- Hydrophilic compounds such as organic polar acids and bases, polar natural compounds, nucleosides, oligonucleotides, amino acids, peptides, water soluble vitamins

Hydrophilic interaction chromatography





Especially for polar compounds reversed phase HPLC – the most common analytical method – is often limited. Here, hydrophilic stationary phases provide an additional tool for the separation of polar analytes in HPLC.

The expression HILIC (Hydrophilic Interaction Chromatography) was firstly published by Andrew Alpert in 1990 – since then it took quite some efforts to develop robust and reproducible hydrophilic HPLC phases for HILIC chromatography [7].

HILIC combines the characteristics of the 3 major methods in liquid chromatography – reversed phase (RPC), normal phase (NPC) and ion chromatography (IC):

- Stationary phases (adsorbents) are mostly polar modifications of silica or polymers (SiOH, NH_2, Diol, (zwitter) ions, …) – like in NPC.
- Mobile phases (eluents) are mixtures of aqueous buffer systems and organic modifier like acetonitrile or methanol - like in RPC.
- Fields of application include quite polar compounds as well as organic and inorganic ions – like in IC.

Summarized: “HILIC is NP chromatography of polar and ionic compounds under RP conditions.”

NUCLEODUR® HILIC is a special zwitterionic modified stationary phase based on ultra spherical NUCLEODUR® particles. The betaine character of the ammoniumsulfonic acid ligands results in total charge equalization and in an overall neutrally charged but highly polar surface

Retention characteristic

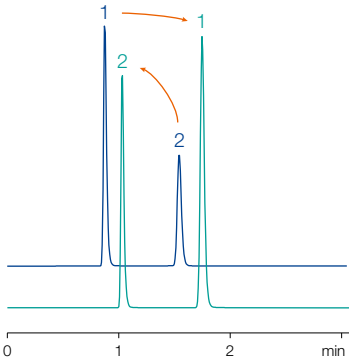
Commonly HILIC is described as partition chromatography or liquid-liquid extraction system between mobile and stationary phases. Versus a water-poor mobile phase a water-rich layer on the surface of the polar stationary phase is formed. Thus, a distribution of the analytes between these two layers will occur. Furthermore HILIC includes weak electrostatic mechanisms as well as hydrogen donor interactions between neutral polar molecules under high organic elution conditions. This distinguishes HILIC from ion exchange chromatography - main principle for HILIC separation is based on compound’s polarity and degree of solvation.

Separation of uracil and naphthalene

MN Appl. Nos. 122911 / 122912

Columns:	A) 125 x 4 mm NUCLEODUR® C_{18} Pyramid, 3 µm
	B) 125 x 4 mm NUCLEODUR® HILIC, 3 µm
Eluent:	acetonitrile – water (90:10, v/v))
Flow rate:	1.0 mL/min, temperature 25 °C
Detection:	UV, 254 nm

Peaks:
1. Uracil
2. Naphthalene



More polar compounds will have stronger interaction with the stationary aqueous layer than less polar compounds – resulting in a stronger retention. Nonpolar compounds exhibit faster elution profiles due to minor hydrophobic interactions. In the separation of uracil and naphthalene the elution order is quite often inverse on HILIC columns compared to RP columns.

Stability features

Due to an advanced and unique surface modification procedure (pat. pend.) NUCLEODUR® HILIC columns provide short equilibration times – after just 20 min equilibration already the 2nd injection shows stable and reproducible results.

Beyond this, NUCLEODUR® HILIC columns are characterized by an outstanding column life time - even after nearly 800 runs the columns show no loss of pristine performance - peak shape and retention are still immaculate. Due to its high loading capacity NUCLEODUR® HILIC is absolutely suitable for preparative and semi-preparative applications.

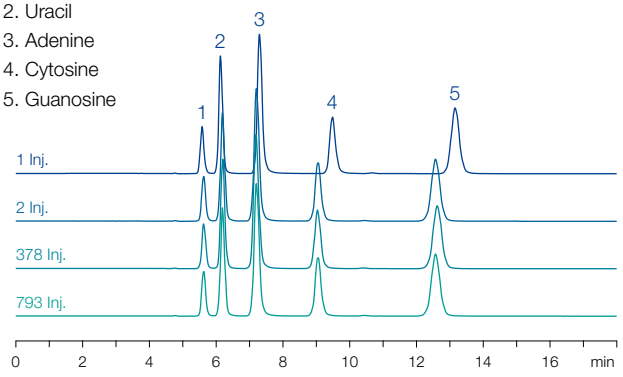
Overall NUCLEODUR® HILIC provides excellent chromatographic features and is hereby the perfect choice for separation of polar or charged compounds.

Stability and equilibration

MN Appl. No. 123100

Column:	250 x 4 mm NUCLEODUR® HILIC, 5 µm
Eluent:	CH_3CN – 5 mmol/L ammonium acetate (80:20, v/v)
Flow rate:	0.6 mL/min
Temperature:	25 °C
Detection:	UV, 254 nm

Peaks:
1. Thymine
2. Uracil
3. Adenine
4. Cytosine
5. Guanosine



Ordering information

Eluent in column acetonitrile – water (80:20, v/v)

ID	Length → 30 mm	50 mm	75 mm	100 mm	125 mm	150 mm	250 mm
NUCLEODUR® HILIC, 1.8 µm particle size 1.8 µm · UHPLC							
Analytical EC columns							
2 mm	760521.20	760523.20	760525.20	760526.20		760528.20	
3 mm	760521.30	760523.30		760526.30			
4 mm	760521.40	760523.40		760526.40			
4.6 mm	760521.46	760523.46		760526.46			
EC guard columns*		4 x 2 mm: 761960.20		4 x 3 mm: 761960.30			
NUCLEODUR® HILIC, 3 µm particle size 3 µm							
Analytical EC columns							
2 mm		760532.20		760534.20	760531.20	760533.20	760530.20
3 mm		760532.30		760534.30	760531.30	760533.30	760530.30
4 mm		760532.40		760534.40	760531.40	760533.40	760530.40
4.6 mm		760532.46		760534.46	760531.46	760533.46	760530.46
EC guard columns*		4 x 2 mm: 761961.20		4 x 3 mm: 761961.30			
NUCLEODUR® HILIC, 5 µm particle size 5 µm							
Analytical EC columns							
2 mm		760552.20		760554.20	760551.20	760553.20	760550.20
3 mm		760552.30		760554.30	760551.30	760553.30	760550.30
4 mm		760552.40		760554.40	760551.40	760553.40	760550.40
4.6 mm		760552.46		760554.46	760551.46	760553.46	760550.46
EC guard columns*		4 x 2 mm: 761962.20		4 x 3 mm: 761962.30			

Guard column system

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966

For details of our column systems see page 250.

NUCLEODUR® CN / CN-RP cyano-modified high purity silica phase · USP L10

Key feature

- High retention capacity especially for very polar and unsaturated compounds
- Multi-mode column (RP and NP) widens scope of selectivity
- Stable against hydrolysis at low pH (working range pH 1–8)

Technical data

- Cyanopropyl-modified high purity silica; pore size 110 Å; particle sizes 3 µm and 5 µm; carbon content 7 %; special endcapping
- High reproducibility from lot to lot; different retention characteristics in comparison to C_8 and C_{18}

Recommended application

- Tricyclic antidepressants, steroids, organic acids

Alternative bonded-phase functionality

In reversed phase HPLC it is fairly common to start with C_{18} or C_8 columns, if new methods have to be developed. However, superior polarity and selectivity properties often required for more sophisticated separations, are not always sufficiently provided by classical RP phases, which are usually characterized by a hydrophobic layer of monomeric or polymeric bonded alkylsilanes.

One approach to improve the resolution of compounds poorly separated on nonpolar stationary phases, is to change bonded-phase functionality.

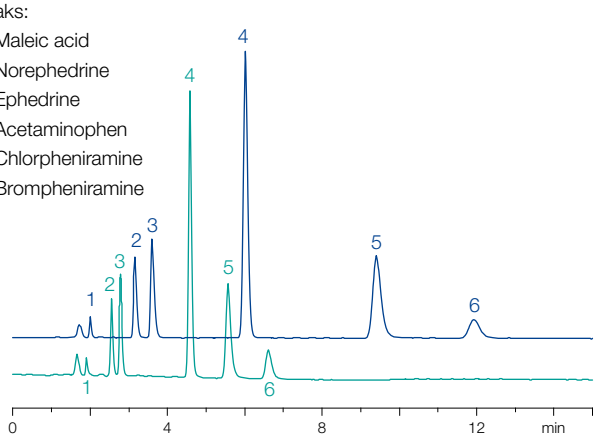
The fully endcapped and highly reproducible NUCLEODUR® 100-5 CN-RP phase has cyanopropyl groups on the surface able to generate a clearly recognizable different retention behavior compared to purely alkyl-functionalized surface modifications (see figure below).

Separation of cold medicine ingredients on two different NUCLEODUR® phases

MN Appl. No. 119340

Columns:	250 x 4 mm NUCLEODUR® 100-5 C_{18} ec
	250 x 4 mm NUCLEODUR® 100-5 CN-RP
Eluent:	acetonitrile – 100 mmol/L sodium citrate pH 2.5 (15:85, v/v)
Flow rate:	1.0 mL/min, temperature 25 °C
Detection:	UV, 254 nm, injection 10 µL

Peaks:
1. Maleic acid
2. Norephedrine
3. Ephedrine
4. Acetaminophen
5. Chlorpheniramine
6. Brompheniramine



The polarity of NUCLEODUR® 100-5 CN-RP can be classified as intermediate based on multiple retention mechanisms such as dipole-dipole, π-π, and also hydrophobic interactions [8]. Therefore, this phase shows a distinct selectivity for polar organic compounds as well as for molecules containing π electron systems (e.g., analytes with double bonds, tricyclic antidepressants) [9].

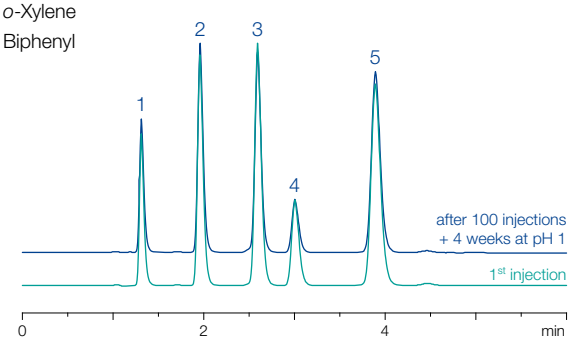
Short-chain bonded phases are sometimes suspected of revealing shortcomings in stability towards hydrolysis at low pH [10]. Application 119350 shows that even after 100 sample injections and four weeks storage at pH 1 (blue curve), neither a considerable shift in retention, nor a visible change in peak symmetry could be noticed (green curve = new column)

Stability of NUCLEODUR® CN-RP at pH 1

MN Appl. No. 119350

Columns:	125 x 4 mm NUCLEODUR® 100-5 CN-RP
Eluent:	acetonitrile – water, 2 % TFA pH 1 (50:50, v/v)
Flow rate:	1.0 mL/min
Temperature:	25 °C
Detection:	UV, 254 nm
Injection:	5 µL

Peaks:
1. Benzamide
2. Dimethyl phthalate
3. Phenetole
4. *o*-Xylene
5. Biphenyl



Multi-mode columns

Due to its polarity the cyano phase can also be run in normal phase mode. NUCLEODUR® CN columns for NP applications are shipped in *n*-heptane. The change in selectivity and order of elution for a mixture of various steroids in NP and RP mode is displayed below. The high coverage combined with a thorough endcapping makes NUCLEODUR® 100-5 CN-RP suitable for separation of ionizable compounds such as basic drugs.

Separation of steroids in normal phase and reversed phase mode

MN Appl. Nos. 119271 / 119272

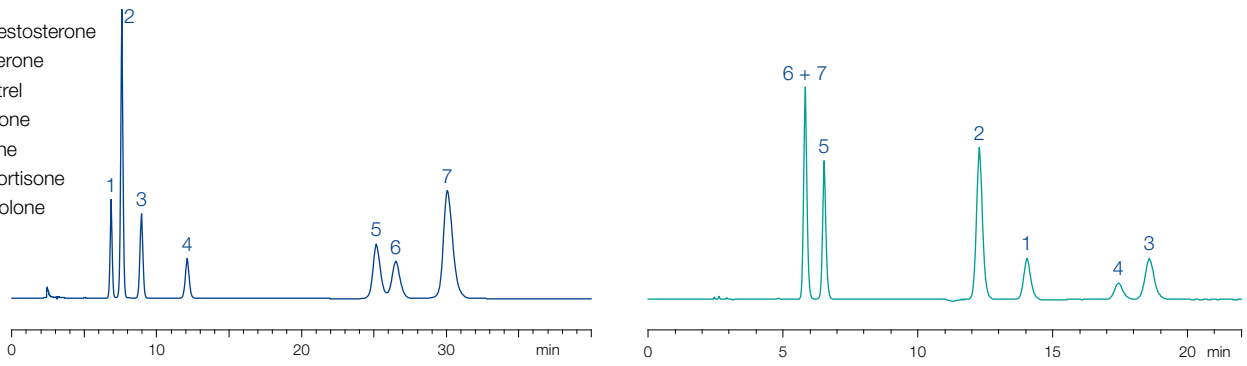
Normal phase mode

Column: 250 x 4 mm NUCLEODUR® 100-5 CN
Eluent: *n*-heptane – 2-propanol (90:10, v/v)
Flow rate: 1.0 mL/min, temperature 25 °C
Detection: UV, 254 nm, injection 10 µL

Reversed phase mode

Column: 250 x 4 mm NUCLEODUR® 100-5 CN-RP
Eluent: acetonitrile – water (25:75, v/v)
other conditions as for normal phase mode

Peaks:
1. Methyltestosterone
2. Testosterone
3. Norgestrel
4. Medrysone
5. Cortisone
6. Hydrocortisone
7. Prednisolone



Ordering information

	ID	Length → 50 mm	125 mm	150 mm	250 mm
NUCLEODUR® 100-3 CN-RP particle size 3 µm; eluent in column acetonitrile – water					
Analytical EC columns					
	2 mm	760159.20	760157.20		
	3 mm		760157.30		
	4 mm			760156.40	
	4.6 mm			760156.46	
EC guard columns*		4 x 2 mm: 761941.20		4 x 3 mm: 761941.30	
NUCLEODUR® 100-5 CN-RP particle size 5 µm; eluent in column acetonitrile – water					
Analytical EC columns					
	4 mm		760153.40		760152.40
	4.6 mm		760153.46	760154.46	760152.46
EC guard columns*				4 x 3 mm: 761944.30	
NUCLEODUR® 100-5 CN particle size 5 µm; eluent in column *n*-heptane					
Analytical EC columns					
	4 mm		760151.40	760149.40	760150.40
	4.6 mm		760151.46	760149.46	760150.46
EC guard columns*				4 x 3 mm: 761943.30	
EC columns in packs of 1, guard columns in packs of 3.					

Guard column system

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966

For details of our column systems see page 250.

NUCLEODUR® columns

NUCLEODUR® NH_2 / NH_2-RP amino-modified high purity silica · USP L8

Key feature

- Multi-mode columns (for RP, NP and IC)
- Stable against hydrolysis at low pH (working range pH 2–8), 100 % stable in water; suitable for LC/MS
- Widens scope of analytical HPLC into the polar range

Technical data

- Aminopropyl modified high purity silica; pore size 110 Å; particle sizes 3, 5 and 7 µm; carbon content 2.5 %; not endcapped

Recommended application

- Polar compounds under RP conditions (sugars, DNA bases), hydrocarbons under NP conditions

- Normal phase chromatography (NP) with hexane, dichloromethane or 2-propanol as mobile phase for polar compounds such as substituted anilines, esters, chlorinated pesticides
- Reversed phase chromatography (RP) of polar compounds in aqueous-organic eluent systems
- Ion exchange chromatography of anions and organic acids using conventional buffers and organic modifiers

Some compounds, especially polar substances, cannot be sufficiently resolved on C_{18} phases. Polar-modified silica phases offer alternative selectivities thus expanding the spectrum of analytical HPLC into the polar range.

Multi-mode columns

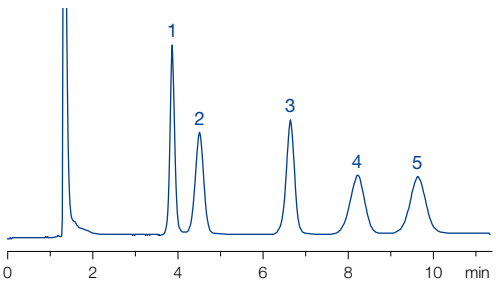
Besides cyano modifications, amino modifications belong to the most frequently used polar silica phases – both feature the important advantage, that they can be run in the RP mode using aqueous-organic eluent mixtures as well as in the NP mode, e.g., with hexane as mobile phase.

Reversed phase separation of sugars

MN Appl. No. 122160

Column: 250 x 4 mm NUCLEODUR® 100-5 NH_2-RP
Eluent: acetonitrile – water (79:21, v/v)
Flow rate: 2 mL/min
Detection: RI

Peaks:
1. Fructose
2. Glucose
3. Saccharose
4. Maltose
5. Lactose



NUCLEODUR® NH_2, too, belongs to the so-called multimode columns. It can be used for RP chromatography of polar compounds such as sugars in aqueous-organic eluent systems, for NP chromatography of substituted aromatics or chlorinated pesticides with organic mobile phases such as hexane, dichloromethane or 2-propanol, but also for ion exchange chromatography of anions and organic acids using conventional buffers and organic modifiers.

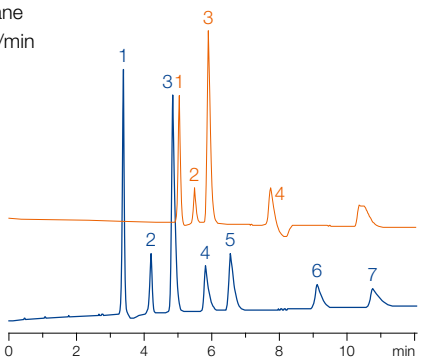
Main field of application of NUCLEODUR® NH_2 is the separation of simple and complex sugars, sugar alcohols and other hydroxy compounds under RP conditions as well as hydrocarbons under NP conditions.

Normal phase separation of middle distillates in accordance with DIN EN 12916

MN Appl. No. 122180

Columns: A) 250 x 4 mm NUCLEODUR® 100-5 NH_2
B) conventional aminopropyl phase
Eluent: heptane
Flow rate: 1 mL/min
Detection: RI

Peaks:
1. Cyclohexane
2. 1-Phenyldodecane
3. 1,2-Dimethylbenzene
4. Hexamethylbenzene
5. Naphthalene
6. Dibenzothiophene
7. 9-Methylanthracene



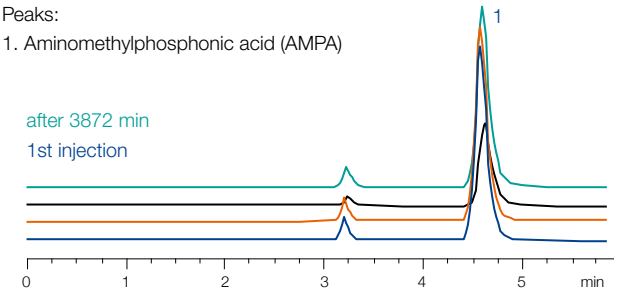
Due to the special method of surface modification NUCLEODUR® NH_2 features a pronounced stability at higher as well as at lower pH values. The following figure shows, that even after several days of exposure of the column material at pH 1.75 good separation efficiency and peak symmetry are maintained. The resulting high column life allows cost reduction due to lower column consumption.

This example shows the enhanced pH stability of NUCLEODUR® NH_2 and the outstanding suitability for the separation of total herbicides (AMPA, glyphosate, glufonisate, …) - see application 122190 in our online data base at *www.mn-net.com/apps*.

Hydrolytical resistance of NUCLEODUR® NH_2-RP

Column: 250 x 4 mm NUCLEODUR® 100-5 NH_2-RP
Eluent: acetonitrile – 50 mmol/L KH_2PO_4, pH 1.75 (50:50, v/v)
Flow rate: 0.6 mL/min
Detection: UV, 254 nm

Peaks:
1. Aminomethylphosphonic acid (AMPA)

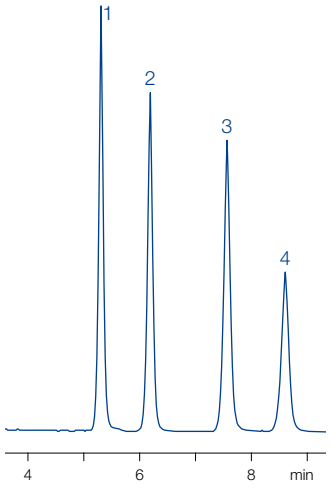


Separation of DNA bases

MN Appl. No. 122170

Column: 250 x 4 mm NUCLEODUR® 100-5 NH_2-RP
Eluent: acetonitrile – water (80:20, v/v)
Flow rate: 0.6 mL/min
Temperature: 35 °C
Pressure: 30 bar
Detection: UV, 254 nm

Peaks:
1. Thymine
2. Uracil
3. Cytosine
4. Adenine



Based on superspherical NUCLEODUR® this phase features a high pressure stability, which makes it the perfect choice for preparative separations as well as for LC/MS. Additionally, the high batch-to-batch reproducibility of NUCLEODUR® NH_2 enables reliable analyses especially for routine work.

Ordering information

ID	Length → 100 mm	125 mm	150 mm	250 mm
NUCLEODUR® 100-3 NH_2-RP particle size 3 µm; eluent in column acetonitrile – water				
Analytical EC columns				
2 mm	760740.20	760741.20		
4.6 mm			760742.46	760739.46
EC guard columns*	4 x 2 mm: 761951.20		4 x 3 mm: 761951.30	
NUCLEODUR® 100-5 NH_2-RP particle size 5 µm; eluent in column acetonitrile – water				
Analytical EC columns				
2 mm		760730.20		760732.20
3 mm		760730.30		760732.30
4 mm		760730.40		760732.40
4.6 mm		760730.46	760731.46	760732.46
EC guard columns*	4 x 2 mm: 761953.20		4 x 3 mm: 761953.30	
NUCLEODUR® 100-5 NH_2 particle size 5 µm; eluent in column *n*-heptane				
Analytical EC columns				
4 mm		760720.40		760722.40
4.6 mm		760720.46	760721.46	760722.46
EC guard columns*			4 x 3 mm: 761952.30	

EC columns in packs of 1, guard columns in packs of 3.

Guard column system

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966

For details of our column systems see page 250.

NUCLEODUR® columns

NUCLEODUR® SiOH unmodified silica for normal phase · USP L3

Key feature

- Totally spherical high purity silica
- Pressure stable up to 600 bar
- Suitable for analytical and preparative separation of polar and midpolar compounds

Technical data

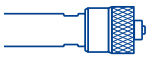
- Unmodified high purity silica; pore size 110 Å; particle sizes 3 to 50 µm; pore volume 0.9 mL/g; surface area (BET) 340 m^2/g; pH stability 2–8; metal content < 10 ppm (see page 150)

Recommended application

- Polar and midpolar compounds under normal phase conditions

Ordering information

Eluent in column *n*-heptane

	ID	Length → 50 mm	125 mm	150 mm	250 mm
NUCLEODUR® 100-3 particle size 3 µm					
Analytical EC columns					
	4.6 mm	760170.46		760172.46	760173.46
EC guard columns*				4 x 3 mm: 761966.30	
NUCLEODUR® 100-5 particle size 5 µm					
Analytical EC columns					
	4 mm				760007.40
	4.6 mm	760023.46		760012.46	760007.46
EC guard columns*				4 x 3 mm: 761967.30	
Preparative VarioPrep columns					
	10 mm	762077.100	762078.100		762007.100
	21 mm	762077.210	762078.210		762007.210
	40 mm			762075.400	762007.400
VP guard columns *		10 x 8 mm: 762094.80		10 x 16 mm: 762094.160	
		15 x 32 mm: 762330.320			
EC and VarioPrep columns in packs of 1, guard columns see below.					

Guard column systems

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966
Guard columns for VarioPrep columns with ID		**8, 10 mm**	**16, 21 mm**	**32, 40 mm**	**≥ 50 mm**	
** VP guard columns (pack of)	VP	10/8 (2)	10/16 (2)	15/32 (1)	15/50 (1)	
VP guard column holder		718251	718256	718253	718255	

For details of our column systems see page 250.
Unmodified NUCLEODUR® bulk material in 10–50 µm for self-packing of preparative columns see page 256.

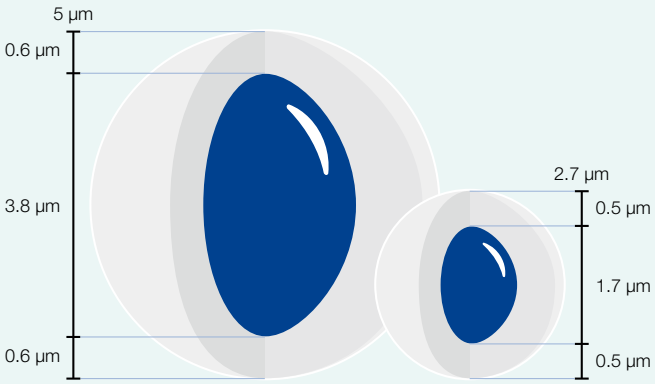
MACHEREY-NAGEL
your partner in HPLC · also online

Besides to this catalog our website provides useful information

- Applications
  Database without registration, with more than 3000 free chromatography applications for your separation task.
- Instruction manuals
  General advises for column care and individual column cleaning are available in the attached instruction manual or online.
- HPLC troubleshooting
  Sometimes during chromatographic separation unexpected effects occur. We give advise of possible reasons and how to avoid or remedy these.
- Flyers, brochures, catalogs
  Our product information is available online as PDF file at any time.

NUCLEOSHELL® core-shell silica for HPLC

Core-shell technology

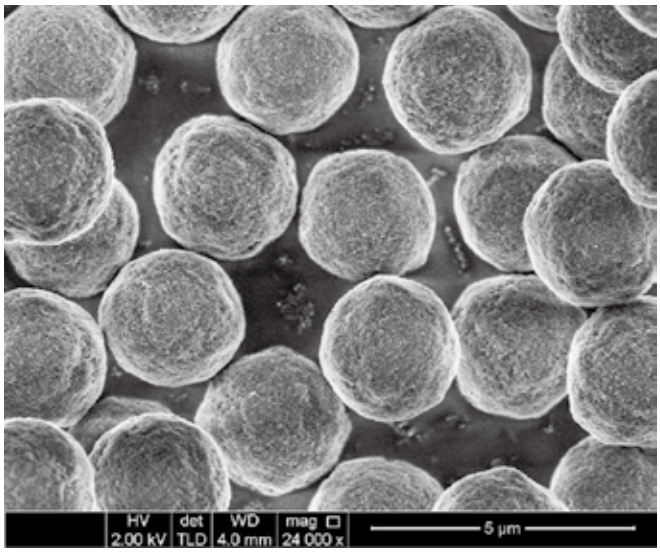


Key feature

- Solid core of silicon dioxide, homogeneous shell of porous silica
- Highest efficiency compared to traditional totally porous materials
- Pore size 90 Å; particle size 2.7 µm (core 1.7 µm) and 5 µm (core 3.8 µm); specific surface 130 (2.7 µm) and 90 (5 µm) m^2/g lower back pressure enables use on conventional LC systems
- Pressure stability 600 bar

Demands on HPLC separations are constantly increasing with respect to separation efficiency, detection limits, and the time requirements for each analysis.

Several approaches have been made to achieve fast separations without losing chromatographic performance. HPLC columns packed with particles < 2 µm show very high efficiencies (plates/meter) and allow the use of smaller column sizes with the positive side effect of significant solvent saving. However they generate a high back pressure of the mobile phase during column runs which requires specifically designed equipment.



Electron microscopic image of NUCLEOSHELL®

NUCLEOSHELL® silica particles consist of a non-porous solid core of 1.7 µm diameter and a porous outer shell of 0.5 µm thickness. Accordingly the total diameter of the particle is 2.7 µm.

Utilizing a proprietary process of synthesis, NUCLEOSHELL® particles exhibit a distinct narrow particle size distribution (d90/d10 ~ 1.1). Columns packed with NUCLEOSHELL core shell particles feature exceptional separation efficiencies with theoretical plate numbers easily comparable to totally porous sub 2 micron particles.

$$R = \frac{\sqrt{N}}{4}\left(\frac{\alpha - 1}{\alpha}\right)\left(\frac{k'_i}{k'_i + 1}\right)$$

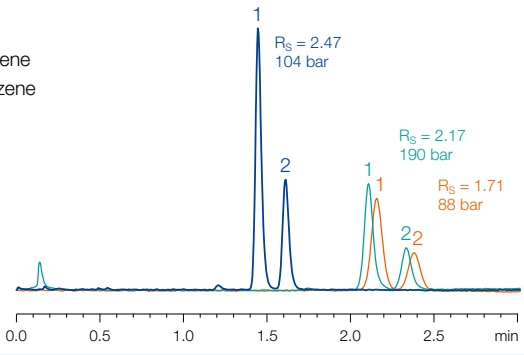
R = resolution, α = selectivity (separation factor), k_i' = retention
N = plate number with $N \propto 1/d_P$, d_P = particle diameter

Resolution R_S as function of particle size

MN Appl. No. 125270

Columns:	50 x 4 mm NUCLEOSHELL® RP 18, 2.7 µm NUCLEODUR® C_{18} Gravity, 3 µm NUCLEODUR® C_{18} Gravity, 1.8 µm
Eluent:	acetonitrile – water (60:40, v/v)
Flow rate:	1 mL/min
Temperature:	25 °C
Detection:	UV, 254 nm

Peaks:
1. Naphthalene
2. Ethylbenzene

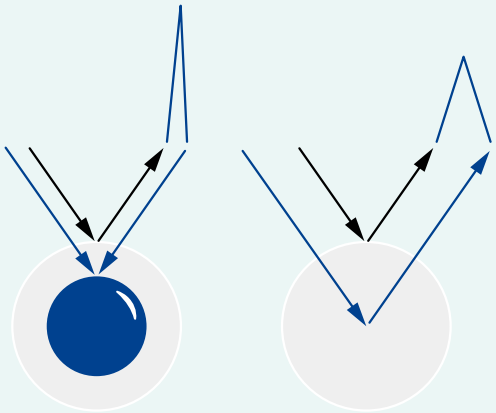


Theoretical column efficiency (optimal conditions)

Silica	d_p [µm]	L [m]	HETP [µm]	Efficiency [plates/m]	L [mm]	N	R_s	Analysis time
NUCLEOSHELL®	2.7	1	4	250 000	100	25 000	112 %	40 %
	5	1	6.5	154 000	150	23 000	115 %	60 %
NUCLEODUR®	1.8	1	4.5	222 222	100	22 000	105 %	40 %
	3	1	7.5	133 333	150	20 000	100 %	60 %
	5	1	12.5	80 000	250	20 000	100 %	100 %

Benefits of core-shell technology

Core-shell particles vs. totally porous silica



Short diffusion paths

- Fast mass transfer (term C of Van Deemter equation)
- High flow velocity without peak broadening for fast LC

Narrow particle size distribution (d_{90}/d_{10} ~ 1.1)

- Stable packing

High heat transfer

- Minimized influence of frictional heat
- Efficiency of NUCLEOSHELL® ~ 250 000 m^{-1} (HETP ~ 4 µm)

With conventional fully porous particles the mass transfer between stationary and mobile phase usually results in peak broadening at higher flow rates (C-term in van Deemter equation). The short diffusion paths in the core-shell particles reduce the dwell time of the analyte molecules in the stationary phase, so that even at high flow velocities of the mobile phase, optimal separation results can be obtained.

The van Deemter plots demonstrate how efficiency is affected by flow rate.

In comparison with fully porous silicas, core-shell particles from various manufacturers maintain the efficiency optimum (max. plates/m) over a long range of increasing linear mobile phase velocity.

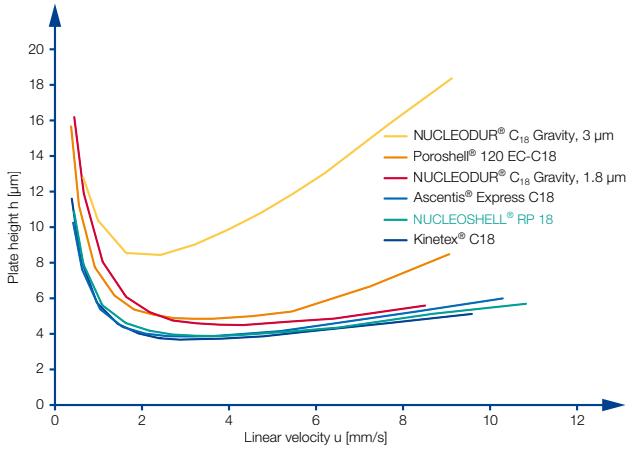
$$H = A + \frac{B}{u} + C \cdot u$$

A term = eddy-diffusion, B term = longitudinal diffusion coefficient, C term = mass transfer coefficient

Van Deemter curves

MN Appl. No. 125500

Column: 50 x 4.6 mm
Eluent: $CH_3CN – H_2O$ (70:30, v/v)
Temperature: 25 °C
Sample: Acenaphthene



NUCLEOSHELL® core-shell silica for HPLC

In direct comparison with conventional sub 2 micron phases, NUCLEOSHELL® columns only generate about 60 % of the back pressure and can be operated with the majority of conventional HPLC systems. In order to develop the maximum performance of NUCLEOSHELL® columns, we recommend reducing extra column voids by using suitable capillaries (< 0.15 mm inner diameter) and specially adapted detector cells. Moreover detector settings should be optimized by increasing the measuring rate or by decrease of the time constant.

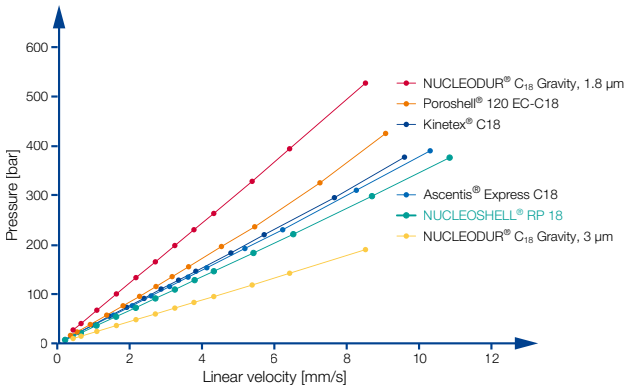
$$\Delta_P = \frac{\Phi \cdot L_C \cdot \eta \cdot u}{d_P^2}$$

Δ_P = pressure drop, Φ = flow resistance (nondimensional), LC = column length, η = viscosity, u = linear velocity, d_P = particle diameter

Pressure drop

MN Appl. No. 125510

Column: 50 x 4.6 mm
Eluent: $CH_3CN - H_2O$ (70:30, v/v)
Temperature: 25 °C



Core-shell particle technology from MACHEREY-NAGEL is an alternate route to gain highest column efficiency and resolution in HPLC at short run time, but with moderate back pressure.

Features of NUCLEOSHELL® particles

A criterion for the long-term stability of the column at pH extremes is the percentage decrease of initial retention and initial plates, respectively.

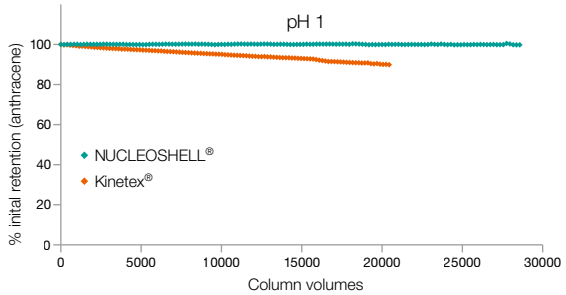
The following figure shows a column stability test of NUCLEOSHELL® RP 18 at mobile phase levels pH 1 and pH 10 compared with three competing phases.

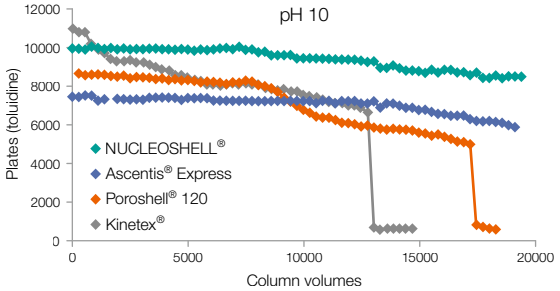
Stability under acidic and basic conditions

MN Appl. Nos. 125520 / 125530

Columns: 50 x 4.6 mm NUCLEOSHELL® RP 18, 2.7 µm
50 x 4.6 mm Kinetex® 2.6 µm C18
Eluent: acetonitrile – 1 % TFA in water, pH 1 (50:50, v/v)
Flow rate: 1.3 mL/min
Temperature: 80 °C
Detection: UV, 254 nm
Analyt: anthracene

Columns: 50 x 4.6 mm NUCLEOSHELL® RP 18, 2.7 µm
50 x 4.6 mm Ascentis® Express C18, 2.7 µm
50 x 4.6 mm Poroshell® 120 EC-C18
50 x 4.6 mm Kinetex® 2,6 µm C18
Eluent: 20 mmol/L Na borate – 10 mmol/L NaOH – methanol, pH 10 (21:49:30, v/v/v)
Flow rate: 1.5 mL/min
Temperature: 40 °C
Detection: UV, 220 nm
Analyt: toluidine





Columns can be operated at elevated temperatures without loss in retention, efficiency or peak symmetry.

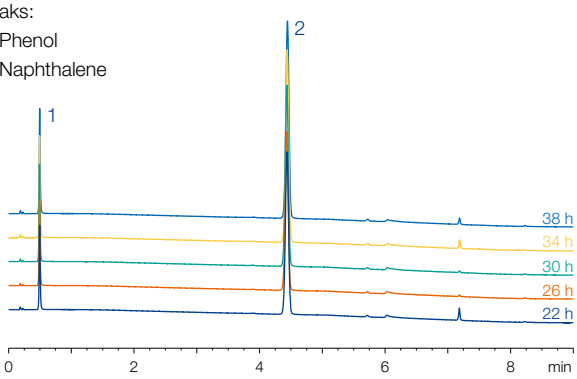
Temperature stability

MN Appl. No. 125400

Stability test:

Column:	50 x 2 mm NUCLEOSHELL® RP 18, 2.7 µm
Eluent:	A) 10 mmol/L ammonium formate – methanol (9:1, v/v) + 120 µL formic acid, ~ pH 4 B) 10 mmol/L ammonium formate – methanol (1:9, v/v) + 120 µL formic acid, ~ pH 4 0–100 % B in 7 min
Flow rate:	0.5 mL/min,
Temperature:	100 °C
Detection:	UV, 220 nm

Peaks:
1. Phenol
2. Naphthalene



Efficiency test:

Eluent:	Acetonitrile – water (60:40, v/v)
Flow rate:	0.33 mL/min;
Temperature:	25 °C
Detection:	UV, 254 nm
Analyte:	Anthracene

	HETP [µm]	Asymmetry
Start (t = 0)	5.2	0.98
End (t = 40 h)	5.2	1.01

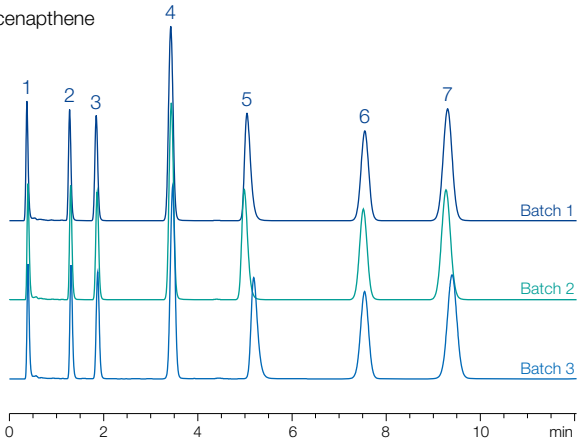
Uniformly shaped NUCLEOSHELL® particles combined with optimized bonding technology safeguard tightly packed columns for 100 % reproducible results.

Batch-to-batch reproducibility

MN Appl. No. 125410

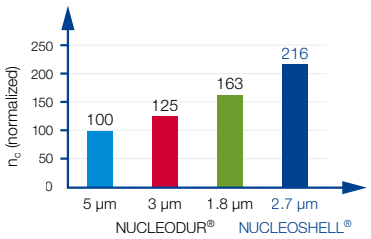
Column:	50 x 4 mm NUCLEOSHELL® RP 18, 2.7 µm
Eluent:	methanol – 25 mmol/L KH_2PO_4, pH 7 (70:30, v/v)
Flow rate:	1 mL/min
Temperature:	40 °C
Detection:	UV, 254 nm

Peaks:
1. Uracil
2. Toluene
3. Ethylbenzene
4. Acenapthene
5. Amitriptyline
6. *o*-Terphenyl
7. Triphenylene



Peak capacity

The peak capacity is a measure for the number of sample analytes that can be separated on HPLC columns per time unit. Narrow peaks increase the peak capacity and thus the efficiency of the analytical column.



Comparison of peak capacities

$$n_c = 1 + \left(\frac{t_g}{W}\right)$$

n_c: peak capacity
t_g: gradient time
W: peak width (at baseline)

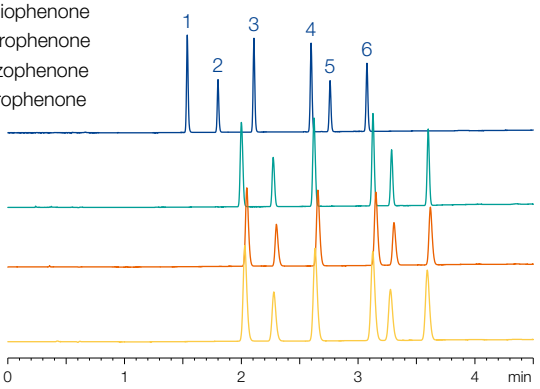
The example shows, that in comparison with totally porous NUCLEODUR® silica (1.8 µm) NUCLEOSHELL® provides 33 % higher peak capacity.

Peak capacity

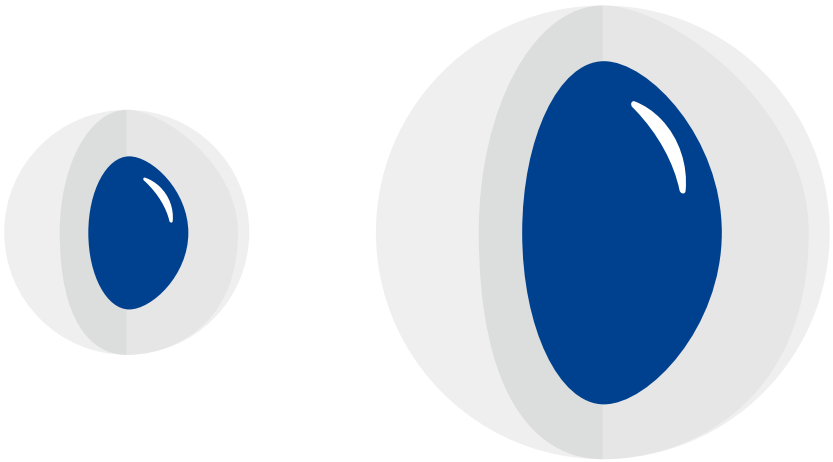
MN Appl. No. 125540

Columns:	100 x 4.6 mm each NUCLEOSHELL® RP 18, 2.7 µm NUCLEODUR® C_{18} Gravity, 1.8 µm NUCLEODUR® C_{18} Gravity, 3 µm NUCLEODUR® C_{18} Gravity, 5 µm
Eluent:	A) acetonitrile, B) water, 40–100 % A in 4 min
Flow rate:	1.5 mL/min
Temperature:	25 °C
Detection:	UV, 230 nm

Peaks:
1. Acetophenone
2. Benzoin
3. Propiophenone
4. Butyrophenone
5. Benzophenone
6. Valerophenone



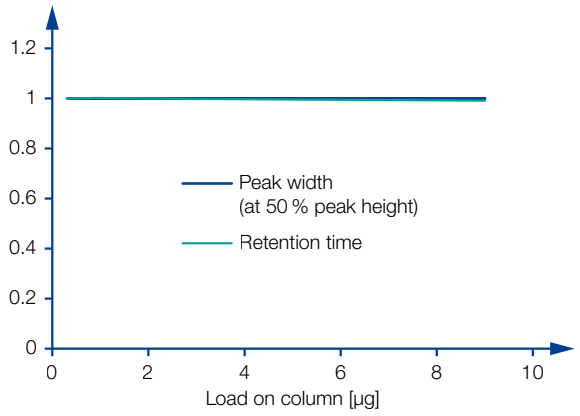
	Max. pressure [bar]	Resolution (4.5)
NUCLEOSHELL®, 2.7 µm	255	5.45
NUCLEODUR®, 1.8 µm	450	4.14
NUCLEODUR®, 3 µm	214	2.97
NUCLEODUR®, 5 µm	142	2.30



Loading capacity

NUCLEOSHELL® columns allow reliable quantification in a wide analytical detection range. Retention time and peak width at 50 % height remain constant with increasing columns load although core-shell particles are suspected of showing a slightly lower loading capacity compared to fully porous silica materials.

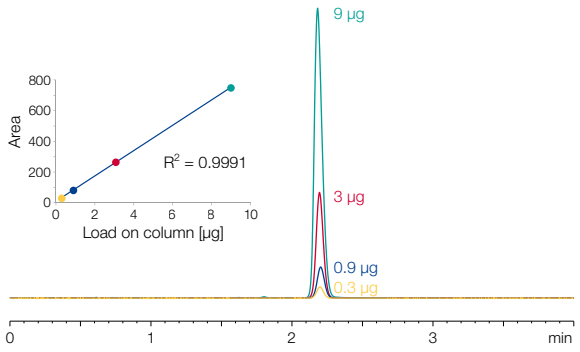
Normalized column parameters



Loading capacity

Column:	50 x 3 mm NUCLEOSHELL® RP 18, 2.7 µm
Eluent:	acetonitrile – 25 mmol/L KH_2PO_4, pH 3 (70:30, v/v)
Flow rate:	0.66 mL/min
Temperature:	30 °C
Detection:	UV, 285 nm

Peaks:
1. Valerophenone



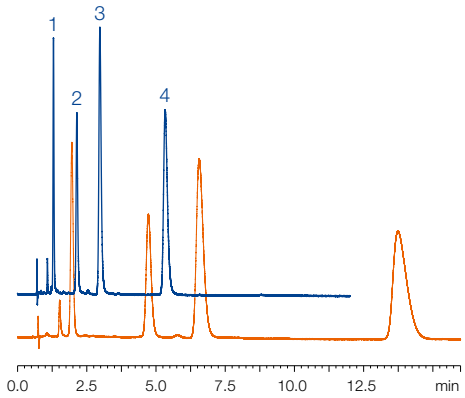
Method transfer of 5 µm particle columns

NUCLEOSHELL® is also available in 5 µm particle size to offer all benefits of core-shell technology to all applications which are bound to particle size.

Separation of cephalosporin antibiotics

MN Appl. No. 126630

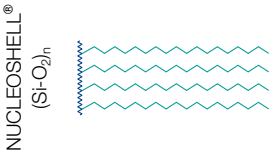
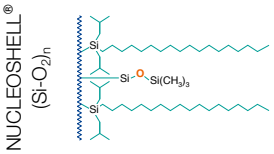
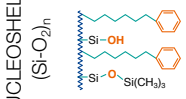
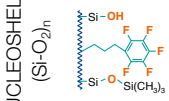
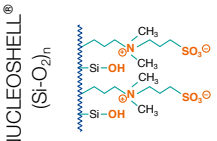
Comparison of 5 µm core-shell and totally porous phase	
Columns:	each 100 x 4.6 mm
	A) NUCLEOSHELL® RP 18plus, 5 µm
	B) NUCLEODUR® Gravity C_{18}, 5 µm
Eluent:	methanol – water + 0.1 % formic acid (35:65, v/v)
Flow rate:	1.3 mL/min
Pressure:	182 bar, 219 bar
Temperature:	25 °C
Detection:	UV, 254 nm
Injection:	4.0 µL



Peaks:	Ret. time [min]		Asymmetry (EP)		Plates (EP)	
	A	B	A	B	A	B
1 Cefotaxime	1.30	1.96	1.19	1.12	6800	2218
2 Cefoxitin	2.14	4.72	1.22	1.20	6599	3471
3 Cefamandole	2.97	6.57	1.24	1.25	6259	3367
4 Cefalotine	5.33	13.73	1.32	1.61	6948	3672

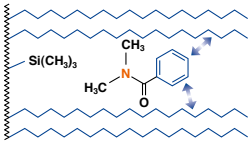
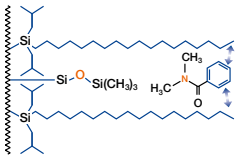
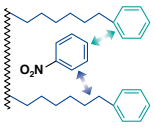
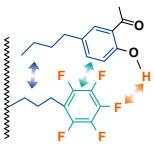
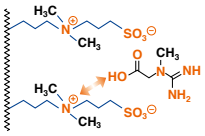
NUCLEOSHELL® phase overview

Overview of NUCLEOSHELL® HPLC phases

Phase	Specification	Page	Characteristic*	Stability	Structure
RP 18	octadecyl, multi-endcapping 7.8 % C (2.7 µm particles) 6.1 % C (5 µm particles) USP L1	200	A ●●●●● B ◖ C ●●◗	pH 1–11, suitable for LC/MS	
RP 18plus	octadecyl (monomeric), multi-endcapping 5.7 % C (2.7 µm particles) 4.4 % C (5 µm particles) USP L1	202	A ●●●● B ●●◖ C -	pH 2–9, suitable for LC/MS	
Phenyl-Hexyl	phenylhexyl, multi-endcapping 4.5 % C (2.7 µm particles) USP L11	204	A ●● B ●●● C ●	pH 1–10, suitable for LC/MS	
PFP	pentafluorophenyl, multi-endcapping ~ 3 % C (2.7 µm particles) USP L43	206	A ●● B ●●●● C ●●●●	pH 1–9, suitable for LC/MS	
HILIC	zwitterionic ammonium – sulfonic acid 1.3 % C (2.7 µm particles)	208	A ● B ●●●●● C -	pH 2–8.5, suitable for LC/MS	

* A = ● hydrophobic selectivity, B = ● polar / ionic selectivity, C = ● steric selectivity

NUCLEOSHELL® phase overview

Application	Similar phases**	Interactions · retention mechanism	
overall sophisticated analytical separations, e.g., analgesics, anti-inflammatory drugs, antidepressants; herbicides; phytopharmaceuticals; immunosuppressants	Kinetex® C18; Cortecs® C18; Raptor® C18; Accucore® C18; Ascentis® Express C18	hydrophobic (van der Waals interactions)	
overall sophisticated analytical separations, especially for polar compounds, e.g., pharmaceuticals like antibiotics, water-soluble vitamins, organic acids	Kinetex® XB-C18; Bonshell® ASB-C18; Raptor® ARC-C18;	hydrophobic (van der Waals interactions)	
aromatic and unsaturated compounds, polar compounds like pharmaceuticals, antibiotics	Ascentis® Express Phenyl-Hexyl; Kinetex® Phenyl-Hexyl; Accucore® Phenyl-Hexyl; Ultracore® Phenyl-Hexyl; Poroshell® Phenyl-Hexyl	π-π and hydrophobic	
aromatic and unsaturated compounds, phenols, halogenated hydrocarbons, isomers, polar compounds like pharmaceuticals, antibiotics	Kinetex® PFP; Ascentis® Express F5; Accucore® PFP	polar (H bond), dipole-dipole, π-π and hydrophobic	
hydrophilic compounds such as organic polar acids and bases, polar natural compounds	–	ionic / hydrophilic and electrostatic	

** phases which provide a similar selectivity based on chemical and physical properties

NUCLEOSHELL® columns

NUCLEOSHELL® RP 18 nonpolar high density phase · USP L1

Key feature

- Core-shell technology for fast and efficient HPLC
- Suitable for LC/MS and HPLC at pH extremes (pH 1–11)
- Superior base deactivation, ideal for method development

Technical data

- Octadecyl modification, multi-end-capped; pore size 90 Å, particle size 2.7 and 5 µm, carbon content 7.8 % for 2.7 µm, 6.1 % for 5 µm; pH stability 1–11; suitable for LC/MS

Recommended application

- Overall sophisticated analytical separations, e.g., analgesics, anti-inflammatory drugs, antidepressants; herbicides; phytopharmaceuticals; immunosuppressants

NUCLEOSHELL® RP 18 is based on core-shell silica. A unique derivatization process generates a homogeneous surface with a high density of bonded silanes. The following thorough endcapping suppresses any unwanted polar interactions between the silica surface and the sample, which makes NUCLEOSHELL® RP 18 particularly suitable for the separation of basic and other ionizable analytes. The extremely reduced silanol activity of the phase can be demonstrated by applying basic analytes, such as tricyclic antidepressants. The chromatogram below shows a sharp elution profile (superior resolution!) of these highly polar compounds with an excellent asymmetry value for amitriptyline of 1.12.

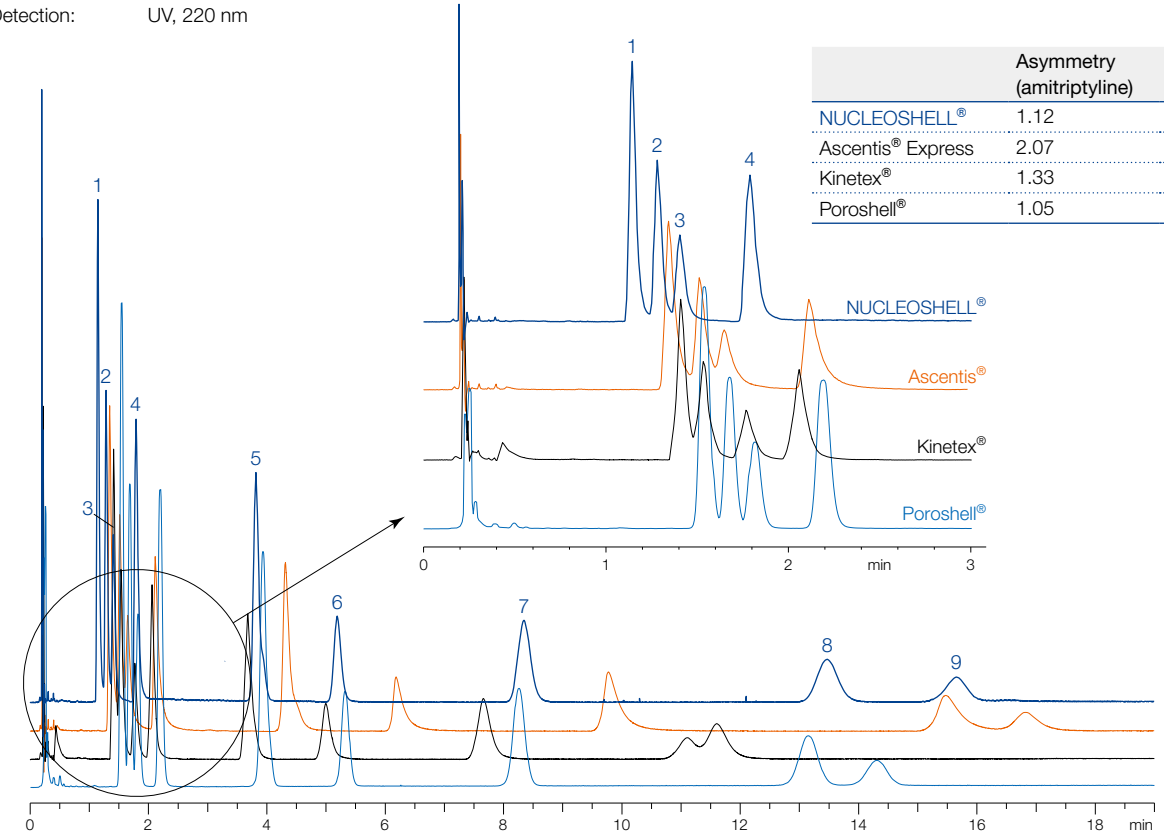
Tricyclic antidepressants · comparison of selectivity and resolution

MN Appl. No. 124960

Columns:	50 x 4.6 mm each NUCLEOSHELL® RP 18, 2.7 µm Ascentis® Express C18 Kinetex® 2.6 µm C18 Poroshell® 120 EC-C18
Eluent:	methanol – acetonitrile – 25 mmol/L KH_2PO_4, pH 7 (22.5:22.5:55, v/v/v)
Flow rate:	2 mL/min
Pressure:	224 bar, 239 bar, 248 bar, 212 bar
Temperature:	40 °C
Detection:	UV, 220 nm

Peaks:
1. Protriptyline
2. Desipramine
3. Maprotiline
4. Nortriptyline
5. Doxepin
6. Imipramine
7. Amitriptyline
8. Clomipramine
9. Trimipramine

	Asymmetry (amitriptyline)	Resolution (8, 9)
NUCLEOSHELL®	1.12	3.35
Ascentis® Express	2.07	1.91
Kinetex®	1.33	n.a.
Poroshell®	1.05	1.95



NUCLEOSHELL® RP 18 combines innovative silica technology and excellent surface deactivation, that outperforms conventional C_{18} silicas in terms of efficiency, resolution and speed.

Due to the applied core-shell particle design the back pressure at elevated flow rates remains at a moderate level and in many cases permits the use of existing HPLC equipment. NUCLEOSHELL® RP 18 with extended pH stability, low bleed characteristics in LC/MS applications, and overall robustness is an ideal tool for method development and routine analyses in modern HPLC.

The separation of 13 β-lactam antibiotics illustrates how time of analysis can be shortened to a fractional part by using core-shell particles without loss of resolution at moderate back pressure.

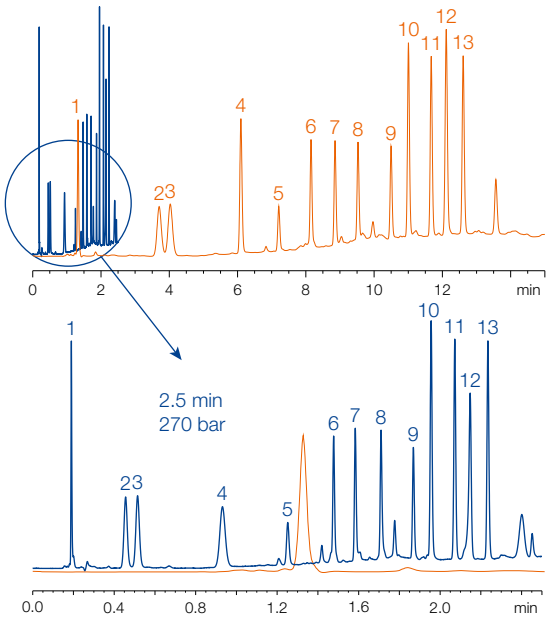
13 β-lactam antibiotics in less than 3 min

MN Appl. No. 124940

Columns:	50 x 4 mm NUCLEOSHELL® RP 18, 2.7 µm 150 x 4 mm NUCLEODUR® C_{18} Gravity, 5 µm
Eluent:	A) acetonitrile B) 20 mmol/L KH_2PO_4, pH 3.5 10 % A (0,5 min) → 50 % A in 1.5 min (0.5 min 50 % A) 10 % A (3 min) → 50 % A in 9 min (3 min 50 % A)
Flow rate:	2 mL/min, 1 mL/min
Pressure:	270 bar, 110 bar
Temperature:	25 °C
Detection:	UV, 220 nm

Peaks:

1. Amoxicillin
2. Ampicillin
3. Cephalexin
4. Cefotaxime
5. Cefoxitin
6. Cefamandole
7. Cephalothin
8. Piperacillin
9. Penicillin V
10. Oxacillin
11. Cloxacillin
12. Nafcillin
13. Dicloxacillin



Ordering information

Eluent in column acetonitrile – water

ID	Length → 50 mm	100 mm	150 mm	250 mm	EC guard columns*
NUCLEOSHELL® RP 18, 2.7 µm particle size 2.7 µm					
Analytical EC columns					
2 mm	763132.20	763134.20	763136.20		763138.20
3 mm	763132.30	763134.30	763136.30		763138.30
4 mm	763132.40	763134.40	763136.40		763138.30
4.6 mm	763132.46	763134.46	763136.46		763138.30
NUCLEOSHELL® RP 18, 5 µm particle size 5 µm					
Analytical EC columns					
2 mm	763152.20	763154.20	763156.20	763157.20	763158.20
3 mm	763152.30	763154.30	763156.30	763157.30	763158.30
4 mm	763152.40	763154.40	763156.40	763157.40	763158.30
4.6 mm	763152.46	763154.46	763156.46	763157.46	763158.30

EC columns in packs of 1, guard columns in packs of 3.

Guard column system

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966

For details of the EC column system please see page 250.

NUCLEOSHELL® RP 18plus C_{18} phase with polar selectivity · USP L1

Key feature

- Based on core-shell particle technology for fast and efficient HPLC
- Hydrophobic C_{18} phase with distinct polar selectivity, ideal for method development
- Excellent performance under highly aqueous conditions

Technical data

- Monomeric octadecyl modification, multi-endcapped; pore size 90 Å, available particle sizes 2.7 µm and 5 µm, carbon content 5.7 % for 2.7 µm, 4.4 % for 5 µm; pH stability 2–9; suitable for LC/MS

Recommended application

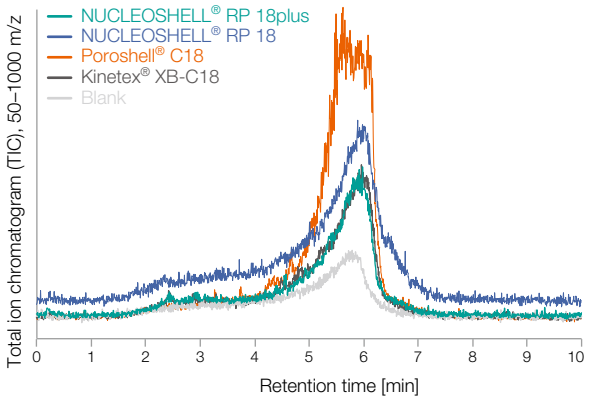
- Overall sophisticated analytical separations, especially for polar compounds, e.g., pharmaceuticals like antibiotics, water-soluble vitamins, organic acids

NUCLEOSHELL® RP 18plus is a C_{18} modified core-shell silica. Due to a monomeric bonding chemistry this HPLC phase offers hydrophobic characteristics with distinct polar selectivity. A special derivatization process generates a medium density of bonded silanes with reduced steric selectivity compared to NUCLEOSHELL® RP 18.

Bleeding characterisitics

MN Appl. No. 126640

Column:	50 x 2 mm NUCLEOSHELL® RP 18plus, 2.7 µm
Eluent:	A) 0.1 % formic acid in water B) 0.1 % formic acid in acetonitrile 95 % A → 5 % A in 4.5 min (0.5 min) → 95 % A in 0.5 min (4.5 min)
Flow rate:	0.5 mL/min
Temperature:	25 °C
Detection:	MS

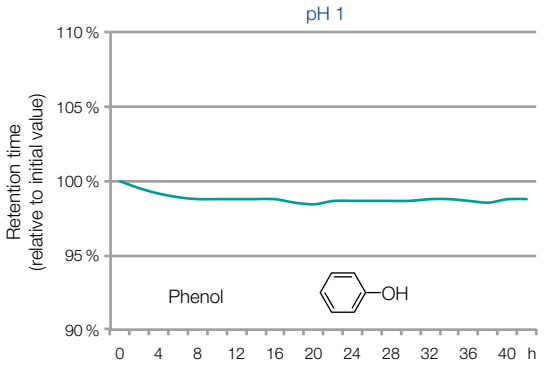


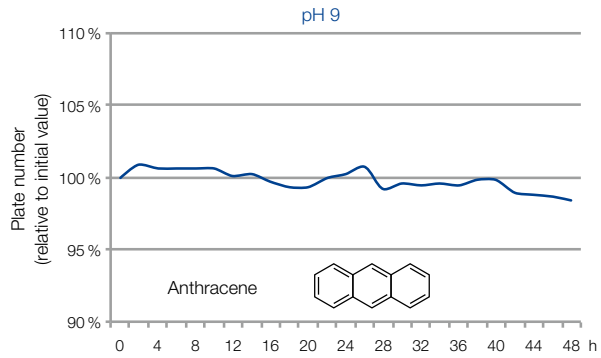
NUCLEOSHELL® RP 18plus combines superbly hydrophobic and polar selectivity – so it is a useful tool for method development in RP chromatography. Good pH stability and low bleeding characteristics make it ideal especially for LC/MS applications.

pH stability of NUCLEOSHELL® RP 18plus

MN Appl. No. 126650

Column:	100 x 4 mm NUCLEOSHELL® RP 18plus, 2.7 µm
Eluent pH 1:	1 % TFA in water - acetonitrile (50:50, v/v)
Eluent pH 9:	50 mmol/L triethylammonium acetate adjusted to pH 9
Flow rate:	for pH 1: 0.8 mL/min, for pH 9: 0.56 mL/min
Temperature:	for pH 1: 60 °C, for pH 9: 50 °C
Detection:	UV, 254 nm
Injection:	1 µL



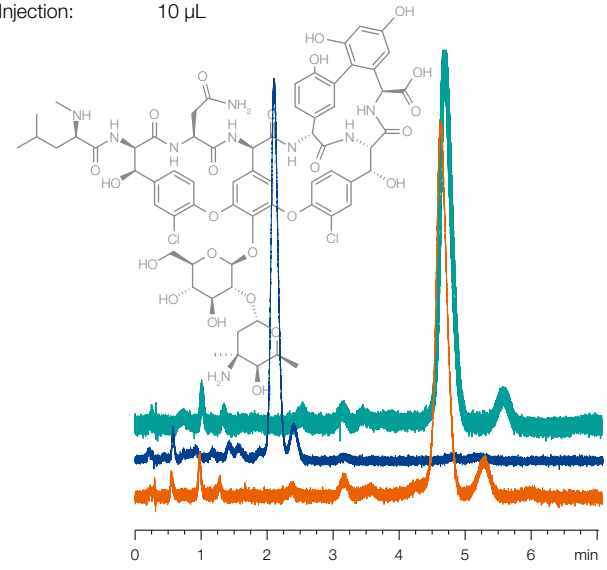


Also a comparison of retention of the glycopeptide antibiotic vancomycin on several octadecyl modified core-shell phases underlines the polar selectivity of NUCLEOSHELL® RP 18plus.

Polar selectivity shown for vancomycin

MN Appl. No. 126660

Columns:	50 x 3 mm each NUCLEOSHELL® RP 18plus, 2.7 µm NUCLEOSHELL® RP 18, 2.7 µm Kinetex® 2.6 µm C18
Eluent:	water – methanol – acetonitrile – glacial acetic acid (100:8:2:0.3, v/v/v/v) adjusted to pH 3.2 with sodium hydroxide solution
Flow rate:	0.9 mL/min
Temperature:	35 °C
Detection:	UV, 240 nm
Injection:	10 µL

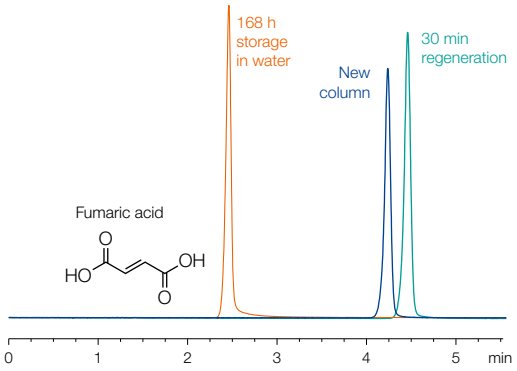


In addition NUCEOSHELL® RP 18plus provides a good stability under highly aqueous conditions. Even by long term usage or storage of the phase phase collapse and loss of retention are hardly observed. The original performance can be regained after a short regeneration procedure.

Phase collapse and regeneration

MN Appl. No. 126670

Column:	100 x 4 mm NUCLEOSHELL® RP 18plus, 2.7 µm
Eluent:	20 mmol/L KH_2PO_4, pH 2.6
Flow rate:	0.5 mL/min
Temperature:	20 °C
Detection:	UV, 215 nm
Injection:	0.5 µL



Ordering information

Eluent in column acetonitrile – water

	ID	Length → 50 mm	100 mm	150 mm	250 mm	EC guard columns*
NUCLEOSHELL® RP 18plus, 2.7 µm particle size 2.7 µm						
Analytical EC columns						
	2 mm	763232.20	763234.20	763236.20		763238.20
	3 mm	763232.30	763234.30	763236.30		763238.30
	4 mm	763232.40	763234.40	763236.40		763238.30
	4.6 mm	763232.46	763234.46	763236.46		763238.30
NUCLEOSHELL® RP 18plus, 5 µm particle size 5 µm						
Analytical EC columns						
	2 mm	763252.20	763254.20	763256.20	763257.20	763258.20
	3 mm	763252.30	763254.30	763256.30	763257.30	763258.30
	4 mm	763252.40	763254.40	763256.40	763257.40	763258.30
	4.6 mm	763252.46	763254.46	763256.46	763257.46	763258.30

EC columns in packs of 1, guard columns in packs of 3.

Guard column system

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966

For details of the EC column system please see page 250.

NUCLEOSHELL® columns

NUCLEOSHELL® Phenyl-Hexyl nonpolar high density phase · USP L11

Key feature
- Based on core-shell particle technology for fast and efficient HPLC
- Hydrophobic phase with alternative selectivity compared to classical C_{18} modifications
- Separation principle based on 2 retention mechanisms: π-π interactions and hydrophobic interactions

Technical data
- Phenyl-Hexyl modification, multi-end-capped; pore size 90 Å, particle size 2.7 µm; carbon content 4.5 %; pH stability 1–10; suitable for LC/MS

Recommended application
- Aromatic and unsaturated compounds, polar compounds like pharmaceuticals, antibiotics

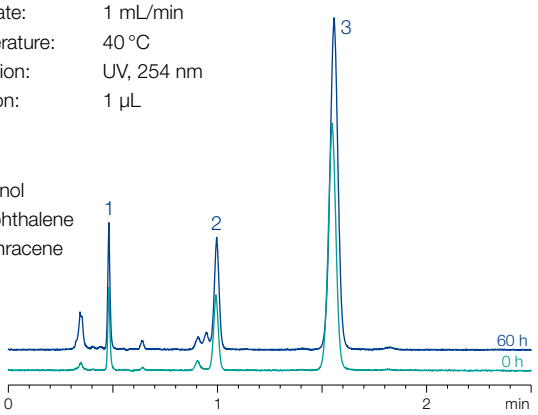
Phenyl-Hexyl modified phases offer an excellent separation efficiency especially for aromatic and unsaturated compounds with electron-withdrawing groups. The combination of hydrophobic and π-π interactions results in an alternative and interesting selectivity profile compared to C_{18} or C_8 modifications. NUCLEOSHELL® Phenyl-Hexyl is based on a unique surface bonding chemistry - therefore it is suitable for LC/MS due to low bleeding characteristics and offers high temperature stability and pH stability from 1 to 10.

Stability of NUCLEOSHELL® Phenyl-Hexyl at pH 10

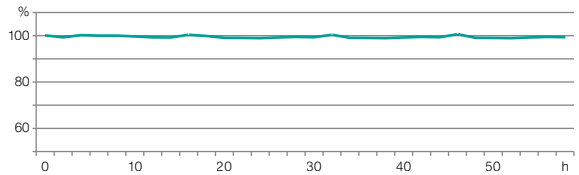
MN Appl. No. 126420

Column:	50 x 4 mm NUCLEOSHELL® Phenyl-Hexyl, 2.7 µm
Eluent:	acetonitrile – 50 mmol/L TEA pH 10 (60:40, v/v); pH of the mixture 10.4
Flow rate:	1 mL/min
Temperature:	40 °C
Detection:	UV, 254 nm
Injection:	1 µL

Peaks:
1. Phenol
2. Naphthalene
3. Anthracene



Relative plate numbers

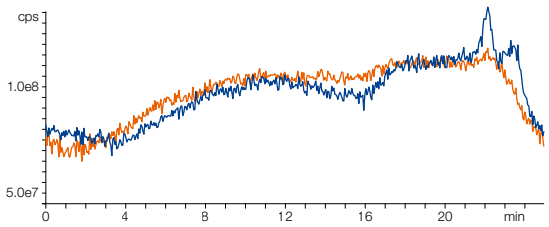


NUCLEOSHELL® Phenyl-Hexyl is a robust phase with an alternative RP selectivity for aromatic and unsaturated analytes compared to classical C_{18} / C_8 phases – it is an additional and useful tool for all chromatography users.

Bleeding characteristics of NUCLEOSHELL® Phenyl-Hexyl

MN Appl. No. 126400

Columns:	50 x 2 mm each NUCLEOSHELL® Phenyl-Hexyl, 2.7 µm Kinetex® Phenyl-Hexyl
Eluent:	A) acetonitrile, B) water 5–95 % A in 25 min
Flow rate:	0.2 mL/min
Temperature:	25 °C
Detection:	MS



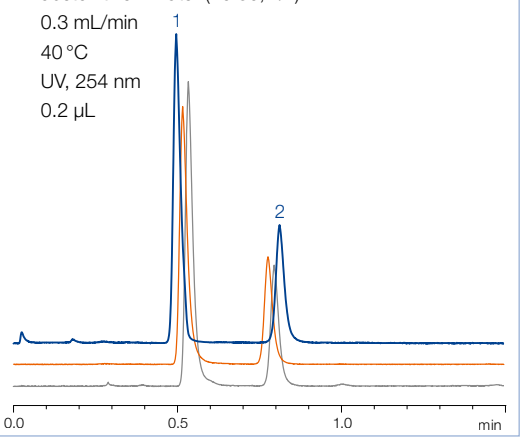
The pyridine-phenol test shows that NUCLEOSHELL® Phenyl-Hexyl provides a symmetrical peak for pyridine and higher resolution in comparison to other core-shell based Phenyl-Hexyl phases, which underlines the excellent base deactivation.

Pyridine-phenol test of NUCLEOSHELL® Phenyl-Hexyl

MN Appl. No. 126410

Columns:	50 x 2 mm each NUCLEOSHELL® Phenyl-Hexyl, 2.7 µm Kinetex® Phenyl-Hexyl Ascentis® Express Phenyl-Hexyl
Eluent:	acetonitrile – water (70:30, v/v)
Flow rate:	0.3 mL/min
Temperature:	40 °C
Detection:	UV, 254 nm
Injection:	0.2 µL

Peaks:
1. Pyridine
2. Phenol



Comparing the separation of sulfonamides on NUCLEODUR® Phenyl-Hexyl with different particle sizes

MN Appl. No. 125860

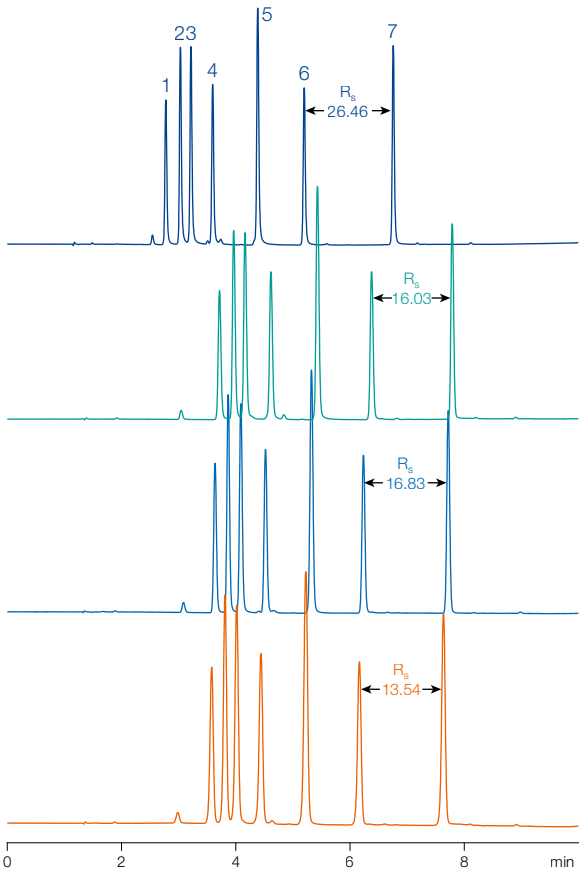
Columns: 150 x 3 mm each
NUCLEOSHELL® Phenyl-Hexyl, 2.7 µm
NUCLEODUR® Phenyl-Hexyl, 1.8 µm
NUCLEODUR® Phenyl-Hexyl, 3 µm
NUCLEODUR® Phenyl-Hexyl, 5 µm

Eluent: A) methanol
B) 0.1 % formic acid in water
20–80 % A in 10 min

Flow rate: 0.56 mL/min
Temperature: 40 °C
Detection: UV, 254 nm
Injection: 0.5 µL

Peaks:
1. Sulfadiazine
2. Sulfachlorpyridazine
3. Sulfapyridine
4. Sulfamerazine
5. Sulfadimidine
6. Sulfathiazole
7. Sulfadimethoxine

On NUCLEOSHELL® Phenyl-Hexyl the resolution of the last two peaks is higher than on the fully porous 1.8 µm NUCLEODUR® Phenyl-Hexyl.



The separation of sulfonamides proves the scalability from fully porous NUCLEODUR® to NUCLEOSHELL® Phenyl-Hexyl. Hereby the core-shell silica exhibits identical selectivity, narrower peaks and slightly shorter retention under the same conditions.

Thus, method transferability between NUCLEODUR® and NUCLEOSHELL® is guaranteed, either for speeding up your methods or scaling up for preparative requirements.

Ordering information

Eluent in column acetonitrile – water

ID	Length → 50 mm	100 mm	150 mm	EC guard columns*
NUCLEOSHELL® Phenyl-Hexyl, 2.7 µm particle size 2.7 µm				
Analytical EC columns				
2 mm	763732.20	763734.20	763736.20	763738.20
3 mm	763732.30	763734.30	763736.30	763738.30
4 mm	763732.40	763734.40	763736.40	763738.30
4.6 mm	763732.46	763734.46	763736.46	763738.30

EC columns in packs of 1, guard columns in packs of 3.

Guard column system

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966

For details of the EC column system please see page 250.

NUCLEOSHELL® PFP hydrophobic pentafluorophenyl phase · USP L43

Key feature

- Core-shell technology for fast and efficient HPLC
- Hydrophobic phase with alternative selectivity in comparison to classical C_{18} modifications
- Separation principle based on 4 retention mechanisms (polar interactions (H bonds), dipole-dipole, π-π, hydrophobic interactions)

Technical data

- Phase with pentafluorophenylpropyl modification, multi-endcapping; pore size 90 Å, particle size 2.7 µm; carbon content ~ 3 %; pH stability 1–9; suitable for LC/MS

Recommended application

- Aromatic and unsaturated compounds, phenols, halogen compounds, isomers, polar compounds like pharmaceuticals, antibiotics; strong retention of basic compounds

Orthogonality in selectivity

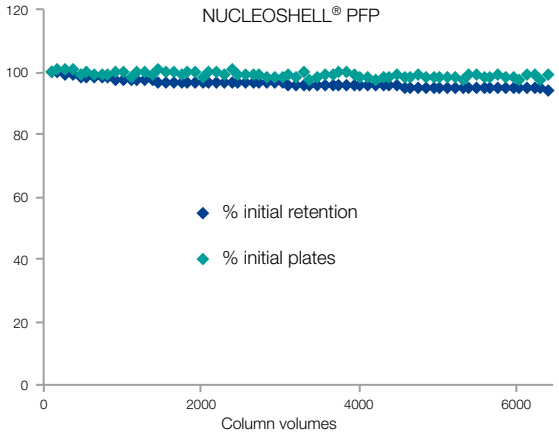
Fluorinated stationary phases in HPLC have gained increasing interest over the last years. Most common representative of fluorinated silica phases is the pentafluorophenyl modification (PFP or F_5). Especially the orthogonal selectivity compared to traditional alkyl phases widens the scope in analytical HPLC. Thus NUCLEOSHELL® PFP offers an excellent selectivity especially for highly polar analytes, aromatic and unsaturated compounds, phenols or halogenated hydrocarbons.

While a typical C_{18} phase just provides hydrophobic interactions between stationary phase and analyte NUCLEOSHELL® PFP offers four different retention mechanisms: polar interactions (H bonds), dipole-dipole interactions, π-π interactions and hydrophobic interactions. Especially the pronounced ion exchange capacity and distinct steric selectivity are typical for the character of fluorinated phases.

Stability of NUCLEOSHELL® PFP at pH 1

MN Appl. No. 125560

Columns:	100 x 4.6 mm NUCLEOSHELL® PFP, 2.7 µm 100 x 4.6 mm Kinetex® PFP, 2.6 µm F5
Eluent:	acetonitrile – 0.5 % TFA, pH 1 (50:50, v/v)
Flow rate:	1.3 mL/min
Temperature:	60 °C
Detection:	UV, 254 nm
Sample:	ethylbenzene



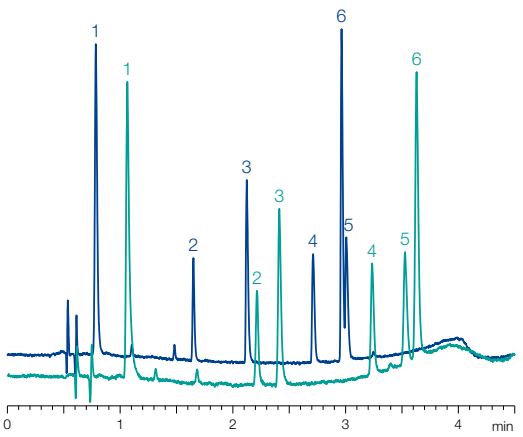
β-Blockers · orthogonal selectivity of NUCLEOSHELL® PFP

MN Appl. No. 125610

Columns:	100 x 4.6 mm NUCLEOSHELL® RP 18, 2.7 µm NUCLEOSHELL® PFP, 2.7 µm
Eluent:	A) acetonitrile + 0.1 % formic acid B) 0.1 % formic acid 10–35 % A in 2.5 min, 35-50 % A in 2 min
Flow rate:	1.7 mL/min
Temperature:	25 °C
Detection:	UV, 280 nm

Peaks:

1. Atenolol
2. Pindolol
3. Metroprolol
4. Labetalol
5. Alprenolol
6. Propranolol



Methylacetophenones

MN Appl. No. 125590

Columns: 100 x 4.6 mm NUCLEOSHELL® PFP, 2.7 µm
250 x 4 mm NUCLEODUR® PFP, 5 µm
100 x 4.6 mm Kinetex® 2.6 µm F5

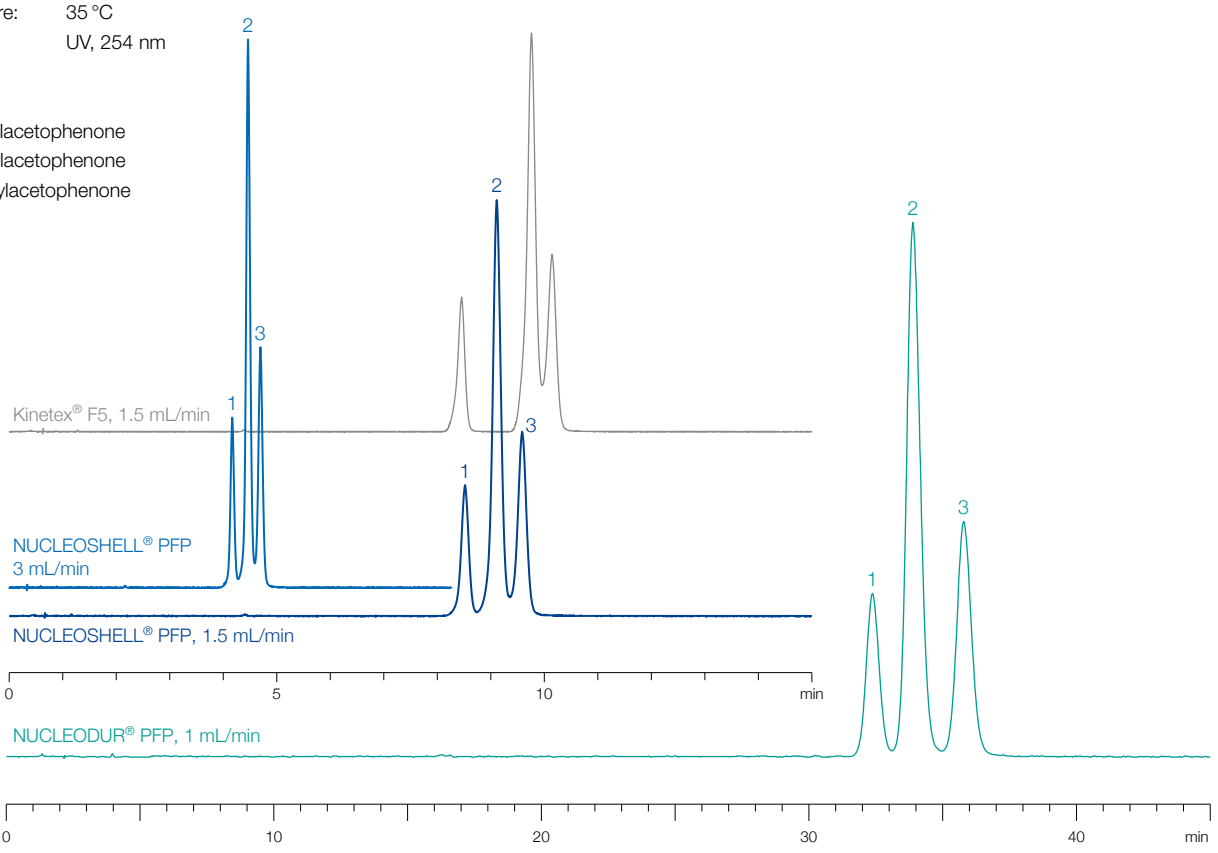
Eluent: Methanol – water (35:65, v/v)

Flow rate: 1.5 mL/min, 3 mL/min, 1 mL/min, 1.5 mL/min

Temperature: 35 °C

Detection: UV, 254 nm

Peaks:
1. *o*-Methylacetophenone
2. *p*-Methylacetophenone
3. *m*-Methylacetophenone



NUCLEOSHELL® PFP combines the benefits of core-shell technology, high stability, and orthogonal selectivity. Thus it is a useful complementary tool for highly efficient separations especially of isomers, halogenated, aromatic and / or polar compounds.

Ordering information

Eluent in column acetonitrile – water

ID	Length → 50 mm	100 mm	150 mm	EC guard columns*
NUCLEOSHELL® PFP, 2.7 µm particle size 2.7 µm				
Analytical EC columns				
2 mm	763532.20	763534.20	763536.20	763538.20
3 mm	763532.30	763534.30	763536.30	763538.30
4 mm	763532.40	763534.40	763536.40	763538.30
4.6 mm	763532.46	763534.46	763536.46	763538.30

EC columns in packs of 1, guard columns in packs of 3.

Guard column system

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966

For details of the EC column system please see page 250.

NUCLEOSHELL® columns

NUCLEOSHELL® HILIC zwitterionic phase

Key feature

- Core-shell technology for fast and efficient HPLC
- Ideal for reproducible and stable chromatography of highly polar analytes
- Very short column equilibration times

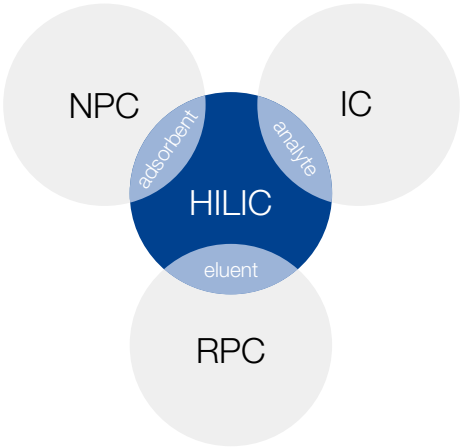
Technical data

- Ammonium - sulfonic acid modified silica; pore size 90 Å, particle size 2.7 µm; carbon content 1.3 %; pH stability 2–8.5; suitable for LC/MS

Recommended application

- Hydrophilic compounds such as polar organic acids and bases, polar natural compounds, nucleosides, oligonucleotides, amino acids, peptides, water-soluble vitamins

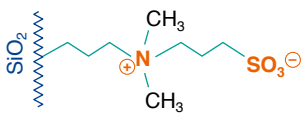
Hydrophilic interaction chromatography



Hydrophilic interaction chromatography (HILIC) is a separation technique using polar stationary phases and organic-aqueous mobile phases. A minimum water content of at least 2 % is indispensable to provide a permanent water layer between the adsorbent surface and the organic fraction of the mobile phase. The sample molecules become separated in a partition chromatography, in which polar analytes are more strongly retained than neutral, less hydrophilic compounds. Consequently, increasing the aqueous part in the mobile phase will diminish retention of the polar sample constituents. In this way HILIC behaves inverse to classical RP chromatography. The particular retention profile of HILIC enables the chromatography of very polar and often small molecules, which won't show any retention on C_8 or C_{18} reversed phases.

Ultra-fast separations at moderate back pressure

NUCLEOSHELL® HILIC is a core-shell technology based stationary phase with a covalently bonded 3-*N,N*-dimethylaminopropane sulfonic acid ligand (pat. p nd.). The betaine character of the strong ion-exchanger results in full charge balancing and facilitates fast equilibration times.



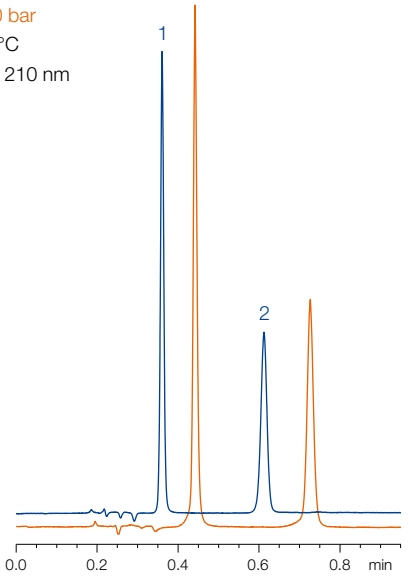
Good separation of polar compounds like the physiologically important substances creatine and creatinine can be achieved on NUCLEOSHELL® HILIC as well as on NUCLEODUR® HILIC, 1.8 µm at similar retention, but much lower back pressure.

Separation of creatine and creatinine

MN Appl. No. 124990

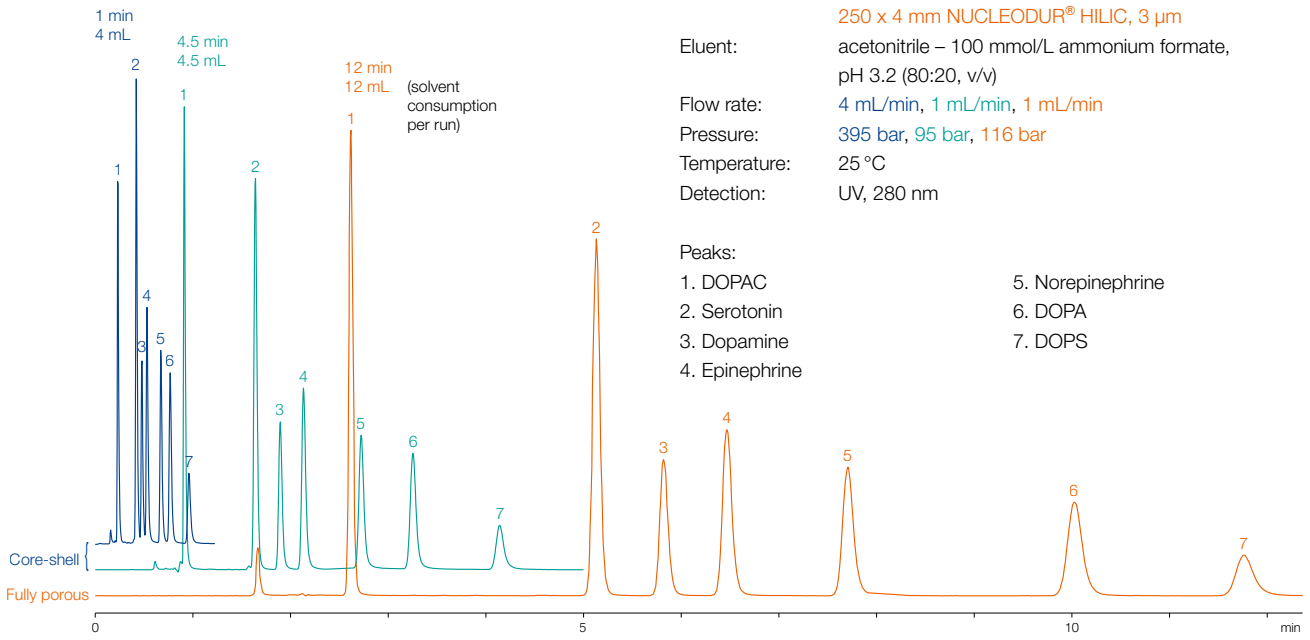
Columns:	50 x 4 mm NUCLEOSHELL® HILIC, 2.7 µm 50 x 4 mm NUCLEODUR® HILIC, 1.8 µm
Eluent:	acetonitrile – 10 mmol/L ammonium acetate, pH 4.0 (90:10, v/v)
Flow rate:	1.7 mL/min
Pressure:	129 bar 180 bar
Temperature:	25 °C
Detection:	UV, 210 nm

Peaks:
1. Creatinine
2. Creatine



The following chromatograms show the method transfer from a fully porous 3 µm HILIC phase to 2.7 µm core-shell silica with equal selectivity features.

Run time has been cut down to 1 min. Column back pressure remains modest < 400 bar, while solvent demand is reduced to less than 35 %.



Core-shell silica: separation in 1 min pressure < 400 bar

NUCLEOSHELL® HILIC provides stable and reproducible chromatography, comprising all the benefits of a state-of-the-art core-shell silica.

Ordering information

Eluent in column acetonitrile – water

ID	Length → 50 mm	100 mm	150 mm	EC guard columns*
NUCLEOSHELL® HILIC, 2.7 µm particle size 2.7 µm				
Analytical EC columns				
2 mm	763332.20	763334.20	763336.20	763338.20
3 mm	763332.30	763334.30	763336.30	763338.30
4 mm	763332.40	763334.40	763336.40	763338.30
4.6 mm	763332.46	763334.46	763336.46	763338.30

EC columns in packs of 1, guard columns in packs of 3.

Guard column system

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966

For details of the EC column system please see page 250.

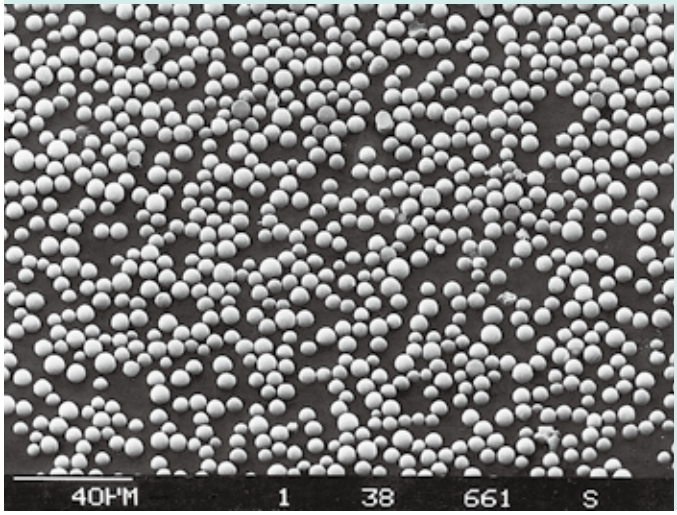
MACHEREY-NAGEL
Column Protection System

The guard column system for HPLC / UHPLC from MN

- Ideal protection for your analytical main column: significant increase in column lifetime
- Minimized void volume: suitable also for ultra fast HPLC (UHPLC)
- Special ferrules: pressure stability up to 1300 bar (18850 psi)
- Cartridges filled with NUCLEODUR®, NUCLEOSIL® and NUCLEOSHELL® HPLC adsorbents.
- Universal screw-on guard column holder system
- Suitable for all analytical HPLC columns with 1/16“ fittings

Further information on page 251.

NUCLEOSIL®



Key feature

- NUCLEOSIL® is a family of totally porous spherical silicas. They feature a very pure and uniform SiO_2 structure and have gained wide acceptance as routine chromatographic packings for very different fields of modern chromatography.
- One of the first spherical silicas used in HPLC
- Developed in the early seventies, it became a world-renowned HPLC packing
- Absolutely reliable choice for routine analyses
- Largest variety of modified HPLC silicas available
- pH stability 2–8 (for NUCLEOSIL® 100-5 C_{18} AB 1–9)
- Due to its particle sizes NUCLEOSIL® finds application in analytical as well as in preparative columns.

Benefits of NUCLEOSIL® silica

- High efficiency due to narrow particle size distribution
- High separation performance due to optimized binding techniques
- High chemical and mechanical stability
- High load capacity and recovery rates
- High reproducibility from lot to lot

Physical properties

NUCLEOSIL® is manufactured with different pore diameters (50, 100, 120, 300, 500, 1000 and 4000 Å) and particle sizes from 3 µm (only NUCLEOSIL® 50, 100 and 120) to 10 µm with very narrow fractionation. All narrow-pore NUCLEOSIL® packings are stable up to 500 bar (7 250 psi), the wide-pore NUCLEOSIL® silicas are stable up to 300 or 400 bar (4 200 or 5 600 psi).

Physical properties of unmodified NUCLEOSIL® materials

Phase	Pore size	Pore volume	Surface (BET)	Density	Pressure stability*
NUCLEOSIL® 50	50 Å	0.8 mL/g	420 m^2/g	0.45 g/mL	500 bar
NUCLEOSIL® 100	100 Å	1 mL/g	350 m^2/g	0.36 g/mL	500 bar
NUCLEOSIL® 120	120 Å	0.65 mL/g	200 m^2/g	0.55 g/mL	500 bar
NUCLEOSIL® 300	300 Å	0.8 mL/g	100 m^2/g	0.45 g/mL	400 bar
NUCLEOSIL® 500	500 Å	0.8 mL/g	35 m^2/g	0.45 g/mL	400 bar
NUCLEOSIL® 1000	1000 Å	0.8 mL/g	25 m^2/g	0.45 g/mL	300 bar
NUCLEOSIL® 4000	4000 Å	0.7 mL/g	10 m^2/g	0.48 g/mL	300 bar

* Maximum packing pressure of NUCLEOSIL® bulk packings

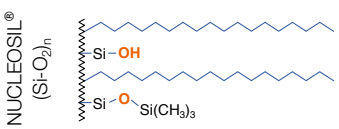
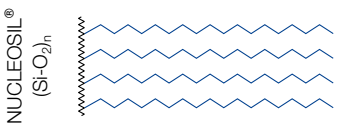
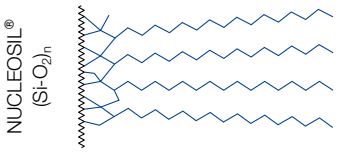
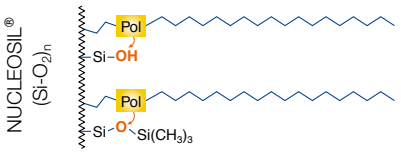
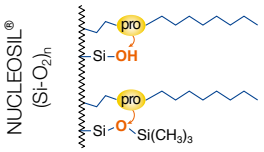
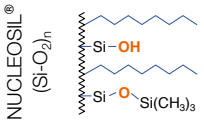
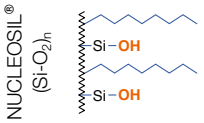
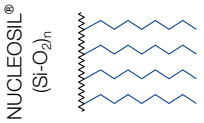
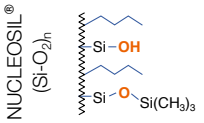
NUCLEOSIL® modifications

- NUCLEOSIL® packings are available as unmodified silica or with numerous chemically bonded phases: RP phases like C_{18} AB, C_{18} HD, C_{18} Nautilus, C_{18}, C_{18} ec, Protect I, C_8 HD, C_8 ec, C_8, C_4, C_2 and C_6H_5 separate mainly by hydrophobic interactions (van der Waals forces). The less polar the sample molecules, the more they are retained – the more polar the sample, the weaker are the hydrophobic interactions and consequently the retention times are shorter.
- Phases with chemically bonded polar groups such as CN, NH_2, $N(CH_3)_2$, OH show selective separation properties. Due to the availability of different functional groups it is possible to vary the chemical characteristics of the surface and consequently the adsorption characteristics of the stationary phase.
- Silica-based ion exchangers (NUCLEOSIL® SA and SB) are stable from pH 2 to 8 and do not swell. Compared to resin-based ion exchangers they offer the advantage of constant permeability, even when the ionic strength and/or pH of the eluent are changed. The separation can be influenced by
  – the type of buffer
  – the ionic strength and
  – the pH value.

A tabular overview of NUCLEOSIL® phases can be found on page 212.

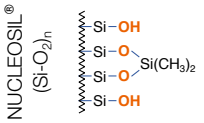
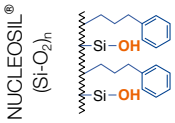
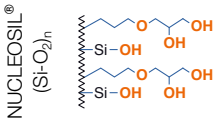
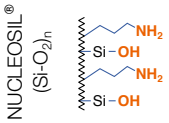
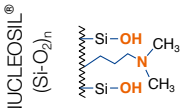
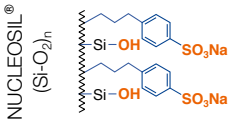
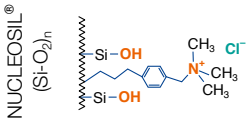
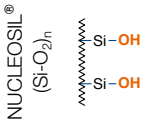
NUCLEOSIL® phase overview

Overview of NUCLEOSIL® HPLC phases

Phase	Specification	Page	Stability	Interactions	Structure
NUCLEOSIL® RP-Phasen					
C_{18}	octadecyl phase, medium density modification, endcapping 15 % C · USP L1	214	pH 2–8	hydrophobic (van der Waals) interactions slight residual silanol interactions	
C_{18} HD	octadecyl phase, high density monomeric modification, endcapping 20 % C · USP L1	214	pH 2–9	hydrophobic (van der Waals) interactions	
C_{18} AB	octadecyl phase, special crosslinked modification, endcapping 25 % C · USP L1	214	pH 1–9	steric and hydrophobic interactions	
C_{18} Nautilus	octadecyl phase, embedded polar group, endcapping 16 % C · USP L60	214	pH 2–8 up to 100 % H_2O	hydrophobic and polar interactions	
Protect I	special RP phase, protective polar group, monomeric modification, endcapping 11 % C	216	pH 2–8 up to 100 % H_2O	hydrophobic and polar interactions	
C_8 ec	octyl phase, medium density modification, endcapping 9 % C · USP L7	217	pH 2–8	hydrophobic (van der Waals) interactions slight residual silanol interactions	
C_8	octyl phase, no endcapping 8.5 % C · USP L7	217	pH 2–8	hydrophobic (van der Waals) interactions noticeable residual silanol interactions	
C_8 HD	octyl phase, high density modification, endcapping 13 % C · USP L7	218	pH 2–8	hydrophobic (van der Waals) interactions	
C_4	butyl phase, medium density modification, endcapping ~ 2 % C · USP L26	219	pH 2–8	hydrophobic (van der Waals) interactions residual silanol interactions	

NUCLEOSIL® phase overview

Overview of NUCLEOSIL® HPLC phases

Phase	Specification	Page	Stability	Interactions	Structure
C_2	dimethyl phase 3.5 % C · USP L16	219	pH 2–8	hydrophobic (van der Waals) interactions noticeable residual silanol interactions	
C_6H_5	phenyl phase, no endcapping 8 % C · USP L11	220	pH 2–8	π–π interactions and hydrophobic interactions noticeable residual silanol interactions	
Polar NUCLEOSIL® phases and NUCLEOSIL® ion exchangers					
CN / CN-RP	cyano (nitrile) phase USP L10	222	pH 2–8	π–π, polar and hydrophobic interactions	
OH (Diol)	diol · USP L20	220	pH 2–8	polar interactions (hydrogen bonds)	
NH_2 / NH_2-RP	amino · USP L8	221	pH 2–8	polar and hydrophobic interactions, weak ion exchange interactions	
$N(CH_3)_2$	dimethylamino	221	pH 2–8	polar and hydrophobic interactions, weak ion exchange interactions	
SA	sulfonic acid, strongly acid cation exchanger (SCX) USP L9	223	pH 2–8	strong ion exchange interactions	
SB	quaternary ammonium, strongly basic anion exchanger (SAX) USP L14	223	pH 2–8	strong ion exchange interactions	
SiOH	unmodified spherical silica USP L3	224	pH 2–8	polar	

NUCLEOSIL® octadecyl phases (C_{18})

NUCLEOSIL® standard octadecyl phases · USP L1

$-(CH_2)_{17}-CH_3$

Technical data

- Nonpolar phases
- pH stability at 20 °C: 2–8
- carbon content depending on pore size (see table)
- Corresponding NUCLEODUR® phases see C_{18} ec page 181

NUCLEOSIL® C_{18} HD · USP L1

$-(CH_2)_{17}-CH_3$

Technical data

- Nonpolar hydrophobic high density phases; monomeric modification
- pH stability 2–9
- Carbon content 20 %
- Corresponding NUCLEODUR® phases see C_{18} Gravity page 158

NUCLEOSIL® C_{18} AB · USP L1

$-(CH_2)_{17}-CH_3$

Technical data

- Crosslinked hydrophobic phase; polymeric modification; inert towards acidic and basic substances with high affinity for silica
- pH stability 1–9
- Carbon content 25 %; distinct steric selectivity
- Corresponding NUCLEODUR® phases see C_{18} Isis page 164

NUCLEOSIL® C_{18} Nautilus · USP L60

$-(CH_2)_{17}-CH_3$

Technical data

- Stable in 100 % aqueous eluents
- Carbon content 16 %
- Interesting polar selectivity features; very good base deactivation
- Corresponding NUCLEODUR® phases see C_{18} PolarTec page 168

All NUCLEOSIL® octadecyl phases are endcapped.
Custom-packed columns with different column dimensions are available on request.

Ordering information

Eluent in column acetonitrile – water

ID	Length → 100 mm	125 mm	150 mm	250 mm	EC guard columns*
NUCLEOSIL® 50-5 C_{18} ec particle size 5 µm, pore size 50 Å, endcapped, 14.5 % C					
Analytical EC columns					
4.6 mm				720098.46	721473.30
NUCLEOSIL® 100-3 C_{18} particle size 3 µm, pore size 100 Å, endcapped, 15 % C					
Analytical EC columns					
4 mm		720150.40		720133.40	721022.30
4.6 mm	720841.46	720150.46	720949.46	720133.46	721022.30
NUCLEOSIL® 100-5 C_{18} particle size 5 µm, pore size 100 Å, endcapped, 15 % C					
Analytical EC columns					
2 mm		720002.20		720014.20	721074.20
3 mm		720002.30		720014.30	721074.30
4 mm	720141.40	720002.40	720120.40	720014.40	721074.30
4.6 mm	720141.46	720002.46	720120.46	720014.46	721074.30

Ordering information

Eluent in column acetonitrile – water

ID	Length → 100 mm	125 mm	150 mm	250 mm	EC guard columns*
NUCLEOSIL® 100-7 C_{18} particle size 7 µm, pore size 100 Å, endcapped, 15 % C					
Analytical EC columns					
4 mm				720018.40	
4.6 mm		720951.46	720110.46	720018.46	
NUCLEOSIL® 100-10 C_{18} particle size 10 µm, pore size 100 Å, endcapped, 15 % C					
Analytical EC columns					
4 mm				720023.40	
4.6 mm		720701.46	720140.46	720023.46	
NUCLEOSIL® 120-3 C_{18} particle size 3 µm, pore size 120 Å, endcapped, 11 % C					
Analytical EC columns					
4 mm	720149.40	720040.40		720055.40	721075.30
4.6 mm	720149.46	720040.46	720740.46	720055.46	721075.30
NUCLEOSIL® 120-5 C_{18} particle size 5 µm, pore size 120 Å, endcapped, 11 % C					
Analytical EC columns					
4 mm		720051.40		720041.40	721070.30
4.6 mm		720051.46	720730.46	720041.46	721070.30
NUCLEOSIL® 120-7 C_{18} particle size 7 µm, pore size 120 Å, endcapped, 11 % C					
Analytical EC columns					
4 mm				720042.40	
NUCLEOSIL® 120-10 C_{18} particle size 10 µm, pore size 120 Å, endcapped, 11 % C					
Analytical EC columns					
4 mm				720043.40	
4.6 mm				720043.46	
NUCLEOSIL® 100-3 C_{18} HD particle size 3 µm, pore size 100 Å, 20 % C					
Analytical EC columns					
4 mm		720191.40			721196.30
4.6 mm		720191.46	720193.46		721196.30
NUCLEOSIL® 100-5 C_{18} HD particle size 5 µm, pore size 100 Å, 20 % C					
Analytical EC columns					
4 mm		720296.40		720280.40	721072.30
4.6 mm		720296.46	720294.46	720280.46	721072.30
NUCLEOSIL® 100-5 C_{18} AB particle size 5 µm, pore size 100 Å, 25 % C					
Analytical EC columns					
4 mm		720935.40		720936.40	721073.30
4.6 mm		720935.46	720305.46	720936.46	721073.30
NUCLEOSIL® 100-3 C_{18} Nautilus particle size 3 µm, pore size 100 Å, 16 % C					
Analytical EC columns					
4 mm		720472.40			721649.30
4.6 mm		720472.46	720471.46		721649.30
NUCLEOSIL® 100-5 C_{18} Nautilus particle size 5 µm, pore size 100 Å, 16 % C					
Analytical EC columns					
4 mm		720430.40		720431.40	721133.30
4.6 mm		720430.46	720432.46	720431.46	721133.30

Guard column system

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966

EC columns in packs of 1, guard columns in packs of 3. For details of our column systems see page 250.

NUCLEOSIL® columns

NUCLEOSIL® octadecyl phases (C_{18}) wide pore octadecyl phases · USP L1

$-(CH_2)_{17}-CH_3$

Technical data

- Many biologically interesting molecules can not be separated using conventional narrow pore silicas with pore sizes of about 100 Å. This is why MACHEREY-NAGEL offers a complete line of wide pore packings with pore sizes of 300, 500, 1000 and 4000 Å.
- These materials can also be used for size exclusion chromatography (SEC).

All NUCLEOSIL® octadecyl phases are endcapped.
Custom-packed columns with different column dimensions are available on request.

Ordering information

Eluent in column acetonitrile – water

ID	Length → 250 mm	EC guard columns*
NUCLEOSIL® 300-5 C_{18} particle size 5 µm, pore size 300 Å, endcapped, 6.5 % C		
Analytical EC columns		
4 mm	720065.40	721085.30
4.6 mm	720065.46	721085.30
NUCLEOSIL® 500-7 C_{18} particle size 7 µm, pore size 500 Å, endcapped, 2 % C		
Analytical EC columns		
4.6 mm	720074.46	
NUCLEOSIL® 1000-7 C_{18} particle size 7 µm, pore size 1000 Å, endcapped, ~ 1 % C		
Analytical EC columns		
4.6 mm	720077.46	

EC columns in packs of 1, guard columns in packs of 3.

VarioPrep preparative HPLC columns with NUCLEOSIL® packing material on request.

NUCLEOSIL® 100 Protect I special RP phase with protective polar group

Technical data

- RP phase with pronounced hydrophilic properties
- Monomeric coating
- Endcapped
- Carbon content 11 %

Ordering information

Eluent in column acetonitrile – water

ID	Length → 125 mm	150 mm	250 mm	EC guard columns*
NUCLEOSIL® 100-5 Protect I particle size 5 µm, pore size 100 Å				
Analytical EC columns				
4 mm	720175.40		720170.40	721157.30
4.6 mm	720175.46	720174.46	720170.46	721157.30

Guard column system

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966

EC columns in packs of 1, guard columns in packs of 3. For details of our column systems see page 250.

NUCLEOSIL® octyl phases (C_8) NUCLEOSIL® standard octyl phases · USP L7

$-(CH_2)_7-CH_3$

Technical data

- Nonpolar phases for RP and ion-pairing chromatography
- Endcapped and non-endcapped modifications available; pH stability at 20 °C: 2–8
- Carbon content depending on pore size (see table)

Recommended application

- Separation of moderately to highly polar (water-soluble) compounds: steroids, nucleosides, cyclodextrins, pharmacological plant constituents
- Corresponding NUCLEODUR® phases see C_8 ec page 183

Ordering information

Eluent in column acetonitrile – water

ID	Length → 125 mm	150 mm	250 mm	EC guard columns*
NUCLEOSIL® 100-5 C_8 ec particle size 5 µm, pore size 100 Å, endcapped, 9 % C				
Analytical EC columns				
4.6 mm			720165.46	721096.30
NUCLEOSIL® 100-5 C_8 particle size 5 µm, pore size 100 Å, not endcapped, 8.5 % C				
Analytical EC columns				
4 mm	720001.40		720013.40	721194.30
4.6 mm	720001.46	720990.46	720013.46	721194.30
NUCLEOSIL® 100-7 C_8 particle size 7 µm, pore size 100 Å, not endcapped, 8.5 % C				
Analytical EC columns				
4.6 mm			720017.46	
NUCLEOSIL® 100-10 C_8 particle size 10 µm, pore size 100 Å, not endcapped, 8.5 % C				
Analytical EC columns				
4 mm			720022.40	
4.6 mm			720022.46	
NUCLEOSIL® 120-3 C_8 particle size 3 µm, pore size 120 Å, not endcapped, 6.5 % C				
Analytical EC columns				
4 mm	720071.40			721093.30
4.6 mm	720071.46	720214.46		721093.30
NUCLEOSIL® 120-5 C_8 particle size 5 µm, pore size 120 Å, not endcapped, 6.5 % C				
Analytical EC columns				
4 mm	720050.40		720052.40	721095.30
4.6 mm	720050.46	720735.46	720052.46	721095.30
NUCLEOSIL® 300-5 C_8 particle size 5 µm, pore size 300 Å, not endcapped, ~ 3 % C				
Analytical EC columns				
4.6 mm			720062.46	721061.30

EC columns in packs of 1, guard columns in packs of 3.

Custom-packed columns with different column dimensions are available on request.

Guard column system

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966

EC columns in packs of 1, guard columns in packs of 3. For details of our column systems see page 250.

NUCLEOSIL® columns

NUCLEOSIL® octyl phases (C_8) NUCLEOSIL® C_8 HD · USP L7

$-(CH_2)_7-CH_3$

Technical data

- Nonpolar high density phases; monomeric modification; endcapped; carbon content 13 %
- Corresponding NUCLEODUR® phases see C_8 Gravity page 158

Recommended application

- Separation of moderate to strong polar (water soluble) analytes like steroids, cyclodextrines, pharmalogical plant ingredients

Ordering information

Eluent in column acetonitrile – water

ID	Length → 125 mm	150 mm	250 mm	EC guard columns*
NUCLEOSIL® 100-5 C_8 HD particle size 5 µm, pore size 100 Å				
Analytical EC columns				
4 mm			720196.40	721071.30
4.6 mm		720194.46	720196.46	721071.30

EC columns in packs of 1, guard columns in packs of 3.

Custom-packed columns with different column dimensions are available on request.

Guard column system

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966

EC columns in packs of 1, guard columns in packs of 3. For details of our column systems see page 250.

Beside analytical HPLC columns we also produce VarioPrep columns (see page 252) for preparative applications.

NUCLEOSIL® butyl phases (C_4) · USP L26

$-(CH_2)_3-CH_3$

Technical data

- Endcapped phases for RP and ion-pairing chromatography
- pH stability at 20 °C: 2–8; carbon content ~ 2 %
- Retention times are shorter than on C_8 and C_{18} phases

Recommended application

- For separation of macromolecules and hydrophobic substances
- For butyl phases for biochemical separations please refer to page 241

Ordering information

Eluent in column acetonitrile – water

ID		Length → 250 mm	EC guard columns*
NUCLEOSIL® 120-5 C_4 particle size 5 µm, pore size 120 Å			
Analytical EC columns			
	4.6 mm	720096.46	721083.30
NUCLEOSIL® 300-5 C_4 particle size 5 µm, pore size 300 Å			
Analytical EC columns			
	4 mm	720059.40	721916.30
	4.6 mm	720059.46	721916.30

EC columns in packs of 1, guard columns in packs of 3.

Guard column system

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966

NUCLEOSIL® dimethyl phase (C_2) · USP L16

$-(CH_3)_2$

Technical data

- Non-endcapped phase for RP and ion-pairing chromatography
- pH stability at 20 °C: 2–8; carbon content 3.5 %
- Retention times are much shorter than for the other RP phases

Ordering information

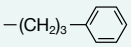
Eluent in column acetonitrile – water

ID		Length → 250 mm	EC guard columns*
NUCLEOSIL® 100-7 C_2 particle size 7 µm, pore size 100 Å			
Analytical EC columns			
	4.6 mm	720089.46	721030.30

EC columns in packs of 1, guard columns in packs of 3. For details of our column systems see page 250.

NUCLEOSIL® columns

NUCLEOSIL® phenyl phases (C_6H_5) · USP L11



Technical data

- Relatively nonpolar, non-endcapped phases for RP and ion pairing chromatography
- Polarity similar to C_8, but with different selectivity for PAHs, polar aromatics, fatty acids etc.
- pH stability at 20 °C: 2–8; carbon content 8 %

Recommended application

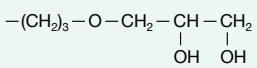
- Separation of moderately polar compounds

Ordering information

Eluent in column acetonitrile – water

ID	Length → 250 mm	EC guard columns*
NUCLEOSIL® 100-5 C_6H_5 particle size 5 µm, pore size 100 Å, not endcapped		
Analytical EC columns		
4.6 mm	720956.46	721137.30
NUCLEOSIL® 100-7 C_6H_5 particle size 7 µm, pore size 100 Å, not endcapped		
Analytical EC columns		
4 mm	720019.40	
4.6 mm	720019.46	

NUCLEOSIL® diol phases · USP L20



Technical data

- Dihydroxypropyl modified silica for RP and NP chromatography
- Less polar than unmodified silica, very easily wettable with water
- pH stability at 20 °C: 2-8; carbon content 5 %

Ordering information

Eluent in column is *n*-heptane. When using an eluent which is not miscible with *n*-heptane (e.g., water), it is necessary to rinse the column with THF first.

ID	Length → 250 mm	EC guard columns*
NUCLEOSIL® 100-5 OH (Diol) particle size 5 µm, pore size 100 Å		
Analytical EC columns		
4.6 mm	720143.46	721142.30

Guard column system

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966

EC columns in packs of 1, guard columns in packs of 3. For details of our column systems see page 250.

NUCLEOSIL® amino phases · USP L8

$-(CH_2)_3-NH_2$

Technical data

- Aminopropyl modified polar silica phase; pH stability at 20 °C: 2–8; carbon content 3.5 %
- Corresponding NUCLEODUR® phases see page 188

Recommended application

Multi-mode chromatography

- NP chromatography with hexane, dichloromethane or 2-propanol as mobile phase for polar compounds such as substituted anilines, esters, chlorinated pesticides
- RP chromatography of polar compounds like carbohydrates in aqueous-organic eluent systems
- Anion exchange chromatography of anions and organic acids using common buffers (e.g., acetate or phosphate) in conjunction with organic modifiers (e.g., acetonitrile)

Ordering information

Eluent in column is *n*-heptane (except for NH_2 RP). When using an eluent which is not miscible with *n*-heptane (e.g., water), it is necessary to rinse the column with THF first.

ID	Length → 250 mm	EC guard columns*
NUCLEOSIL® 100-5 NH_2 particle size 5 µm, pore size 100 Å; eluent in column *n*-heptane		
Analytical EC columns		
4.6 mm	720095.46	721020.30
NUCLEOSIL® 100-5 NH_2-RP particle size 5 µm, pore size 100 Å; eluent in column acetonitrile – water (80:20)		
Analytical EC columns		
4.6 mm	720095.46RP	721155.30
NUCLEOSIL® 100-10 NH_2 particle size 10 µm, pore size 100 Å; eluent in column *n*-heptane		
Analytical EC columns		
4.6 mm	720025.46	

NUCLEOSIL® dimethylamino phase

$-(CH_2)_3-N(CH_3)_2$

Technical data

- Weakly basic anion exchanger, pH stability at 20 °C: 2–8; carbon content 4 %

Recommended application

- Separation of many anions; can also be used in a similar way as the NH_2 phase

Ordering information

Eluent in column is *n*-heptane. When using an eluent which is not miscible with *n*-heptane (e.g., water), it is necessary to rinse the column with THF first.

ID	Length → 250 mm	EC guard columns*
NUCLEOSIL® 100-5 $N(CH_3)_2$ particle size 5 µm, pore size 100 Å		
Analytical EC columns		
4.6 mm	720994.46	721158.30

Guard column system

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966

EC columns in packs of 1, guard columns in packs of 3. For details of our column systems see page 250.

NUCLEOSIL® cyano phases · USP L10

$-(CH_2)_3-CN$

Technical data

- Polar to midpolar cyano (nitrile) modified silica
- pH stability at 20 °C: 2–8; carbon content 5 % for 100 Å pores, ~ 3 % for 120 Å pores
- Corresponding NUCLEODUR® phases see page 186

Recommended application

Reversed phase and normal phase chromatography

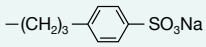
- Normal phase:
  with low-polarity solvents for many compounds, which can also be separated on unmodified silica, however, due to the rapid equilibration much more suitable for gradient separations
- Reversed phase:
  with different selectivity than C_{18}, C_8 or phenyl modified packings

Ordering information

Eluent in column (except for NUCLEOSIL® 100-5 CN-RP) is *n*-heptane. When using an eluent which is not miscible with *n*-heptane (e.g., water), it is necessary to rinse the column with THF first.

ID	Length → 250 mm	EC guard columns*
NUCLEOSIL® 100-5 CN particle size 5 µm, pore size 100 Å; eluent in column *n*-heptane		
Analytical EC columns		
4 mm	720090.40	721078.30
4.6 mm	720090.46	721078.30
NUCLEOSIL® 100-5 CN-RP particle size 5 µm, pore size 100 Å; eluent in column acetonitrile – water		
Analytical EC columns		
4 mm	720205.40	721039.30
4.6 mm	720205.46	721039.30
NUCLEOSIL® 100-10 CN particle size 10 µm, pore size 100 Å; eluent in column *n*-heptane		
Analytical EC columns		
4 mm	720024.40	
4.6 mm	720024.46	
NUCLEOSIL® 120-7 CN particle size 7 µm, pore size 120 Å; eluent in column *n*-heptane		
Analytical EC columns		
4 mm	720057.40	
4.6 mm	720057.46	

NUCLEOSIL® SA phases · USP L9



Technical data

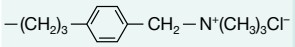
- Strongly acidic cation exchanger (SCX) with benzenesulfonic acid modification
- Capacity ~ 1 meq/g; pH stability at 20 °C: 2–8; carbon content 6.5 %

Ordering information

Eluent in column 0.15 mol/L $(NH_4)_2HPO_4$, pH 5

ID	Length → 125 mm	150 mm	250 mm	EC guard columns*
NUCLEOSIL® 100-5 SA particle size 5 µm, pore size 100 Å				
Analytical EC columns				
4 mm			720097.40	721024.30
4.6 mm	720709.46	720182.46	720097.46	721024.30
NUCLEOSIL® 100-10 SA particle size 10 µm, pore size 100 Å				
Analytical EC columns				
4.6 mm			720028.46	

NUCLEOSIL® SB phases · USP L14



Technical data

- Strongly basic anion exchanger (SAX) with quaternary ammonium modification
- Capacity ~ 1 meq/g; pH stability at 20 °C: 2–8; carbon content 10 %

Ordering information

Eluent in column 0.15 mol/L $(NH_4)_2HPO_4$, pH 5

ID	Length → 125 mm	150 mm	250 mm	EC guard columns*
NUCLEOSIL® 100-5 SB particle size 5 µm, pore size 100 Å				
Analytical EC columns				
4 mm			720996.40	721025.30
4.6 mm	720989.46	720183.46	720996.46	721025.30
NUCLEOSIL® 100-10 SB particle size 10 µm, pore size 100 Å				
Analytical EC columns				
4.6 mm			720029.46	

NUCLEOSIL® columns

NUCLEOSIL® SiOH unmodified silica · USP L3

Technical data

- Spherical silica, pH stability 2–8
- For physical properties of unmodified NUCLEOSIL® materials please see page 211.
- Maximum working pressure for the EC columns listed below is 400 bar.

Ordering information

Eluent in column is *n*-heptane. When using an eluent which is not miscible with *n*-heptane (e.g., water), it is necessary to rinse the column with THF first.

	ID	Length → 250 mm	EC guard columns*
NUCLEOSIL® 50-5 particle size 5 µm, pore size 50 Å			
Analytical EC columns			
	4.6 mm	720093.46	721167.30
NUCLEOSIL® 100-5 particle size 5 µm, pore size 100 Å			
Analytical EC columns			
	4.6 mm	720099.46	721518.30

Guard column system

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966

EC columns in packs of 1, guard columns in packs of 3. For details of our column systems see page 250.

LiChrospher® packings manufactured by E. Merck (D)

Phase	USP	Particle size	Pore size	Modification	Endcapped	Carbon content
LiChrospher® 100 RP 18, 5 µm	L1	nom. 5 µm	100 Å	Octadecyl	–	21 %
LiChrospher® 100 RP 18 ec, 5 µm	L1	nom. 5 µm	100 Å	Octadecyl	+	21 %
LiChrospher® 60 RP select B, 5 µm	L7	nom. 5 µm	60 Å	Octyl	+	12 %
All phases as packed ChromCart® cartridges						
ChromCart® columns require the CC connecting kit (REF 721690).						

Ordering information

Eluent in column acetonitrile – water

ID	Length → 125 mm	150 mm	250 mm	Guard columns*
LiChrospher® 100 RP 18, 5 µm particle size 5 µm, pore size 100 Å				
2 mm	728031.20		728032.20	728053.30
3 mm	728031.30		728032.30	728053.30
4 mm	728031.40		728032.40	728053.40
4.6 mm	728031.46	728033.46	728032.46	728053.40
LiChrospher® 100 RP 18 ec, 5 µm particle size 5 µm, pore size 100 Å				
2 mm	728034.20		728035.20	728054.30
3 mm	728034.30		728035.30	728054.30
4 mm	728034.40		728035.40	728054.40
4.6 mm	728034.46	728036.46	728035.46	728054.40
LiChrospher® 60 RP select B, 5 µm particle size 5 µm, pore size 100 Å				
2 mm	728037.20		728038.20	728055.30
3 mm	728037.30		728038.30	728055.30
4 mm	728037.40		728038.40	728055.40
4.6 mm	728037.46	728039.46	728038.46	728055.40

* can directly be used with the CC connecting kit (REF 721690).

8 mm ChromCart® guard column cartridges in packs of 3, all other columns in packs of 1.

Phase overview for special separations

Overview

Separation / mechanism	Recommended column	Specification of the phase	Page
Environmental analysis			
Anion exchange chromatography of inorganic anions	NUCLEOGEL® Anion I	Strongly basic polymer-based anion exchanger	230
	NUCLEOSIL® Anion II	Strongly basic silica-based anion exchanger	
RP chromatography of PAHs	NUCLEODUR® C_{18} PAH	NUCLEODUR® polymer-coated with C_{18} groups USP L1	227
	NUCLEOSIL® 100-5 C_{18} PAH	NUCLEOSIL® 100 polymer-coated with C_{18} groups USP L1	229
Enantiomer separation			
Polar and π-π interactions	NUCLEOCEL *DELTA*	Silica-based modified cellulose phases USP L40	233
Formation of inclusion complexes	NUCLEODEX α-PM, β-PM, γ-PM and β-OH	Silica-based permethylated and underivatized cyclodextrin phases USP L45	231
Enantioselective binding to chiral protein surface structures	RESOLVOSIL BSA-7	Silica-based protein phase (BSA)	234
Ligand exchange	NUCLEOSIL® CHIRAL-1	Covalently bonded amino acid – Cu(II) complexes USP L32	235
Charge-transfer, dipole-dipole interactions and others	NUCLEOSIL® CHIRAL-2 NUCLEOSIL® CHIRAL-3	Silica-based brush type phases USP L36	236
Separation of biological macromolecules			
Anion exchange chromatography of oligonucleotides and nucleic acids	NUCLEOGEN® DEAE	Silica-based DEAE anion exchanger	237
Anion exchange chromatography of peptides, large proteins and oligonucleotides	NUCLEOGEL® SAX	Polymer-based strongly basic anion exchanger USP L23	240
Cation exchange chromatography of proteins, peptides and carbohydrates	NUCLEOGEL® SCX	Polymer-based strong cation exchanger USP L22	240
Reversed phase chromatography of proteins, peptides and oligonucleotides	NUCLEOSIL® MPN	Monomerically bonded alkyl chains on silica USP L1 / USP L26	243
	NUCLEOSIL® PPN	Polymerically bonded alkyl chains on silica USP L1	244
	NUCLEOGEL® RP 300	Polystyrene – divinylbenzene polymer USP L21	245
Reversed phase chromatography of small molecules	NUCLEOGEL® RP 100	Small pore macroporous PS-DVB polymer USP L21	245
Food analysis · sugars			
RP chromatography of mono- and oligosaccharides	NUCLEOSIL® Carbohydrate	Silica-based special amino phase USP L8	246
Separation of sugars, alcohols, org. acids based on ion exclusion, ion exchange, size exclusion, ligand exchange, NP and RP effects	NUCLEOGEL® SUGAR 810 H, Ca	Resins with sulfonic acid modification in different ionic forms H form USP L17 / Ca form L19 / Pb form L34 / Na form L58	247
Separation of sugars, alcohols, org. acids based on steric exclusion, ligand exchange and partition effects	NUCLEOGEL® SUGAR Ca, Na, Pb NUCLEOGEL® ION 300 OA		248
Gel permeation chromatography (GPC)			
Water-insoluble compounds	NUCLEOGEL® GPC	Polystyrene – divinylbenzene polymer	249

NUCLEODUR® C_{18} PAH special octadecyl phase for PAH analysis · USP L1

Technical data

- Base material NUCLEODUR® silica, particle sizes 1.8 and 3 µm, pore size 110 Å; polymeric coating

Recommended application

- Allows efficient gradient separation of the 16 PAHs according to EPA

Analysis of 16 EPA PAHs with or without acetonitrile

MN Appl. Nos. 123820 / 123830

Separation with acetonitrile

Column: 100 x 4 mm NUCLEODUR® C18 PAH, 3 µm

Eluent: A) methanol – water (80:20, v/v) B) acetonitrile 2–20 % B in 1.2 min, 20–100 % B in 0.5 min, 100 % B for 2.5 min, 100–2 % B in 0.4 min

Flow rate: 2.5 mL/min, temperature 35 °C

Detection: UV, 254 nm fluorescence (see chromatogram)

Separation without acetonitrile

Column: 125 x 4 mm NUCLEODUR® C18 PAH, 3 µm

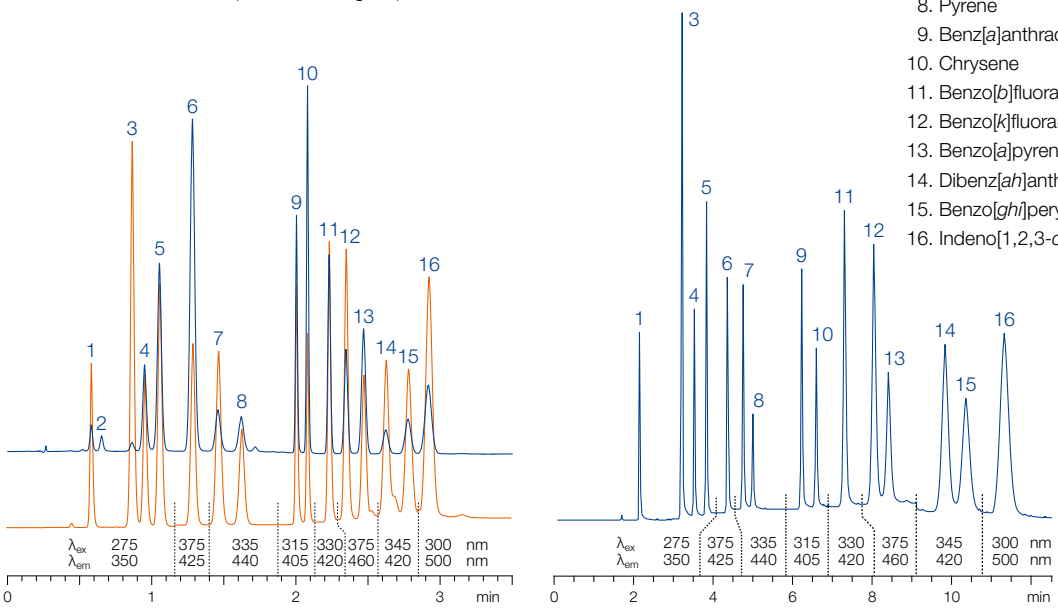
Eluent: A) water B) methanol 65–97 % B in 6 min, 97 % B for 5 min, 97–65 % B in 0.5 min

Flow rate: 2 mL/min, temperature 35 °C

Detection: fluorescence (see chromatogram)

Peaks:

1. Naphthalene
2. Acenaphthylene (not detectable by fluorescence)
3. Acenaphthene
4. Fluorene
5. Phenantrene
6. Anthracene
7. Fluoranthene
8. Pyrene
9. Benz[*a*]anthracene
10. Chrysene
11. Benzo[*b*]fluoranthene
12. Benzo[*k*]fluoranthene
13. Benzo[*a*]pyrene
14. Dibenz[*ah*]anthracene
15. Benzo[*ghi*]perylene
16. Indeno[1,2,3-*cd*]pyrene



Detection of separated PAHs with UV (250–280 nm), diode array or fluorescence detection at different wavelengths for excitation and emission (acenaphthylene cannot be analyzed with fluorescence detection).

Ordering information

Eluent in column acetonitrile – water (70:30, v/v)

ID	Length → 100 mm	125 mm	150 mm	250 mm	EC guard columns*
NUCLEODUR® C_{18} PAH, 1.8 µm particle size 1.8 µm · UHPLC					
Analytical EC columns					
2 mm	760773.20				761970.20
3 mm	760773.30				761970.30
4 mm	760773.40				761970.30
NUCLEODUR® C_{18} PAH, 3 µm particle size 3 µm					
Analytical EC columns					
3 mm	760783.30	760784.30	760785.30	760786.30	761971.30
4 mm	760783.40	760784.40	760785.40	760786.40	761971.30

Guard column system

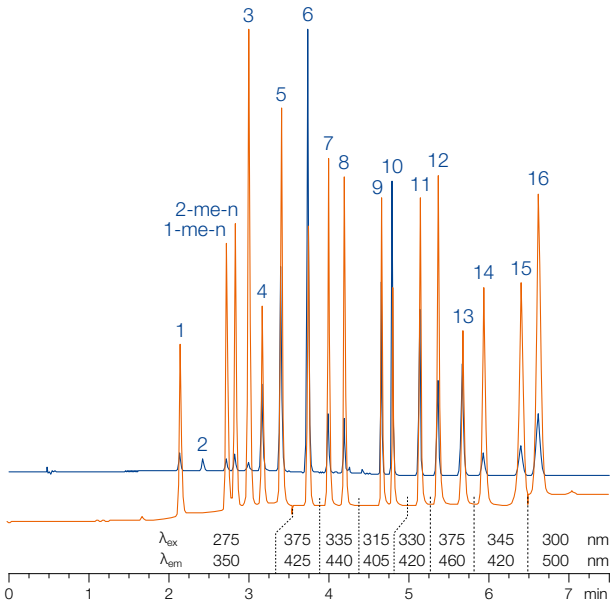
Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966

Separation of 18 PAHs on NUCLEODUR® C_{18} PAH

MN Appl. No. 123840

Column:	125 x 4 mm NUCLEODUR® C_{18} PAH, 3 µm
Eluent:	A) methanol – water (70:30, v/v); B) acetonitrile 0–20 % B in 1.5 min, 20–50 % B in 1.5 min, 50–100 % B in 1.0 min, 100 % B for 3 min, 100–0 % B in 0.5 min
Flow rate:	1.5 mL/min
Temperature:	35 °C
Injection:	UV: 1 µL,
Fluorescence:	0.5 µL
Detection:	UV, 254 nm fluorescence (see chromatogram)

Peaks:
(concentrations 10 ng/µL per compound)
1.–16. see page 227
1-me-n: 1-methylnaphthalene
2-me-n: 2-methylnaphthalene

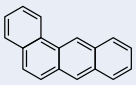


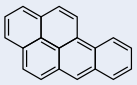
Analysis of polycyclic aromatic hydrocarbons (PAHs) by HPLC

Polycyclic aromatic hydrocarbons (PAHs) are chemical compounds that consist of fused aromatic rings and do not contain heteroatoms or carry substituents. As a pollutant, they are of concern because some compounds have been identified as carcinogenic, mutagenic, and teratogenic. PAHs are natural components of coal or gas. They are delivered to our environment by pyrolysis (incomplete burning) of organic materials like coal, oil, fuel, wood, tobacco, … and hence can be found globally. Today most PAHs accrue from anthropogenic processes – but also natural origins (forest fire) are possible. Regarding to past pollutions an important impact had production of coke and gas from black coal. Waste products (e.g., tar) from coking or gas plants are often origin of serious ground water pollutions.

Since a number of PAHs (e.g., benzo[a]pyrene, 3-methylcholanthrene and benzanthracene) have been proven to be carcinogenic, control of the PAH content of food, water and soil is an important task for routine analysis. For choice and limiting values of the polycyclics we refer to the governmental regulations, which exist in many countries (e.g., EPA method 610 of the United States Environmental Protection Agency).

PAHs can be determined by different chromatographic techniques (TLC, GC, HPLC). Thus the 6 PAHs according to German drinking water specification (TVO) can, e.g., be analyzed by TLC (see German Standard DIN 38 409), while a much larger number of polycyclic aromatics can be determined by GC or HPLC.


Benzo[a]anthracen


Benzo[a]pyren

HPLC columns for PAH analysis

For PAH analyses we have developed specially modified C_{18} phases based on NUCLEODUR® and NUCLEOSIL® which allow efficient gradient separation of 16 PAHs according to EPA. Detection of the separated PAHs can be achieved by UV (250–280 nm), with diode array or with fluorescence detection at different wavelengths for excitation and emission. Acenaphthylene cannot be analyzed with fluorescence detection. For cost-effective routine PAH analysis we recommend applications using methanol instead of acetonitrile as eluent. For rapid analysis NUCLEODUR® C_{18} PAH (3 µm) in short columns (100 mm) provides excellent results at high flow rates. Hereby separation of 16 PAHs according to EPA can be achieved in less than 3 min.

Tightened regulations require determination of 2 additional PAHs (1- and 2-methylnaphthalene) – so we developed highly efficient methods for 18 PAHs on the NUCLEODUR® C_{18} PAH.

NUCLEOSIL® 100-5 C_{18} PAH special octadecyl phase for PAH analysis · USP L1

Technical data

- Base material NUCLEOSIL® silica, particle size 5 µm, pore size 100 Å; polymeric coating
- Detection of the separated PAH with UV (250–280 nm), diode array or fluorescence detection at different wavelengths for excitation and emission (acenaphthylene cannot be analyzed with fluorescence detection)

Recommended application

- Efficient gradient separation of the 16 PAHs according to EPA

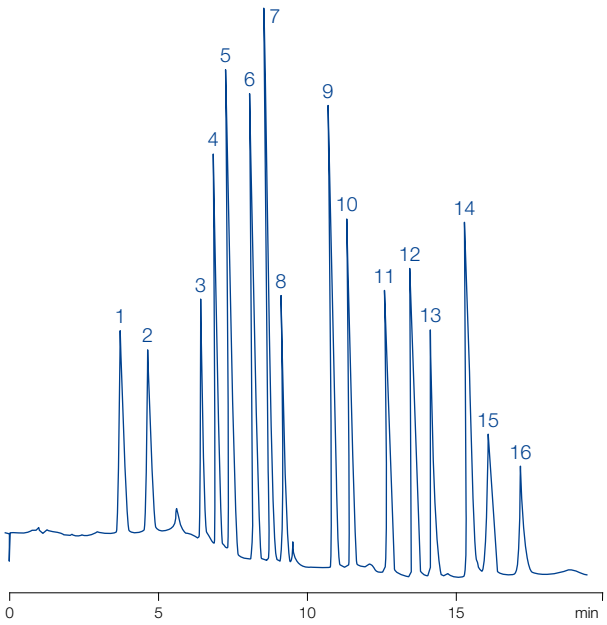
Separation of the PAH standard according to EPA (REF 722393)

MN Appl. No. 115040

Column:	150 x 4 mm NUCLEOSIL® 100-5 C_{18} PAH
Eluent:	A) methanol – water (80:20)
	B) acetonitrile – tetrahydrofuran (93:7)
	0–100 % B in 10 min, 5 min 100 % B
Flow rate:	1 mL/min
Pressure:	140 bar
Temperature:	20 °C
Detection:	UV, 260 nm

Peaks: (10 µg/mL each in acetonitrile)

1. Naphthalene
2. Acenaphthylene
3. Acenaphthene
4. Fluorene
5. Phenanthrene
6. Anthracene
7. Fluoranthene
8. Pyrene
9. Benz[*a*]anthracene
10. Chrysene
11. Benzo[*b*]fluoranthene
12. Benzo[*k*]fluoranthene
13. Benzo[*a*]pyrene
14. Dibenz[*ah*]anthracene
15. Benzo[*ghi*]perylene
16. Indeno[1,2,3-*cd*]pyrene



Ordering information

Eluent in column acetonitrile – water 70:30

ID	Length → 150 mm	250 mm	EC guard columns*
NUCLEOSIL® 100-5 C_{18} PAH particle size 5 µm, pore size 100 Å			
Analytical EC columns			
2 mm		720117.20	721168.20
3 mm	720923.30	720117.30	721168.30
4 mm	720923.40	720117.40	721168.30
4.6 mm		720117.46	721168.30
PAH standard according to EPA for HPLC			
Analytical EC columns			
PAH standard for HPLC	16 PAH according to EPA method 610 in acetonitrile (1 mL) for composition see chromatogram above		722393

Guard column system

Guard columns for EC columns with ID		2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966

EC columns in packs of 1, guard columns in packs of 3. For details of our column systems see page 250.

This product contains harmful substances which must be specially labeled as hazardous. For detailed information please see SDS.

HPLC columns for environmental analyses

Anion columns for analysis of inorganic anions

NUCLEOGEL® Anion I

Technical data

- Strongly basic polymer-based anion exchanger, particle size 10 µm; pH stability 1–14
- Eluent in column 4 mmol/L salicylate buffer pH 7.8
- Contrary to the silica-based phase also suited for fluoride analysis

NUCLEOSIL® Anion II

Technical data

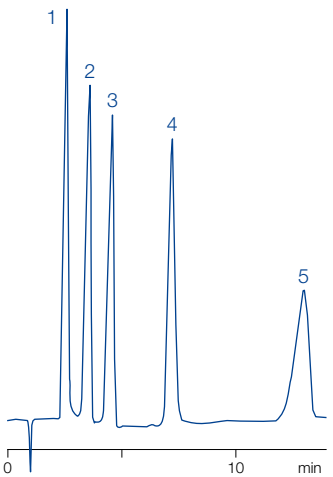
- Base material NUCLEOSIL® silica, particle size 10 µm, pore size 300 Å strongly basic anion exchanger, exchange capacity 50 µeq/g, pH stability 2–7.5
- Eluent in column 0.15 mol/L $(NH_4)_2HPO_4$ buffer pH 5.2 recommended buffer concentration for separation of inorganic anions: 2 mmol/L phthalate
- Preferred method of detection: conductivity or negative UV detection

Separation of an anion standard

MN Appl. No. 106440

Column: 250 x 4 mm NUCLEOSIL® Anion II
Eluent: 2 mmol/L potassium hydrogen phthalate, pH 5.7
Flow rate: 2 mL/min
Detection: UV, 280 nm

Peaks:
1. H_2PO^{4-}
2. Cl^-
3. NO^{2-}
4. NO^{3-}
5. SO_4^{2-}

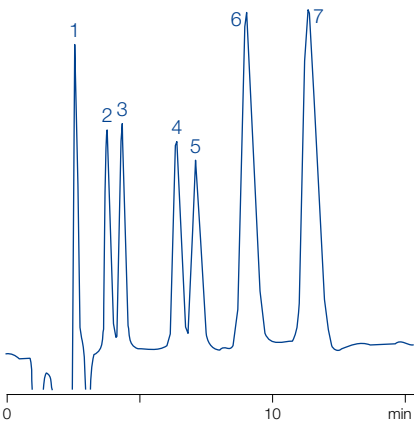


Separation of inorganic anions

MN Appl. No. 115050

Column: 120 x 4.6 mm NUCLEOGEL® Anion I
Eluent: 4 mmol/L salicylic acid – Tris pH 7.8
Flow rate: 1 mL/min
Detection: UV, 254 nm

Peaks:
1. F^-
2. Cl^-
3. NO^{2-}
4. Br^-
5. NO_3^-
6. PO_4^{3-}
7. SO_4^{2-}

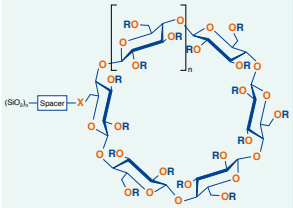


Ordering information

	ID	Length → 120 mm	250 mm	Guard columns*
NUCLEOGEL® Anion I eluent 4 mmol/L salicylate buffer pH 7.8				
Analytical Valco type columns				
	4.6 mm	719533		719543
NUCLEOSIL® Anion II eluent 0.15 mol/L $(NH_4)_2HPO_4$ buffer pH 5.2				
Analytical EC columns				
	4 mm		720094.40	721169.30

* NUCLEOGEL® Anion I Valco type guard columns cartridges are 21 x 4 mm, require guard column holder C, REF 719538, see page 250 (columns in packs of 1, guard columns in packs of 2)
NUCLEOSIL® Anion II guard columns are used with the Column Protection System (REF 718966, see page 251).

NUCLEODEX columns enantiomer separation based on cyclodextrins



NUCLEODEX β-OH β-cyclodextrin (R = H; n = 2) · USP L45

Technical data

- Base material NUCLEOSIL® silica, particle size 5 µm, pore size 100 Å modified cyclodextrins as chiral selectors
- Separation based on hydrogen bonds and dipole interactions between functional groups of the analyte and hydroxyl groups of the cyclodextrin
- Examples for successful enantiomer separations: chlorthalidone and other compounds, which require free hydroxyl groups for enantioselective interactions
- Eluent in column CH_3OH – 0.1 % TEAA pH 4 (55:45)

NUCLEODEX α-PM permethylated α-cyclodextrin (R = CH_3; n = 1)

Technical data

- Base material NUCLEOSIL® silica, particle size 5 µm, pore size 100 Å modified cyclodextrins as chiral selectors
- Examples for successful enantiomer separations: mecoprop and dichlorprop as free carboxylic acids, trans-stilbene oxide, styrene oxide
- Eluent in column CH_3OH – 50 mmol/L phosphate pH 3 (70:30)

NUCLEODEX β-PM permethylated β-cyclodextrin (R = CH_3; n = 2) · USP L45

Technical data

- Base material NUCLEOSIL® silica, particle size 5 µm, pore size 100 Å modified cyclodextrins as chiral selectors
- Examples for successful enantiomer separations: mephobarbital (prominal), pesticide derivatives mecoprop methyl and dichlorprop methyl
- Eluent in column CH_3OH – 0.1 % TEAA pH 4 (65:35)

NUCLEODEX γ-PM permethylated γ-cyclodextrin (R = CH_3; n = 3)

Technical data

- Base material NUCLEOSIL® silica, particle size 5 µm, pore size 100 Å modified cyclodextrins as chiral selectors
- Examples for successful enantiomer separations: steroids or other larger molecules
- Eluent in column CH_3OH – 0.1 % TEAA pH 4 (55:45)

Recommended application

- NUCLEODEX phases are especially suited for the control of optical purity, but also for semipreparative separations and for the analysis of positional and cis-trans isomers.
- For numerous separations on NUCLEODEX phases please visit our website: *www.mn-net.com/apps*

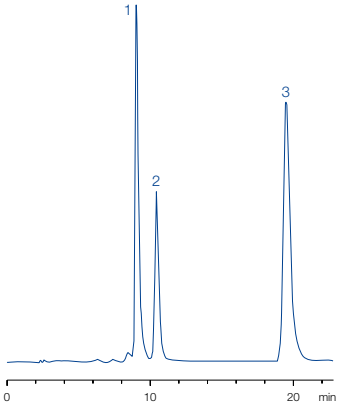
HPLC columns for enantiomer separations

Separation of the positional isomers of nitroaniline

MN Appl. No. 101420

Column:	200 x 4 mm NUCLEODEX β-OH
Eluent:	methanol – 0.1 % triethylammonium acetate pH 4.0 (50:50, v/v)
Flow rate:	0.7 mL/min
Pressure:	180 bar
Detection:	UV, 254 nm
Injection:	1 µL

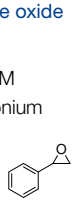
Peaks:
1. *m*-Nitroaniline
2. *o*-Nitroaniline
3. *p*-Nitroaniline



Enantiomer separation of styrene oxide

MN Appl. No. 106160

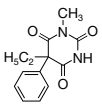
Column:	200 x 4 mm NUCLEODEX α-PM
Eluent:	methanol – 0.1 % triethylammonium acetate pH 4.0 (60:40, v/v)
Flow rate:	0.7 mL/min
Pressure:	160 bar
Detection:	UV, 230 nm
Injection:	2 µL



Enantiomer separation of mephobarbital

MN Appl. No. 105800

Column:	200 x 4 mm NUCLEODEX β-PM
Eluent:	methanol – 0.1 % triethylammonium acetate pH 4.0 (55:45, v/v)
Flow rate:	0.7 mL/min
Pressure:	180 bar
Detection:	UV, 254 nm
Injection:	1 µL

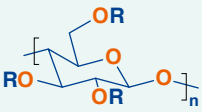


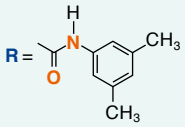
Ordering information

ID	Length → 200 mm	EC guard columns*
NUCLEODEX β-OH eluent methanol – 0.1 % TEAA pH 4 (55:45)		
Analytical EC columns		
4 mm	720124.40	721171.30
NUCLEODEX α-PM eluent methanol – 50 mmol/L phosphate pH 3 (70:30)		
Analytical EC columns		
4 mm	720127.40	721469.30
NUCLEODEX β-PM eluent methanol – 0.1 % TEAA pH 4 (65:35)		
Analytical EC columns		
4 mm	720125.40	721176.30
NUCLEODEX γ-PM eluent methanol – 0.1 % TEAA pH 4 (55:45)		
Analytical EC columns		
4 mm	720752.40	721178.30
NUCLEODEX CC screening kit		
contains one CC 30/4 each with NUCLEODEX β-OH, α-PM, β-PM and γ-PM as well as one CC column holder 30 mm	721920	

* EC 4/3 guard columns for EC columns with 4 mm ID require the Column Protection System guard column holder (REF 718966, see page 251). Columns and guard columns in packs of 1.

NUCLEOCEL *DELTA* enantiomer separation based on a cellulose derivative · USP L40





Technical data

- Base material silica, chiral selector cellulose tris-(3,5-dimethylphenylcarbamate)
  High resolution type (S) with 5 µm particle size, allows use of shorter columns (150 mm) for faster separations, pressure stability up to ~150 bar (2000 psi), pH stability 1–9

NUCLEOCEL *DELTA* for normal phase applications: eluent in column *n*-heptane – 2-propanol (90:10, v/v) typical eluents are heptane – propanol mixtures

NUCLEOCEL *DELTA*-RP for reversed phase applications: eluent in column acetonitrile – water (40:60, v/v) designed for use either in polar organic mode or with eluents containing high concentrations of chaotropic salts such as perchlorate

Recommended application

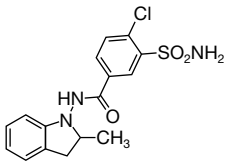
- Pharmaceutically active compounds, chiral pollutants (e.g., herbicides, PCB), chiral compounds in food (dyes, preservatives), chiral catalysts and bioorganic compounds

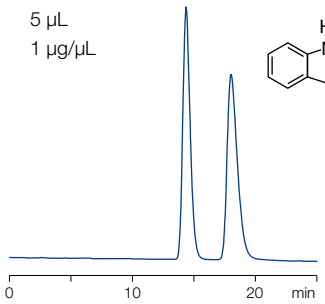
Similar phases: Chiralcel® OD, Kromasil® CelluCoat™, Eurocel® 01, Lux™ Cellulose-1

Enantiomer separation of indapamide

MN Appl. No. 121230

Column:	250 x 4,6 mm NUCLEOCEL *DELTA-RP S*
Eluent:	acetonitrile – water (40:60, v/v)
Flow rate:	0.5 mL/min
Temperature:	40 °C
Detection:	UV, 254 nm
Injection:	5 µL
Concentration:	1 µg/µL
α = 1.3	
R_s = 2.6	

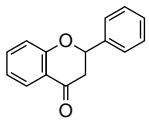


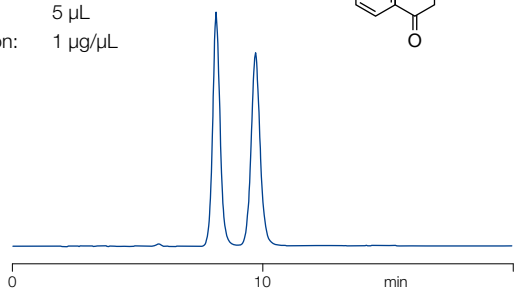


Enantiomer separation of flavanone

MN Appl. No. 121260

Column:	250 x 4,6 mm NUCLEOCEL *DELTA S*
Eluent:	*n*-heptane – 2-propanol (90:10, v/v)
Flow rate:	1 mL/min
Temperature:	25 °C
Detection:	UV, 254 nm
Injection:	5 µL
Concentration:	1 µg/µL
α = 1.29	
R_s = 2.6	





Ordering information

ID	Length → 150 mm	250 mm	EC guard columns*
NUCLEOCEL *DELTA S*, 5 µm eluent n-heptane – 2-propanol (90:10, v/v)			
Analytical EC columns			
4.6 mm		720445.46	721185.30
NUCLEOCEL *DELTA-RP S*, 5 µm eluent acetonitrile – water (40:60, v/v)			
Analytical EC columns			
4.6 mm	720451.46	720450.46	721186.30

* EC 4/3 guard column cartridges are used for EC columns of 4.6 mm ID with the Column Protection System guard column holder (REF 718966, see page 251). Columns and guard columns in packs of 1.

HPLC columns for enantiomer separations

RESOLVOSIL BSA-7 protein phase for enantiomer separation · USP L75

Technical data

- Base material NUCLEOSIL® silica, particle size 7 µm, pore size 300 Å chiral selector bovine serum albumin (BSA)
- Separation based on selective interaction of proteins with low molecular compounds, i.e. principles of bioaffinity, including hydrophobic interactions (similar to a true reversed phase), interactions of polar groups and steric effects

Recommended application

- Amino acid derivatives, aromatic amino acids, aromatic sulfoxides, barbiturates, benzodiazepinones, benzoin and benzoin derivatives, β-blockers, coumarin derivatives, and for monitoring stereoselective microbial and enzymatic conversions

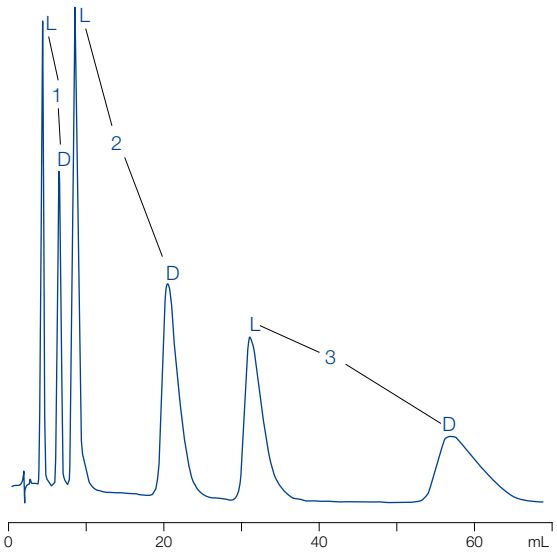
Enantiomer separation of *N*-benzoyl-*D*,*L*-amino acids

MN Appl. No. 105450

S. Allenmark et al. in "Affinity chromatography and biological recognition" (I. Chaiken, M. Wilchek, and I. Parikh. Eds.), Academic Press, New York, 1983, 259–260

Column: 150 x 4 mm RESOLVOSIL BSA-7
Eluent: 50 mmol/L phosphate buffer pH 6.5 + 1 % 1-propanol
Flow rate: 0.70 mL/min
Detection: UV, 225 nm

Peaks:
1. Serine
2. Alanine
3. Phenylalanine



Ordering information

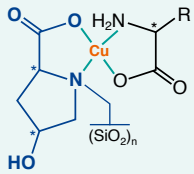
Eluent in column 0.1 mol/L phosphate buffer pH 7.5, 2 % 1-propanol

ID	Length → 150 mm	EC guard columns*
RESOLVOSIL BSA-7		
Analytical EC columns		
4 mm	720046.40	721402.30

* EC 4/3 guard columns for EC columns with 4 mm ID require the Column Protection System guard column holder (REF 718966, see page 251). Columns and guard columns in packs of 1.

HPLC columns for enantiomer separations

NUCLEOSIL® CHIRAL-1 enantiomer separation based on ligand exchange · USP L32



Technical data

- Base material NUCLEOSIL® silica, particle size 5 µm, pore size 120 Å chiral selector L-hydroxyproline – Cu^{2+} complexes
- Principal interaction mode:
- formation of ternary mixed-ligand complexes with Cu(II) ions; differences in the stability of the diastereomeric complexes cause chromatographic separation

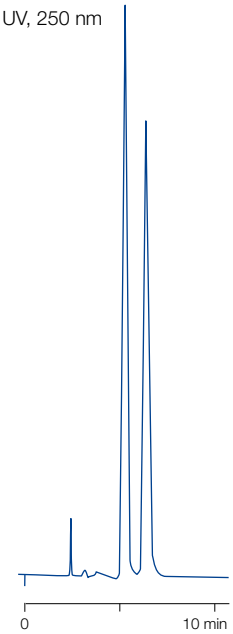
Recommended application

- Enantiomers with two polar functional groups with the correct spacing such as α-amino acids, α-hydroxycarboxylic acids (e.g., lactic acid), *N*-alkyl-α-amino acids etc.

D,L-alanine enantiomers

MN Appl. No. 105410

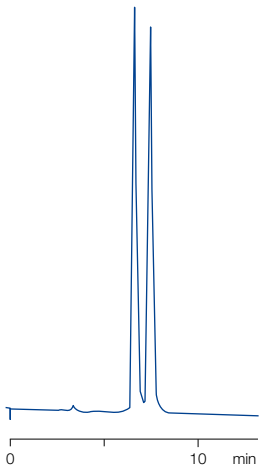
Column:	250 x 4 mm NUCLEOSIL® CHIRAL-1
Eluent:	0.5 mmol/L $CuSO_4$
Flow rate:	1 mL/min
Pressure:	60 bar
Temperature:	60 °C
Detection:	UV, 250 nm



D,L-threonine enantiomers

MN Appl. No. 105410

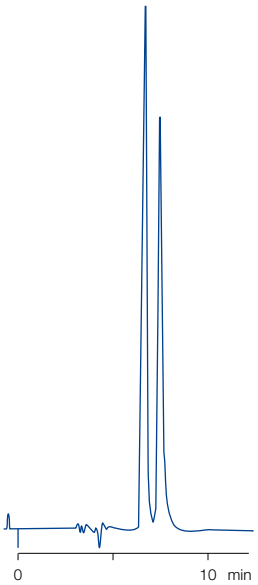
Column:	250 x 4 mm NUCLEOSIL® CHIRAL-1
Eluent:	0.25 mmol/L $CuSO_4$
Flow rate:	0.8 mL/min
Pressure:	65 bar
Temperature:	60 °C
Detection:	UV, 240 nm



Lactic acid enantiomers

MN Appl. No. 105560

Column:	250 x 4 mm NUCLEOSIL® CHIRAL-1
Eluent:	0.5 mmol/L $CuSO_4$
Flow rate:	0.8 mL/min
Temperature:	60 °C
Detection:	UV, 240 nm
Injection:	1 µL



Ordering information

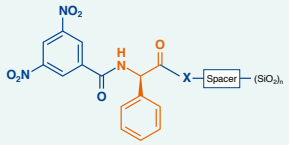
Eluent in column 0.5 mmol/L copper sulfate solution

ID	Length → 250 mm	EC guard columns*
NUCLEOSIL® CHIRAL-1		
Analytical EC columns		
4 mm	720081.40	721188.30

* EC 4/3 guard columns for EC columns with 4 mm ID require the Column Protection System guard column holder (REF 718966, see page 251). Columns and guard columns in packs of 1.

HPLC columns for enantiomer separations

NUCLEOSIL® CHIRAL-2 · CHIRAL-3 enantiomer separation in organic eluent systems · USP L36



Technical data

- Base material NUCLEOSIL® silica, particle size 5 µm, pore size 100 Å chiral selector for NUCLEOSIL® CHIRAL-2 is *N*-(3,5-dinitrobenzoyl)-D-phenylglycine, for CHIRAL-3 the optical antipode is used, "brush type" phases
- Principle interaction modes: charge-transfer interactions, hydrogen bonds, dipole-dipole interactions and steric effects

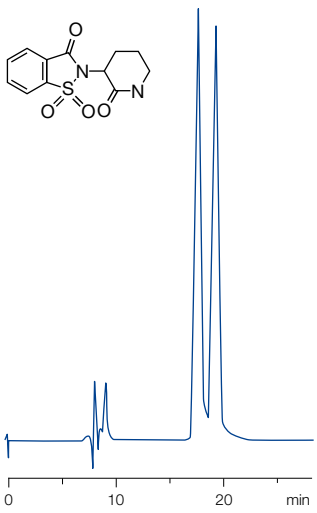
Recommended application

- analysis of stereoisomers such as separation of enantiomers and diastereomers, control of optical purity of plant protectives (pesticides, e.g., propionic acid derived herbicides) pharmaceuticals etc. and for product control in chiral organic syntheses
- For control of optical purity of a substance, the columns NUCLEOSIL® CHIRAL-2 and NUCLEOSIL® CHIRAL-3 allow to select conditions such that the minor enantiomer, present as an impurity, is eluted before the main peak. Overlapping peaks are avoided. This makes an exact quantification of the impurity much easier.

Enantiomer separation of *D*,*L*-supidimide

MN Appl. No. 105690

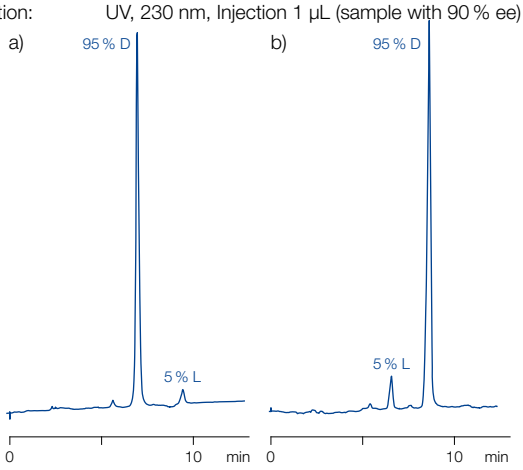
Column: 250 x 4 mm NUCLEOSIL® CHIRAL-2
Eluent: tetrahydrofuran – *n*-heptane (10:3, v/v)
Flow rate: 1.0 mL/min
Detection: UV, 220 nm



Control of optical purity of mecoprop methyl

MN Appl. No. 111360

Columns: a) 250 x 4 mm NUCLEOSIL® CHIRAL-2
b) 250 x 4 mm NUCLEOSIL® CHIRAL-3
Eluent: *n*-heptane – 2-propanol – TFA (100:0.05:0.05, v/v/v)
Flow rate: 1 mL/min, ambient temperature
Detection: UV, 230 nm, Injection 1 µL (sample with 90 % ee)



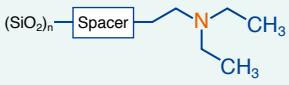
Ordering information

Eluent in column *n*-heptane – 2-propanol – TFAA (100:0.05:0.05, v/v/v)

ID	Length → 250 mm	EC guard columns*
NUCLEOSIL® CHIRAL-2		
Analytical EC columns		
4 mm	720088.40	721190.30
NUCLEOSIL® CHIRAL-3		
Analytical EC columns		
4 mm	720350.40	721190.30

Guard columns for NUCLEOSIL® CHIRAL-2 and CHIRAL-3 are identical.
* EC 4/3 guard columns for EC columns with 4 mm ID require the Column Protection System guard column holder (REF 718966, see page 251). EC columns and EC guard columns in packs of 1.

NUCLEOGEN® columns anion exchange chromatography of nucleic acids



NUCLEOGEN® 60-7 DEAE pore size 60 Å

Technical data

- Base material silica, particle size 7 µm; DEAE anion exchanger
- For the separation of oligonucleotides up to chain lengths of 40 bases with recoveries > 95 % capacity 200 A_{260}/mL (~ 300 A_{260} for a 125 x 4 mm ID column, 1875 A_{260} for a 125 x 10 mm ID column)
- Preparative separations possible when using higher flow rates and longer gradient times

NUCLEOGEN® 500-7 DEAE pore size 500 Å

Technical data

- Base material silica, particle size 7 µm; DEAE anion exchanger
- For the separation of tRNA, 5S RNA, viroids and messenger RNA in the intermediate molecular weight range (25–1 000 kDa) with recoveries > 95 %
- Capacity 730 A_{260} for a 125 x 6 mm ID column, 1940 A_{260} for a 125 x 10 mm ID column

NUCLEOGEN® 4000-7 DEAE pore size 4000 Å

Technical data

- Base material silica, particle size 7 µm; DEAE anion exchanger
- For the separation of plasmids, DNA restriction fragments, ribosomal RNA, messenger RNA and viral RNA, i.e. very high molecular weight nucleic acids (e.g., 1–50 MDa)
- Capacity 120 A_{260} for a 125 x 6 mm ID column, 350 A_{260} for a 125 x 10 mm ID column

For more separations of deoxyoligonucleotides, plasmids and DNA restriction fragments visit our website *www.mn-net.com/apps*

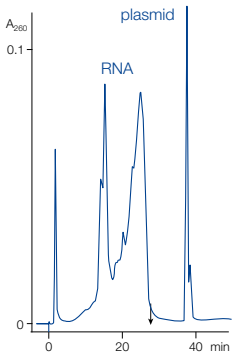
Separation of plasmid pBR 322

MN Appl. No. 107480

M. Colpan, D. Riesner, private communication

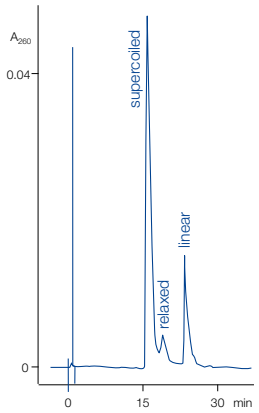
A) isolation of plasmid DNA from a crude cell lysate

Sample: 5 µg plasmid pBR 322 containing cleared lysate from *E. coli*
Column: 125 x 6 mm NUCLEOGEN® 4000-7 DEAE
Eluent: A) 20 mmol/L K phosphate buffer pH 6.9; 5 mol/L urea
B) eluent A + 1.5 mol/L KCl
20–100 % B in 50 min;
arrow = ionic strength of 850 mmol/L
Flow rate: 1.0 mL/min, 70 bar, ambient temperature
Detection: UV, 260 nm



B) separation of supercoiled plasmid from relaxed and linear forms

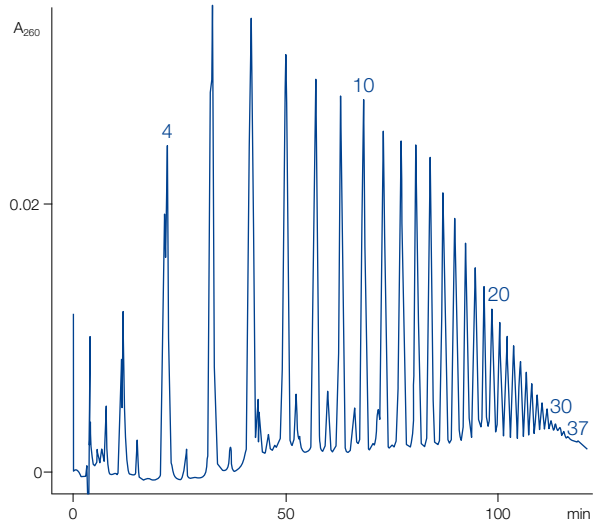
Sample: plasmid pBR 322, supercoiled, relaxed and linear
Column: 125 x 6 mm NUCLEOGEN® 4000-7 DEAE
Eluent: A) 20 mmol/L K phosphate buffer pH 6.8; 6 mol/L urea
B) eluent A + 2 mol/L KCl
42–100 % B in 230 min
Flow rate: 1.5 mL/min, 45 bar, ambient temperature



Separation of oligo(rA)$_n$

MN Appl. No. 115180

Column: 125 x 4 mm NUCLEOGEN® 60-7 DEAE
Eluent: A) 20 mmol/L phosphate buffer, pH 5.5, 5 mol/L urea
B) buffer A + 1 mol/L KCl
0–100 % B in 200 min
Flow rate: 2 mL/min
Pressure: 110 bar
Temperature: ambient
Detection: UV, 260 nm

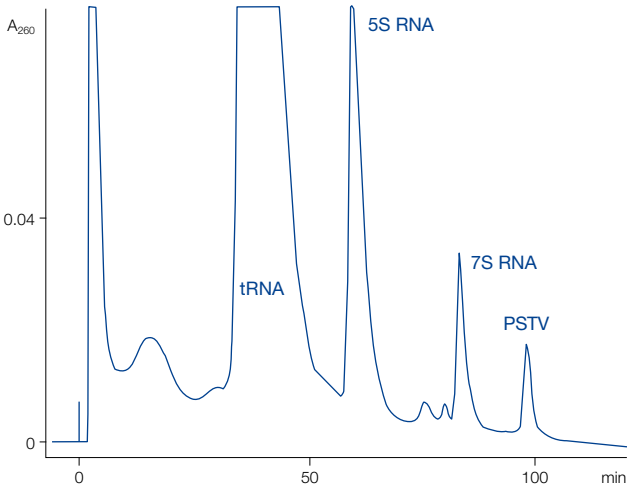


Preparative separation of a crude RNA extract of viroid (PSTV) infected tomato plants

MN Appl. No. 107490

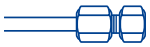
D. Riesner, BioEngineering 1 (1988) 42–48

Column: 125 x 6 mm NUCLEOGEN® 500-7 DEAE
Eluent: A) `250 mmol/L KCl, 20 mmol/L phosphate buffer, pH 6.6, 5 mol/L urea
B) 1 mol/L KCl, 20 mmol/L phosphate buffer, pH 6.6, 5 mol/L urea
0–50 % B in 120 min, 50–100 % B in 250 min
Flow rate: 3 mL/min
Pressure: 40 bar, ambient temperature
Detection: 260 nm



Ordering information

Eluent in column methanol

	ID	Length → 125 mm	Guard columns*
NUCLEOGEN® 60-7 DEAE particle size 7 µm, pore size 60 Å			
Analytical EC columns			
	4 mm	736596.40	736400.40
Preparative VarioPrep columns			
	10 mm	736597.100	736400.40
NUCLEOGEN® 500-7 DEAE particle size 7 µm, pore size 500 Å			
Analytical Valco type columns			
	6 mm	736598	736400.40
Preparative VarioPrep columns			
	10 mm	736599.100	736400.40
NUCLEOGEN® 4000-7 DEAE particle size 7 µm, pore size 4000 Å			
Analytical Valco type columns			
	6 mm	736601	736400.40
Preparative VarioPrep columns			
	10 mm	736602.100	736400.40

* NUCLEOGEN® guard columns are 30 mm long and require the CC column holder 30 mm (REF 721823).
Columns in packs of 1, guard columns in packs of 2.

HPLC columns for biochemical separations

NUCLEOGEL® SAX anion exchange of biological macromolecules · USP L23

Technical data

- Polymer-based strongly basic anion exchanger -$N^+(CH_3)_3$, gel matrix quaternized PEI; particle size 8 µm, pore size 1000 Å
- pH working range 1–13, max. working pressure 200 bar

Recommended application

- Purification of peptides, large proteins and oligonucleotides, high capacity for proteins even at pH 10

NUCLEOGEL® SCX cation exchange of biological macromolecules · USP L22

Technical data

- Polymer-based strongly acidic cation exchanger -SO_3^-, hydrophilic gel matrix; particle size 8 µm, pore size 1000 Å
- pH working range 1–13, max. working pressure 200 bar

Recommended application

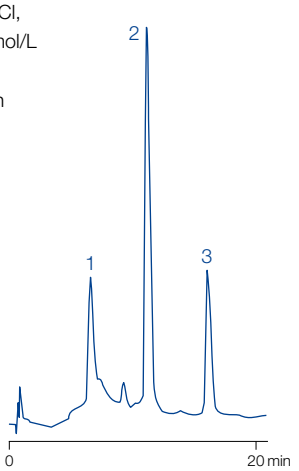
- Proteins, peptides and carbohydrates with high isoelectric point

Separation of hen's egg white

MN Appl. No. 115200

Sample:	frozen egg white was thawed, filtered and diluted 1:8 with eluent A
Column:	50 x 4.6 mm NUCLEOGEL® SAX 1000-8
Eluent:	A) 0.01 mol/L Tris-HCl, pH 7.5; B) A + 0.5 mol/L NaAc, pH 7.5; 0–100 % B in 20 min
Flow rate:	1 mL/min
Inj.volumen:	50 µL
Detection:	UV, 280 nm

Peaks:
1. Conalbumin
2. Ovalbumin
3. not identified

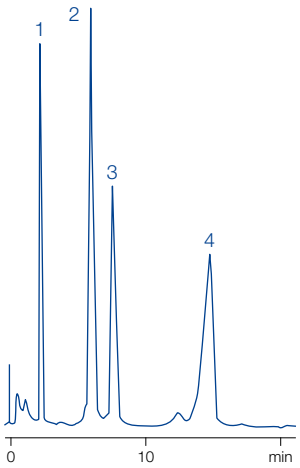


Separation of protein standards

MN Appl. No. 108261

Column:	50 x 4.6 mm NUCLEOGEL® SCX 1000-8
Eluent:	A) 0.02 mol/L KH_2PO_4, pH 6.0 B) A + 0,5 mol/L NaCl, pH 6.0 0–100 % B in 20 min
Flow rate:	1 mL/min
Detection:	UV, 280 nm

Peaks:
1. Myoglobin
2. α-Chymotrypsinogen A
3. Cytochrome C
4. Lysozyme



Ordering information

Eluent in column 0.1 mol/L Na_2SO_4 + 0.2 % NaN_3

ID	Length → 50 mm	Guard columns*
NUCLEOGEL® SAX pore size 1000 Å		
Analytical Valco type columns		
4.6 mm	719469	719600
NUCLEOGEL® SCX pore size 1000 Å		
Analytical Valco type columns		
4.6 mm	719475	719540

* NUCLEOGEL® SAX and SCX Valco type guard columns measure 5 x 3 mm and require the guard column holder B, REF 719539 (see page 250)
Columns in packs of 1, guard columns in packs of 2.

HPLC columns for biochemical separations

NUCLEODUR® 300 C_{18} ec · C_4 ec wide pore silica for biochromatography · USP L1 (C_{18}) · USP L26 (C_4)

Key feature

- Reliable wide pore RP phases for daily routine analysis
- Medium density octadecyl or butyl modification with exhaustive endcapping
- Ideal phases for separation of biomolecules

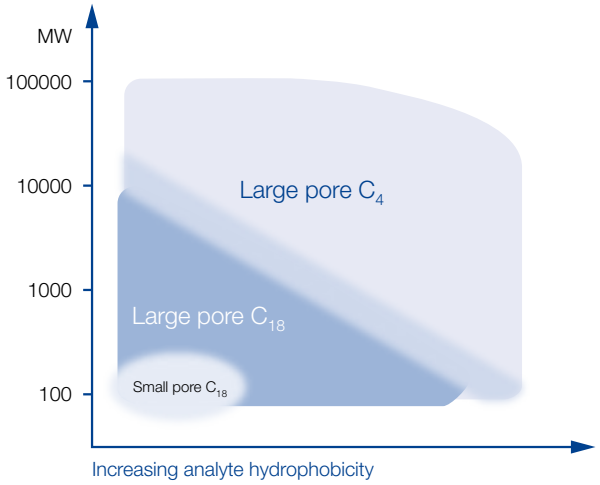
Technical data

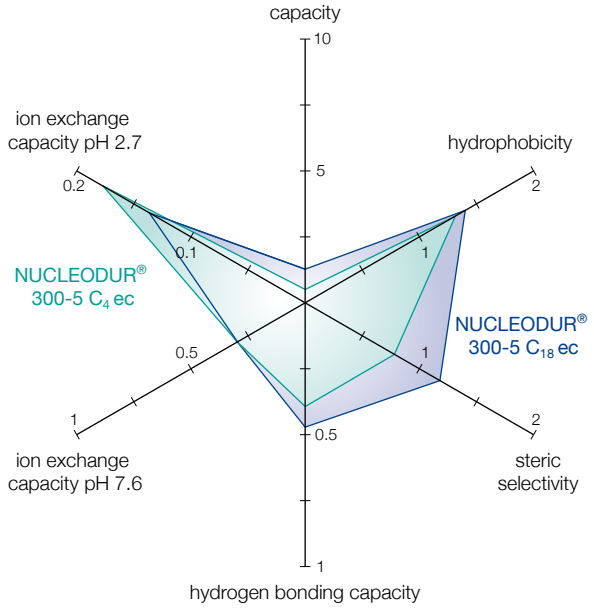
- Pore size 300 Å; particle size 5 µm, carbon content 4 % for C_{18}, 2.5 % for C_4; pH stability 1–9; high reproducibility from lot to lot

Recommended application

- Biological macromolecules like proteins or peptides

Column selection by analyte characteristics





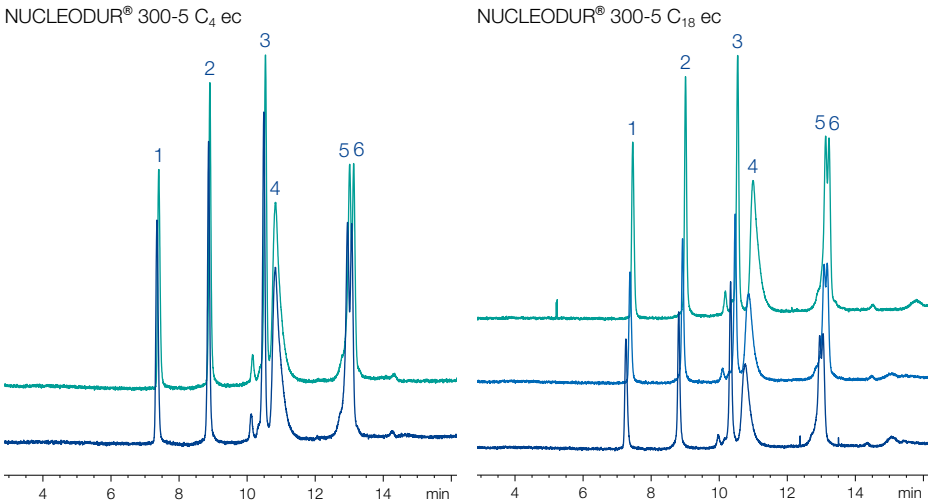
Tanaka plots of NUCLEODUR® wide pore phases

Batch-to-batch reproducibility of NUCLEODUR® 300-5 C_4 ec and NUCLEODUR® 300-5 C_{18} ec

MN Appl. Nos. 126551 / 126552

Columns:	250 x 4 mm
Eluent:	A) 0.1 % TFA in water
	B) 0.08 % TFA in acetonitrilel
	20–60 % B in 15 min
Flow rate:	1 mL/min
Temperature:	25 °C
Detection:	UV, 280 nm

Peaks:
1. Ribonuclease A
2. Cytochrome C
3. Lysozyme
4. BSA
5. β-Lactoglobulin
6. β-Lactoglobulin 2



HPLC columns for biochemical separations

Comparison of narrow and wide pore NUCLEODUR® for the separation of proteins

MN Appl. No. 126590

Columns: 250 x 4,6 mm NUCLEODUR® 300-5 C_{18} ec
250 x 4,6 mm NUCLEODUR® C_{18} Gravity, 5 µm

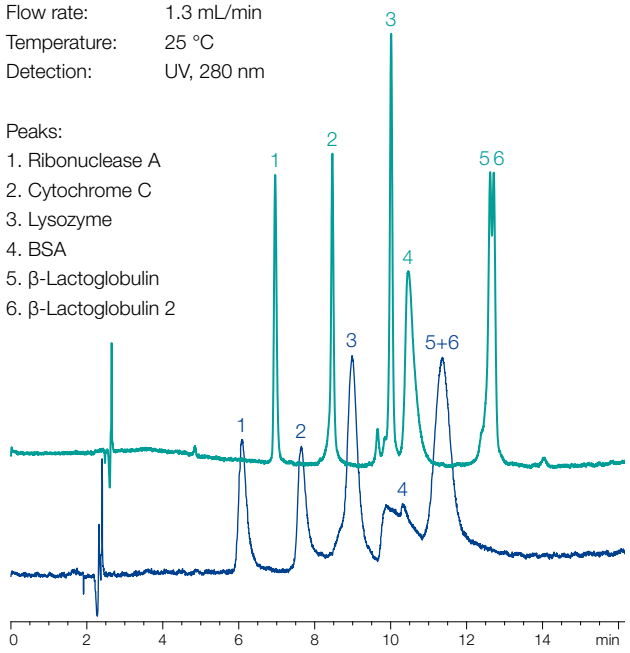
Eluent: A) 0.1 % TFA in water
B) 0.08 % TFA in acetonitrile
20–65 % B in 15 min
(3 min 65 % B)

Flow rate: 1.3 mL/min

Temperature: 25 °C

Detection: UV, 280 nm

Peaks:
1. Ribonuclease A
2. Cytochrome C
3. Lysozyme
4. BSA
5. β-Lactoglobulin
6. β-Lactoglobulin 2



Sharper peaks of larger molecules on wide pore material

Tryptic digest of cytochrome C

MN Appl. No. 126600

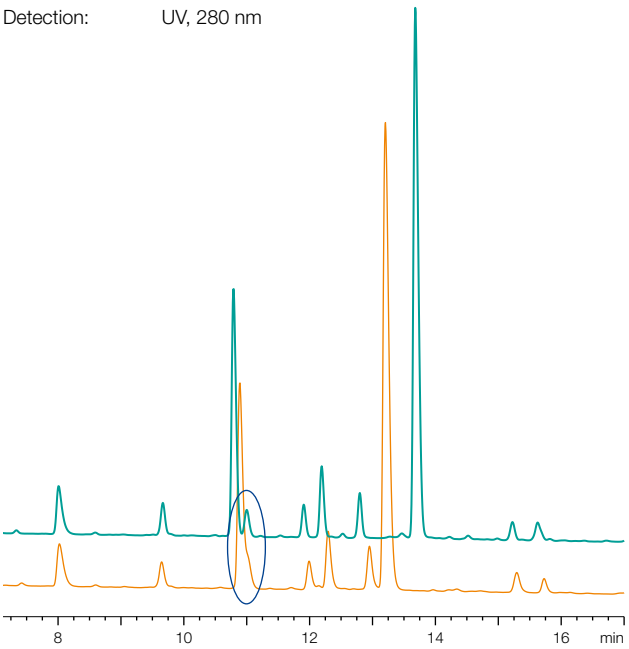
Columns: 250 x 4.6 mm NUCLEODUR® 300-5 C_{18} ec
250 x 4.6 mm Jupiter® C_{18}, 5 µm

Eluent: A) 0.1 % TFA in water
B) 0.08 % TFA in acetonitrile
5–40 % B in 15 min (1 min 40 % B)

Flow rate: 1.3 mL/min

Temperature: 30 °C

Detection: UV, 280 nm



Less tailing and better separation on NUCLEODUR® 300 C_{18} ec

Ordering information

Eluent in column acetonitrile – water

	ID	Length → 100 mm	125 mm	150 mm	250 mm	EC guard columns*
NUCLEODUR® 300-5 C_{18} ec octadecyl phase, particle size 5 µm, pore size 300 Å, endcapped, 4 % C						
Analytical EC columns						
	2 mm	760183.20	760184.20	760185.20	760186.20	761988.20
	3 mm	760183.30	760184.30	760185.30	760186.30	761988.30
	4 mm	760183.40	760184.40	760185.40	760186.40	761988.30
	4.6 mm	760183.46	760184.46	760185.46	760186.46	761988.30
NUCLEODUR® 300-5 C_4 ec butyl phase, particle size 5 µm, pore size 300 Å, endcapped, 2.5 % C						
Analytical EC columns						
	2 mm	760193.20	760194.20	760195.20	760196.20	761989.20
	3 mm	760193.30	760194.30	760195.30	760196.30	761989.30
	4 mm	760193.40	760194.40	760195.40	760196.40	761989.30
	4.6 mm	760193.46	760194.46	760195.46	760196.46	761989.30

* EC guard columns require the Column Protection System guard column holder (REF 718966, see page 251).
EC columns in packs of 1, guard columns in packs of 3.

NUCLEOSIL® MPN RP chromatography of biological macromolecules

NUCLEOSIL® 100-5 C_{18} MPN · USP L1

Key feature

- Octadecyl phase, particle size 5 µm; pore size 100 Å
- Dynamic protein binding capacity per g packing: 6 mg BSA, 110 mg cytochrome C
- pH working range 2–8, max. working pressure 250 bar

Technical data

- Silica-based reversed phase materials with monomerically bonded alkyl chains, brush type structure predominantly hydrophobic forces with a small portion of hydrophilic interactions
- Maximum separation efficiency can be achieved when the injected protein mass does not exceed 1–2 % of the maximum protein loading capacity.

NUCLEOSIL® 300-5 C_4 MPN · USP L26

Key feature

- Butyl phase, particle size 5 µm, pore size 300 Å
- Dynamic protein binding capacity per g packing: 14 mg BSA, 27 mg cytochrome C especially suited for the purification of larger, hydrophobic peptides and very different proteins
- pH working range 2–8, max. working pressure 250 bar

Technical data

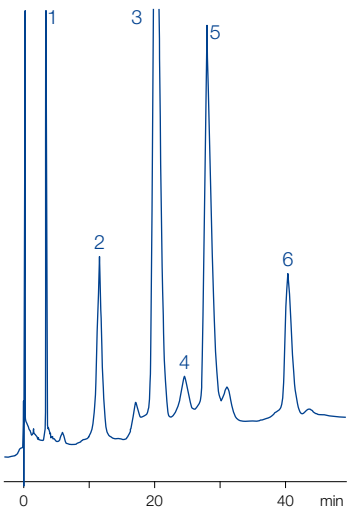
- Silica-based reversed phase materials with monomerically bonded alkyl chains, brush type structure predominantly hydrophobic forces with a small portion of hydrophilic interactions
- Maximum separation efficiency can be achieved when the injected protein mass does not exceed 1–2 % of the maximum protein loading capacity.

Separation of haemoglobin chains

MN Appl. No. 108240

Column:	250 x 4 mm NUCLEOSIL® 300-5 C_4 MPN
Eluent:	A) 20 % acetonitrile, 80 % water, 0.1 % TFA
	B) 60 % acetonitrile, 40 % water, 0.1 % TFA
	40–60 % B in 60 min
Flow rate:	1 mL/min
Detection:	UV, 220 nm

Peaks:
1. Hem
2. β-globin
3. α-globin
4. $^{A}\gamma^{T}$-globin
5. $^{G}\gamma$-globin
6. $^{A}\gamma^{I}$-globin



Ordering information

Eluent in column methanol

	ID	Length → 250 mm	EC guard columns*
NUCLEOSIL® 100-5 C_{18} MPN			
Analytical EC columns			
	4 mm	720231.40	
NUCLEOSIL® 300-5 C_4 MPN			
Analytical EC columns			
	4 mm	720245.40	721119.30

* EC guard columns require the Column Protection System guard column holder (REF 718966, see page 251). Columns in packs of 1, guard columns in packs of 2.

HPLC columns for biochemical separations

NUCLEOSIL® PPN RP chromatography of biological macromolecules

NUCLEOSIL® 100-5 C_{18} PPN · USP L1

Key feature

- Octadecyl phase, particle size 5 µm, pore size 100 Å, dynamic protein binding capacity per g packing: 8 mg BSA, 64 mg cytochrome C; suited for the separation of peptides and proteins up to about 40 kD, also suited for basic peptides

Technical data

- Silica-based reversed phase materials with polymerically bonded alkyl chains; exclusively hydrophobic interactions
- pH working range 1–9, max. working pressure 250 bar

NUCLEOSIL® 500-5 C_{18} PPN · USP L1

Key feature

- Octadecyl phase, particle size 5 µm, pore size 500 Å, dynamic protein binding capacity per g packing: 22 mg BSA, 40 mg cytochrome C; especially suited for large peptides and medium-size hydrophilic proteins

Technical data

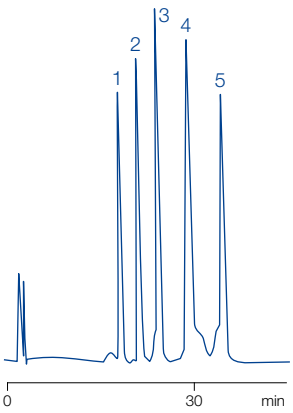
- Silica-based reversed phase materials with polymerically bonded alkyl chains; exclusively hydrophobic interactions
- pH working range 1–9, max. working pressure 250 bar

Separation of a protein standard

MN Appl. No. 108220

Column: 125 x 4 mm NUCLEOSIL® 100-5 C_{18} PPN
Eluent: A) 0.1 % TFA in H_2O
B) 0.08 % TFA in CH_3CN
20–60 % B in 10 min
Flow rate: 1.0 mL/min
Detection: UV, 280 nm

Peaks:
1. Ribonuclease
2. Cytochrome C
3. Lysozyme
4. β-Lactoglobulin
5. Ovalbumin

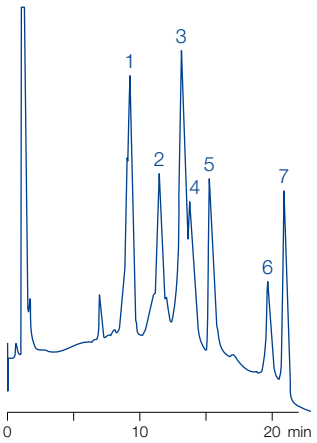


Separation of pancreatic secretion of piglets

MN Appl. No. 108280

Column: 125 x 4 mm NUCLEOSIL® 500-5 C_{18} PPN
Eluent: A) 0.1 % TFA in H_2O
B) 0.08 % TFA in CH_3CN
30–50 % B in 14 min, then 50–65 % B in 6 min
Flow rate: 1 mL/min
Detection: UV, 215 nm

Peaks:
1. Trypsin + trypsinogen
2. Proelastase
3. Lipase + α-Chymotrypsin
4. Chymotrypsinogen
5. α-Amylase
6., 7. Procarboxypeptidase



Ordering information

Eluent in column methanol

	ID	Length → 250 mm	EC guard columns*
NUCLEOSIL® 100-5 C_{18} PPN particle size 5 µm, pore size 100 Å			
Analytical EC columns			
	4 mm	720252.40	721567.30
NUCLEOSIL® 500-5 C_{18} PPN particle size 5 µm, pore size 500 Å			
Analytical EC columns			
	4 mm	720258.40	721924.30

* EC guard columns require the Column Protection System guard column holder (REF 718966, see page 251).
Columns in packs of 1, guard columns in packs of 2.

NUCLEOGEL® RP columns RP columns for biochemical applications · USP L21

Technical data

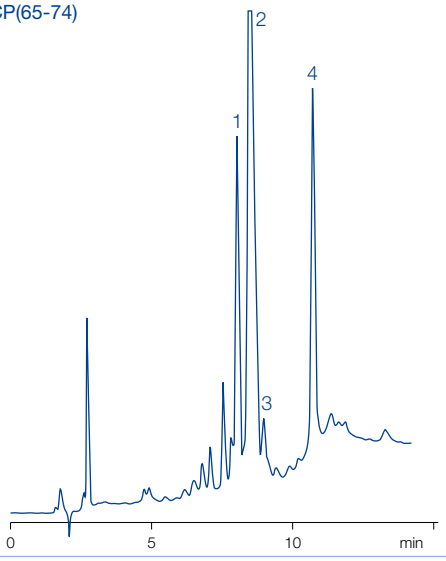
- Polystyrene resin cross-linked with divinylbenzene, available particle sizes 5 µm and 8 µm, available pore sizes 100 Å and 300 Å
- pH working range 1–13, max. working pressure 180 bar
- Small pore columns for reversed phase separation of small molecules such as pharmaceuticals with basic properties, e.g., organic heterocycles; also suited for separation of nucleosides and nucleotides up to 5000 Da; allow gradient as well as isocratic elution
- Wide pore columns are especially recommended for large biomolecules higher background hydrophobicity compared to silica phases

Analysis of the synthetic acyl carrier protein ACP(65-74)

MN Appl. No. 108500

Column: 150 x 4.6 mm NUCLEOGEL® RP 100-8
Eluant: A) 0.1 % TFA in acetonitrile – water (1:99, v/v)
B) 0.1 % TFA in acetonitrile – water (99:1, v/v)
10–60 % B in 20 min
Flow rate: 1 mL/min
Detection: UV, 220 nm

Peaks:
1. ACP(66-74)(H-Gln)
2. ACP(65-74)
3. ACP(66-74)(Glp)
4. Thioanisole



Ordering information

Eluent in column acetonitrile – water

ID	Length → 50 mm	150 mm	250 mm	Guard columns*
NUCLEOGEL® RP 100-5 particle size 5 µm, pore size 100 Å				
Analytical Valco type columns				
4.6 mm		719454	719455	719542
NUCLEOGEL® RP 100-8 particle size 8 µm, pore size 100 Å				
Analytical Valco type columns				
4.6 mm		719456	719520	719542
NUCLEOGEL® RP 300-5 particle size 5 µm, pore size 300 Å				
Analytical Valco type columns				
4.6 mm	719459			719542
NUCLEOGEL® RP 300-8 particle size 8 µm, pore size 300 Å				
Analytical Valco type columns				
4.6 mm	719460			719542

* Valco type guard columns measure 5 x 3 mm and require Guard column holder B, REF 719539, see page 250.
Columns in packs of 1, guard columns in packs of 2.

HPLC columns for sugar analyses

NUCLEOSIL® Carbohydrate separation of mono- and disaccharides · USP L8

Technical data

· Matrix: NUCLEOSIL® silica with amino modification, particle size 10 µm

Recommended application

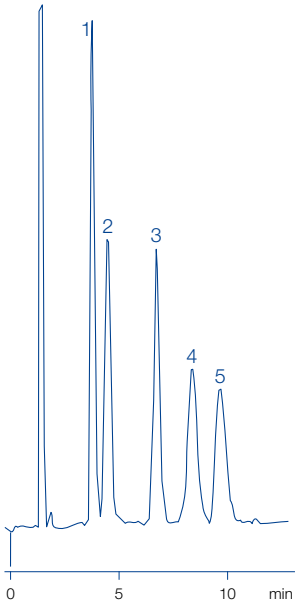
· RP separation of mono- and disaccharides

Separation of sugars

MN Appl. No. 102480

Column:	250 x 4 mm NUCLEOSIL® Carbohydrate
Eluent:	acetonitrile – water (79:21, v/v)
Flow rate:	2 mL/min
Temperature:	25 °C
Detection:	RI
Injection:	10 µL

Peaks:
1. Fructose
2. Glucose
3. Saccharose
4. Maltose
5. Lactose



Ordering information

Eluent in column acetonitrile – water (79:21, v/v)

ID	Length → 250 mm	EC guard columns*
NUCLEOSIL® Carbohydrate		
Analytical EC columns		
4 mm	720905.40	721170.30

* EC 4/3 guard columns for EC columns with 4 mm ID require the Column Protection System guard column holder (REF 718966, see page 251). Columns and guard columns in packs of 1.

NUCLEOGEL® SUGAR 810 separation of sugars · USP L17 (H-Form) · USP L19 (Ca form)

Technical data

- Sulfonated polystyrene - divinylbenzene resins in different ionic forms; due to a different selectivity pattern compared to NUCLEOGEL® SUGAR columns, the range of application is considerably enlarged
- Separation mechanism: ion exclusion, ion exchange, size exclusion, ligand exchange, NP and RP chromatography

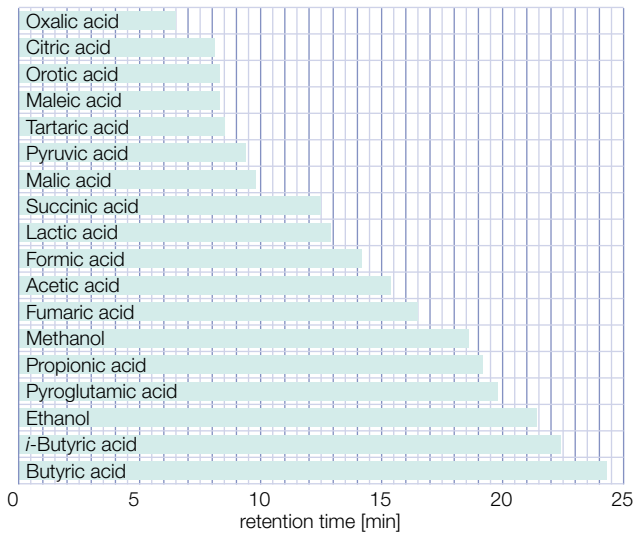
Recommended application

- H^+ form:
  Separation of sugars, sugar alcohols and organic acids; eluent in column 5 mmol/L H_2SO_4
- Ca^{2+} form:
  Separation of mono-, di- and oligosaccharides; eluent in column water

Organic acids and alcohols

MN Appl. No. 113870

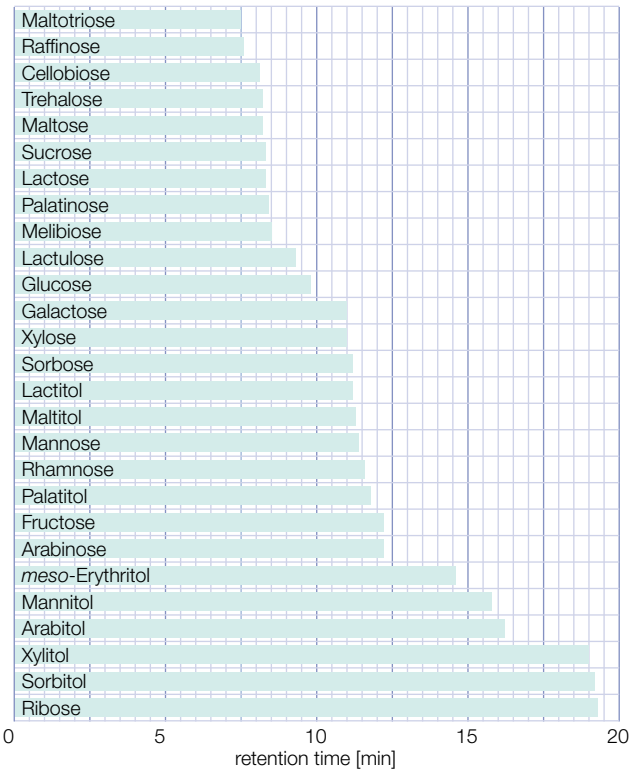
Column:	300 x 7.8 mm NUCLEOGEL® SUGAR 810 H
Eluent:	5 mmol/L H_2SO_4
Flow rate:	0.6 mL/min
Temperature:	35 °C
Detection:	RI
Injection:	5 µL



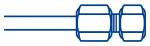
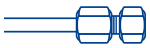
Sugars and sugar alcohols

MN Appl. No. 114160

Column:	300 x 7.8 mm NUCLEOGEL® SUGAR 810 Ca
Eluent:	water
Flow rate:	0.6 mL/min
Temperature:	85 °C
Detection:	RI



Ordering information

	ID	Length → 300 mm	Guard columns*
NUCLEOGEL® SUGAR 810 H eluent in column 5 mmol/L H_2SO_4			
Analytical Valco type columns			
	7.8 mm	719574	719575
NUCLEOGEL® SUGAR 810 Ca eluent in column water			
Analytical Valco type columns			
	7.8 mm	719570	719571

* NUCLEOGEL® SUGAR 810 guard columns measure 30 x 4 mm and require the CC column holder 30 mm (REF 721823)
Columns in packs of 1, guard columns in packs of 2.

HPLC columns for sugar analyses

NUCLEOGEL® ION 300 OA / SUGAR

separation of sugars · USP L17 (H form) · USP L19 (Ca form) · USP L34 (Pb form) · USP L58 (Na form)

Technical data

- Sulfonated spherical PS/DVB resins in different ionic forms; mean particle size 10 µm, pore size 100 Å
- Separation mechanism includes steric exclusion, ligand exchange and partition effects, ligand exchange being the predominant force, since the hydrated metal ions form strong interactions with the hydroxyl groups of the sample molecules. The intensity of these interactions decreases in the sequence Pb > Ca > Na
- Recommended operating temperatures: 60–95 °C; maximum pressure 70 bar

Recommended application

NUCLEOGEL® ION 300 OA:
H^+ form for separation of sugars, alcohols and organic acids

NUCLEOGEL® SUGAR:
Ca^{2+} form: separation of mono- and oligosaccharides, sugar alcohols

Pb^{2+} form: separation of mono- and disaccharides from food and biological samples

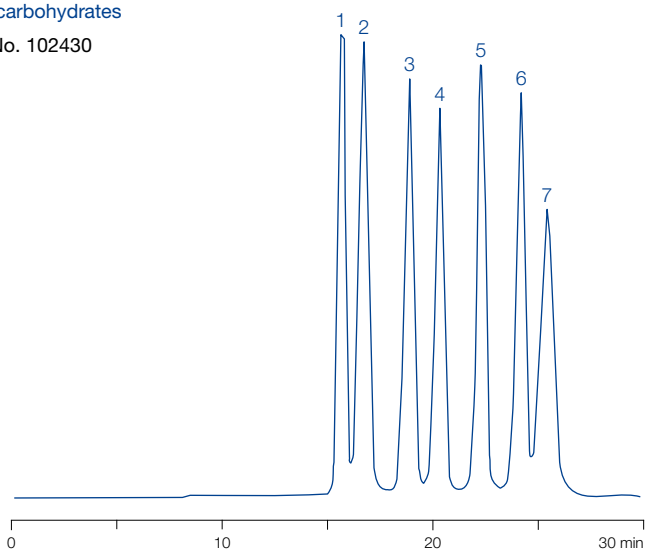
Na^+ form: separation of oligosaccharides from starch hydrolysates and food

Separation of carbohydrates

MN Appl. No. 102430

Column:	300 x 7.8 mm NUCLEOGEL® SUGAR Pb
Eluent:	deionized water
Flow rate:	0.4 mL/min
Temperature:	80 °C
Detection:	RI

Peaks:
1. Sucrose
2. Maltose
3. Glucose
4. Xylose
5. Galactose
6. Arabinose
7. Mannose



Ordering information

	ID	Length → 300 mm	Guard columns*
NUCLEOGEL® ION 300 OA eluent in column 5 mmol/L H_2SO_4 5 mmol/L H_2SO_4			
Analytical Valco type columns			
	7.8 mm	719501	719537
NUCLEOGEL® SUGAR Ca eluent in column water + 0.02 % azide			
Analytical Valco type columns			
	6.5 mm	719531	719535
NUCLEOGEL® SUGAR Pb eluent in column water + 0.02 % azide			
Analytical Valco type columns			
	7.8 mm	719530	719534
NUCLEOGEL® SUGAR Na eluent in column water + 0.02 % azide			
Analytical Valco type columns			
	7.8 mm	719532	719536

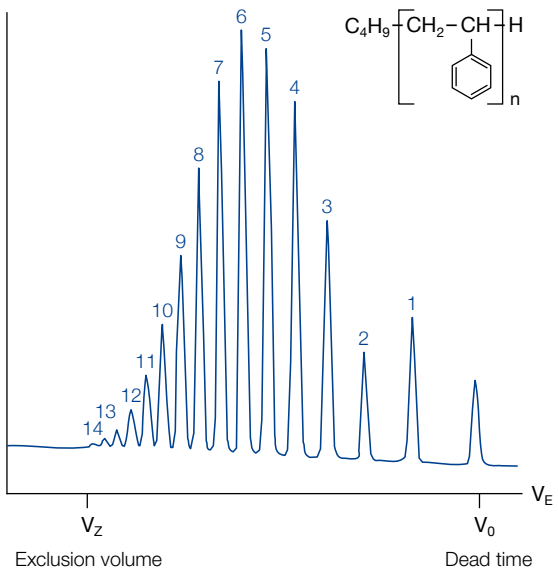
* Valco Type guard columns measure 21 x 4 mm and require the guard column holder C, REF 719538, see page 250.
Columns in packs of 1, guard columns in packs of 2.

NUCLEOGEL® GPC for GPC of water-insoluble substances

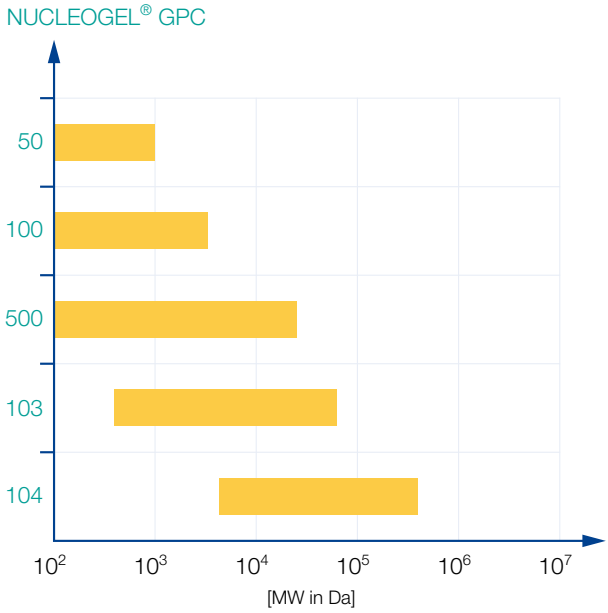
Technical data

- Highly crosslinked macroporous, spherical polystyrene – divinylbenzene polymer matrix with good mechanical stability

Chromatogram of styrene oligomers



Working ranges for polystyrene



Ordering information

Eluent in column toluene

Phase	Exclusion limit [kDalton]	Application	Column 300 x 7.7 mm
5 µm particle size			
Analytical Valco type columns			
NUCLEOGEL GPC 50	2	low molecular weight organics	719402
NUCLEOGEL GPC 100	4	oligomers, oils	719403
NUCLEOGEL GPC 500	25	low molecular weight polymers	719404
NUCLEOGEL GPC 103	60	low molecular weight polymers	719405
NUCLEOGEL GPC 104	500	polymers up to 500 kDa	719406
		guard columns 50 x 7.7 mm	719409
10 µm particle size			
Analytical Valco type columns			
NUCLEOGEL GPC 50	2	low molecular weight organics	719410
NUCLEOGEL GPC 100	4	oligomers, oils	719411
NUCLEOGEL GPC 500	25	low molecular weight polymers	719412
NUCLEOGEL GPC 103	60	low molecular weight polymers	719413
NUCLEOGEL GPC 104	500	polymers up to 500 kDa	719414
		guard columns 50 x 7.7 mm	719418

Columns and guard columns in packs of 1.

EC standard columns for analytical HPLC / UHPLC

- Analytical column system manufactured from stainless steel M8 outer threads on both ends combination of sealing element and very fine-meshed stainless steel screen, PTFE ring and fitting adaptor column heads SW 12, with inner threads M8 x 0.75 and UNF 10-32 (= 1/16“ connection)
- EC column hardware guarantees pressure stability of 1200 bar - hereby EC columns are suitable for UHPLC applications (ultra fast HPLC) and all modern HPLC systems.
- As screw-on guard column system we recommend the Column Protection System used with EC guard column cartridges with 4 mm length.
- EC guard columns supplied with NUCLEODUR®, NUCLEOSIL® spherical silicas and NUCLEOSHELL® spherical core shell silica particles

Available standard dimensions of EC columns

ID	Length → 20 mm	30 mm	50 mm	75 mm	100 mm	125 mm	150 mm	200 mm	250 mm	300 mm
2 mm	+	+	+	+	+	+	+	+	+	+
3 mm	+	+	+	+	+	+	+	+	+	+
4 mm	+	+	+	+	+	+	+	+	+	+
4.6 mm	+	+	+	+	+	+	+	+	+	+

Please ask for availability of certain phases.

Note: NUCLEODUR® and NUCLEOSHELL® column head must not be removed!

Guard columns for EC columns

EC column with ID	EC guard column*
2 mm	4/2
3 mm	4/3
3 mm	4/3
3 mm	4/3

Packs of 3 cartridges

* Information about the Column Protection System on page 251.

For preparative applications MN offers the so-called VarioPrep® hardware system, which is described from page 252 on.

Valco type columns

- Analytical column system manufactured from stainless steel
- Available inner diameters: 4.6 mm ID (1/4“ OD) and 7.7 mm (3/8“ OD)
- Mainly used for NUCLEOGEN® and NUCLEOGEL® (see page 226)

Ordering information

Description	Pack of	REF
Accessories for Valco type columns		
Guard column holder B for VA columns 5 x 3 mm	1	719539
Guard column holder C for VA guard columns 21 x 4 mm	1	719538

Column Protection System

Innovative and universal guard column holder system

- Suitable for all analytical HPLC columns with 1/16” fittings
- Cartridges filled with special NUCLEODUR®, NUCLEOSIL® and NUCLEOSHELL® HPLC adsorbents
- Ideal protection for your analytical main column → significant increase in column lifetime
- Minimized dead volume → suitable also for ultra-fast HPLC
- Special ferrules → pressure stability up to 1300 bar (18 850 psi)
- Visual contamination check → in-time changing of the guard column
- Suitable guard columns with 4 mm length, 2 mm ID (for main columns with 2 mm ID); 3 mm ID (for main columns with 3, 4 and 4.6 mm), respectively
- UNIVERSAL RP guard columns suitable for all HPLC columns under RP conditions

Content of the Column Protection System

Description	Pack of	REF
Guard column holder	1	718966
Capillaries (0.12 mm ID)	2	
Ferrules	3	
Wrenches	2	
Manual	1	

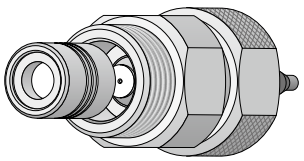
Ordering information

Description	Pack of	REF
Replacement parts for the Column Protection System		
Special ferrules made of PEEK	5	718967
Replacement connector including O-ring	1	718968
Stainless steel capillaries 0.12 mm ID, nuts and metal ferrules	3	718969
Stainless steel capillaries 0.18 mm ID (for higher flow rates), nuts and metal ferrules	3	718971
Wrench (size 12 and 14 mm)	1	718970
EC 4/2 UNIVERSAL RP guard column (for main columns with 2 mm ID)	3	728777.20
EC 4/2 UNIVERSAL RP guard column (for main columns with 2 mm ID), value pack	9	728778.20
EC 4/3 UNIVERSAL RP guard column (for main columns with 3, 4 and 4.6 mm ID)	3	728777.30
EC 4/3 UNIVERSAL RP guard column (for main columns with 3, 4 and 4.6 mm ID), value pack	9	728778.30

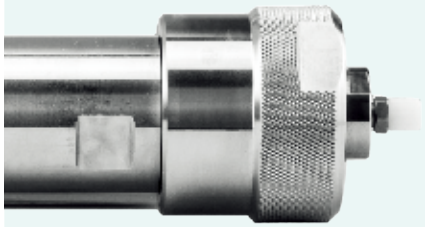
Visual contamination check

The cartridge is fitted with a special filter membrane:

- If this silver membrane is contaminated (bright or dark discoloration), it is advisable to replace the cartridge.
- If the contaminants are colorless, replace the cartridge if the pressure rises or the chromatographic performance decreases.

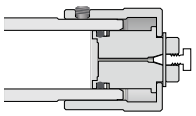


VarioPrep (VP) columns for preparative HPLC



- Column system for preparative HPLC, manufactured from stainless steel with two adjustable end fittings, suitable for frequent use of back-flushing techniques
- Allows compensation of a dead volume, which could occur at the column inlet after some time of operation, without need for opening the column
- Can be packed with all NUCLEODUR® and NUCLEOSIL® spherical silicas

Available standard dimensions of VarioPrep columns with axially adjustable end fittings

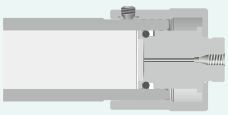


End fitting design	ID	Length → 10* mm	15* mm	Length → 50 mm	75 mm	100 mm	125 mm	150 mm	250 mm	500 mm
	8	+		+		+	+	+	+	
	10			+		+	+	+	+	
	16	+		+		+	+	+	+	
	21			+	+	+	+	+	+	
	32		+			+		+	+	
	40			+		+	+	+	+	+
	50		+			+		+	+	
	80								+	+

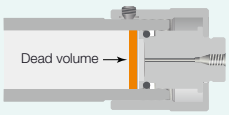
* 10 x 8, 10 x 16, 15 x 32 and 15 x 50 mm ID columns are used as guard columns and require the respective holders, see page 253.

The VarioPrep principle

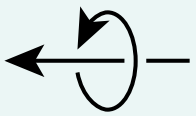
1



2

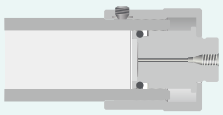


3



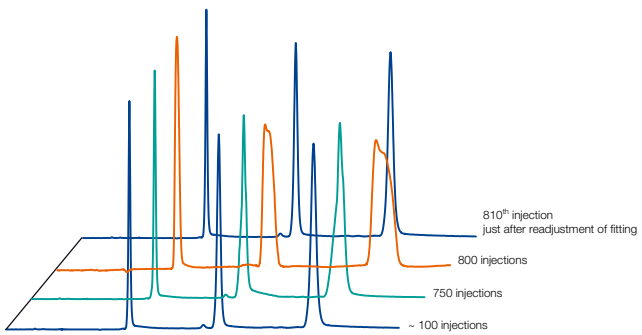
Readjustment of fitting

4



VarioPrep columns are produced with highest packing quality and bed density (1). Due to intensive chemical and/or mechanical exposure of the column adsorbent, shrinking of the column bed can occur (2; orange gap). in this even unlikely case readjustment of the VarioPrep column fitting (3; turning the nut at the column inlet clockwise) will eliminate the emerged dead volume (4). The performance of the VarioPrep column is completely reconstituted and column lifetime is significantly extended.

Column reconstitution



Reconstitution of VarioPrep column performance

- Slight peak broadening and deformation after 800 injections under strongly demanding conditions (pH 11; 50 °C; sample in DMSO)
- Readjustment of the column fitting restores column performance and prolongs column lifetime noticeably.

The improved guard column system for (semi-) preparative HPLC

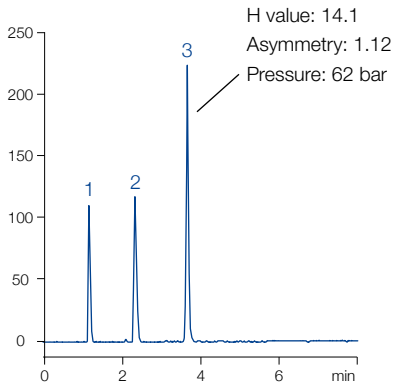
① VP 15/32 for 32 and 40 mm ID columns
② VP 10/16 for 16 and 21 mm ID columns
③ VP 10/8 for 8 and 10 mm ID columns
④ VP 15/50 for ≥ 50 mm ID columns

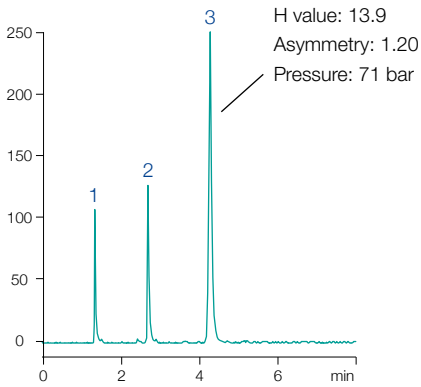
- Easy handling and cartridge exchange
- Robust hardware
- Free rotary plunger fittings – low O-ring abrasion
- Cost-efficient cartridges
- Minimally invasive / no disturbance of the separation efficiency of main column
- Low back pressure
- Designed for pressures up to 400 bar

Column performance without and with guard column

Columns: 125 x 16 mm NUCLEODUR® C_{18} HTec, 5 µm
125 x 16 mm NUCLEODUR® C_{18} HTec, 5 µm + 10 x 16 mm NUCLEODUR® C_{18} HTec guard column
Eluent: acetonitrile – water (80:20, v/v)
Flow rate: 16 mL/min
Temperature: 22 °C

Peaks:
1. Phenol
2. Naphthalene
3. Anthracene





Using VarioPrep guard columns provides ideal protection of your main column – symmetry, pressure and retention stay almost constant.

Technical data

· 1/16“ thread · free rotary plunger fittings – low O-ring abrasion · stainless steel

Guard cartridge	Holder REF	Holder ID	Recommended for column ID	Preferred capillary ID	Typical flow rate
VP 10/8	718251	8 mm	8 and 10 mm ID	0.17 and 0.25 mm	1–12 mL/min
VP 10/16	718256	16 mm	16 and 21 mm ID	0.17, 0.25 and 0.5 mm	2–32 mL/min
VP 15/32	718253	32 mm	32 and 40 mm ID	0.25, 0.5 and 1.0 mm	5–150 mL/min
VP 15/50	718255	50 mm	≥ 50 mm ID	0.5 and 1.0 mm	20–250 mL/min

Ordering information

Guard column holders for VarioPrep columns

	VP Guard columns for VarioPrep columns with ID →				Pack of	Replacement O-ring	Holder	
	8, 10 mm	16, 21 mm	32, 40 mm	≥ 50 mm	guard columns	(pack of 2)	ID	REF
VP	10/8				2	718975	8 mm	718251
VP		10/16			2	718976	16 mm	718256
VP			15/32		1	718977	32 mm	718253
VP				15/50	1	718978	50 mm	718255

For REF numbers of individual VP guard column cartridges see respective NUCLEODUR® and NUCLEOSIL® phases.

Accessories

Accessories for stainless steel HPLC columns

- Stainless steel columns are most frequently used in HPLC.
- The material is corrosion resistant, pressure stable and easy to work mechanically.

Ordering information

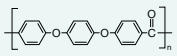
Description	Pack of	REF
Capillary accessories		
1/16“ column end caps (plastic)	4	718582
1/16“ nut for connecting 1/16“ capillaries	5	718583
1/16“ ferrule	5	718584
Capillary unions		
Typ 1: 100 mm x 1/16“ x 0.25 mm	1	718637
Typ 2: 100 mm x 1/16“ x 0.12 mm	1	719489
Cutter for 1/16“ capillary tubing	1	706290

For accessories and replacement parts for EC columns see page 251, for accessories and replacement parts for VarioPrep columns see page 253.

SPE accessories for sample preparation, like e.g., CHROMABOND® vacuum manifolds can be found on page 65.

PEEK accessories

PEEK



- PEEK (= polyether ether ketone) is a high performance polymer belonging to the group of polyarylether ketones (PAEK), which meets all requirements of HPLC columns with respect to chemical resistance and mechanical stability. In some fields of application in HPLC like, e.g., in ion chromatography and chromatography of biopolymers, PEEK fulfils the requirements for a nonmetallic material.
- All fittings can be tightened by hand.

Ordering information

Description	Pack of	REF
PEEK fittings		
1/16“ PEEK fingertight fitting, 1-part combination nut + ferrule	1	718770
1/16“ PEEK fingertight Nut	1	718771
1/16“ PEEK ferrule for REF 718771	1	718772
1/16“ PEEK double ferrule	1	718775

1/16“ PEEK union, both sides inner threads, equipped with 2 fingertight nuts and double ferrules	1	718766

1/16“ PEEK union, both sides inner threads, however without nuts and without ferrules	1	718767

1/16“ PEEK union, both sides outer threads	1	718768

AD	ID [mm]	Length	Pack of	REF
PEEK standard capillaries				
1/16“	0.13	1 m	1	718765
1/16“	0.17	1 m	1	718760
1/16“	0.25	1 m	1	718761
1/16“	0.5	1 m	1	718762
1/16“	0.75	1 m	1	718763

Description	Pack of	REF
Tools for PEEK capillaries		
Guillotine cutter for PEEK and PTFE capillaries	1	718769

Clean-Cut cutter for different capillary outer diameters	1	718755

NUCLEODUR® high purity silica for HPLC

Basics of preparative HPLC

In principal for preparative HPLC the same rules apply than for analytic HPLC. However both differ significantly in their aim. The aim of analytic HPLC is a preferably complete separation of the single components of a mixture with subsequent peak identification. In contrast the goal of preparative HPLC is isolation of the desired product in defined purity, maximum amount while having a cost effective method of operating.

Demand of a preparative separation

- Throughput
- Purity
- Yield

Upscaling table for current MN column dimensions

ID x Length [mm]	4 x 250	8 x 250	10 x 250	16 x 250	21 x 250	32 x 250	40 x 250	50 x 250	80 x 250
Linear scale-up factor	1	4	6.25	16	27.6	64	100	156.3	400
Typical amount of sample* [mg]	0.02–2	0.08–8	0.13–13	0.3–35	0.6–60	1.3–130	2–210	3–350	10–850
Typical flow rate [mL/min]	0.5–1.5	2–6	3–9	8–24	14–40	32–96	50–150	80–250	200–600

* based on RP material; the herein stated maximum amounts of sample are dependent on the separation problem and the sample. In some cases half the maximum amount of sample can already lead to a drastic overload of the column, in other cases the maximum amount of sample still leads to an acceptable separation.

NUCLEODUR® bulk packings

- Fully spherical high purity silica
- Pore size 110 Å; pore volume 0.9 mL/g; surface (BET) 340 m^2/g; density 0.47 g/mL; pressure stable up to 600 bar
- Bigger particles for preparative application

Ordering information

Phase	Endcapped	Carbon content	Particle size	Pack of 100 g	Pack of 1000 g
NUCLEODUR® C_{18} HTec premium octadecyl phase (see page 178)					
NUCLEODUR® C_{18} HTec, 7 µm	yes	18 % C	7 µm	713831.0100	713831.1
NUCLEODUR® C_{18} HTec, 10 µm	yes	18 % C	10 µm	713832.0100	713832.1
NUCLEODUR® C_{18} ec standard octadecyl phase (see page 181)					
NUCLEODUR® 100-10 C_{18} ec	yes	17.5 % C	10 µm	713611.0100	713611.1
NUCLEODUR® 100-12 C_{18} ec	yes	17.5 % C	12 µm	713618.0100	713618.1
NUCLEODUR® 100-16 C_{18} ec	yes	17.5 % C	16 µm	713621.0100	713621.1
NUCLEODUR® 100-20 C_{18} ec	yes	17.5 % C	20 µm	713601.0100	713601.1
NUCLEODUR® 100-30 C_{18} ec	yes	17.5 % C	30 µm	713631.0100	713631.1
NUCLEODUR® 100-50 C_{18} ec	yes	17.5 % C	50 µm	713550.0100	713550.1
Unmodifiziertes NUCLEODUR® SiOH silica (see page 190)					
NUCLEODUR® 100-10			10 µm	713610.0100	713610.1
NUCLEODUR® 100-12			12 µm	713615.0100	713615.1
NUCLEODUR® 100-16			16 µm	713620.0100	713620.1
NUCLEODUR® 100-20			20 µm	713600.0100	713600.1
NUCLEODUR® 100-30			30 µm	713630.0100	713630.1
NUCLEODUR® 100-50			50 µm	713551.0100	713551.1

POLYGOSIL® bulk packings

- Irregular silica for analytical applications
- pH stability 2–8

Physical properties of unmodified POLYGOSIL® materials

Phase	Pore size	Pore volume	Surface (BET)	Density	Pressure stability
POLYGOSIL® 60	60 Å	0.75 mL/g	350 m^2/g	0.45 g/mL	600 bar
POLYGOSIL® 100	100 Å	1 mL/g	280 m^2/g	0.35 g/mL	400 bar
POLYGOSIL® 300	300 Å	0.8 mL/g	100 m^2/g	0.45 g/mL	400 bar
POLYGOSIL® 1000	1000 Å	0.8 mL/g	25 m^2/g	0.45 g/mL	300 bar

Modification of POLYGOSIL® follows the same processes as for NUCLEOSIL® silica.

Ordering information

Phase	Endcapped	Carbon content	Pore size	Particle size	Pack of 10 g	Pack of 100 g
Octadecyl phases $-(CH_2)_{17}-CH_3$						
POLYGOSIL® 60-5 C_{18}	yes	12 % C	60 Å	5 µm	711330.10	711330.100
POLYGOSIL® 60-7 C_{18}	yes	12 % C	60 Å	7 µm	711340.10	711340.100
POLYGOSIL® 60-10 C_{18}	yes	12 % C	60 Å	10 µm	711350.10	711350.100
POLYGOSIL® 100-5 C_{18}	yes	14 % C	100 Å	5 µm	711560.10	711560.100
POLYGOSIL® 100-7 C_{18}	yes	14 % C	100 Å	7 µm	711570.10	711570.100
POLYGOSIL® 100-10 C_{18}	yes	14 % C	100 Å	10 µm	711580.10	711580.100
POLYGOSIL® 300-7 C_{18}	yes	4 % C	300 Å	7 µm	711710.10	711710.100
POLYGOSIL® 1000-7 C_{18}	yes	~ 1 % C	1000 Å	7 µm	711992.10	711992.100
Octyl phases $-(CH_2)_7-CH_3$						
POLYGOSIL® 60-5 C_8	no	7 % C	60 Å	5 µm	711300.10	711300.100
POLYGOSIL® 60-7 C_8	no	7 % C	60 Å	7 µm	711310.10	711310.100
POLYGOSIL® 60-10 C_8	no	7 % C	60 Å	10 µm	711320.10	711320.100
Butyl phases $-(CH_2)_3-CH_3$						
POLYGOSIL® 300-7 C_4	yes	~ 1 % C	300 Å	7 µm	711680.10	711680.100
POLYGOSIL® 1000-7 C_4	yes	< 1 % C	1000 Å	7 µm	711991.10	711991.100
Cyano phases (nitrile) $-(CH_2)_3-CN$						
POLYGOSIL® 60-5 CN		~ 5 % C	60 Å	5 µm	711380.10	711380.100
POLYGOSIL® 60-10 CN		~ 5 % C	60 Å	10 µm	711390.10	711390.100
Amino phases $-(CH_2)_3-NH_2$						
POLYGOSIL® 60-5 NH_2		~ 3 % C	60 Å	5 µm	711360.10	711360.100
POLYGOSIL® 60-10 NH_2		~ 3 % C	60 Å	10 µm	711370.10	711370.100
Dimethylamino phases $-(CH_2)_3-N(CH_3)_2$						
POLYGOSIL® 60-5 $N(CH_3)_2$		~ 3.5 % C	60 Å	5 µm	711420.10	711420.100
POLYGOSIL® 60-10 $N(CH_3)_2$		~ 3.5 % C	60 Å	10 µm	711430.10	711430.100
Unmodified silica SiOH						
POLYGOSIL® 60-5			60 Å	5 µm	711010.10	711010.100
POLYGOSIL® 60-7			60 Å	7 µm	711280.10	711280.100
POLYGOSIL® 60-10			60 Å	10 µm	711020.10	711020.100
POLYGOSIL® 100-5			100 Å	5 µm	711510.10	711510.100
POLYGOSIL® 100-7			100 Å	7 µm	711520.10	711520.100
POLYGOSIL® 100-10			100 Å	10 µm	711530.10	711530.100
POLYGOSIL® 300-7			300 Å	7 µm	711600.10	711600.100
POLYGOSIL® 1000-7			1000 Å	7 µm	711890.10	711890.100

POLYGOPREP irregular silica for HPLC

POLYGOPREP bulk packings

- Irregular silica for preparative applications
- pH stability 2–8

Physical properties of unmodified POLYGOPREP materials

Phase	Pore size	Pore volume	Surface (BET)	Density	Pressure stability
POLYGOPREP 60	60 Å	0.75 mL/g	350 m^2/g	0.45 g/mL	600 bar
POLYGOPREP 100	100 Å	1 mL/g	280 m^2/g	0.35 g/mL	400 bar
POLYGOPREP 300	300 Å	0.8 mL/g	100 m^2/g	0.45 g/mL	400 bar
POLYGOPREP 1000	1000 Å	0.8 mL/g	35 m^2/g	0.45 g/mL	300 bar

Modification of POLYGOPREP follows the same processes as for NUCLEOSIL® silica.

Ordering information

Phase	Endcapped	Carbon content	Pore size	Particle size	Pack of 100 g	Pack of 1 kg
Octadecyl phases $-(CH_2)_{17}-CH_3$						
POLYGOPREP 60-12 C_{18}	no*	12 % C	60 Å	10–15 µm	711009.100	711009.1000
POLYGOPREP 60-20 C_{18}	no*	12 % C	60 Å	15–25 µm	711031.100	711031.1000
POLYGOPREP 60-30 C_{18}	no*	12 % C	60 Å	25–40 µm	711480.100	711480.1000
POLYGOPREP 60-50 C_{18}	no*	12 % C	60 Å	40–63 µm	711500.100	711500.1000
POLYGOPREP 60-80 C_{18}	no*	12 % C	60 Å	63–100 µm	711011.100	711011.1000
POLYGOPREP 60-130 C_{18}	no*	12 % C	60 Å	63–200 µm	711590.100	711590.1000
POLYGOPREP 100-12 C_{18}	no*	14 % C	100 Å	10–15 µm	711018.100	711018.1000
POLYGOPREP 100-20 C_{18}	no*	14 % C	100 Å	15–25 µm	711019.100	711019.1000
POLYGOPREP 100-30 C_{18}	no*	14 % C	100 Å	25–40 µm	711032.100	711032.1000
POLYGOPREP 100-50 C_{18}	no*	14 % C	100 Å	40–63 µm	711021.100	711021.1000
POLYGOPREP 300-12 C_{18}	yes	4 % C	300 Å	10–15 µm	711024.100	711024.1000
POLYGOPREP 300-20 C_{18}	yes	4 % C	300 Å	15–25 µm	711025.100	711025.1000
POLYGOPREP 300-30 C_{18}	yes	4 % C	300 Å	25–40 µm	711720.100	711720.1000
POLYGOPREP 300-50 C_{18}	yes	4 % C	300 Å	40–63 µm	711730.100	711730.1000
POLYGOPREP 1000-30 C_{18}	yes	~ 1 % C	1000 Å	25–40 µm	711028.100	711028.1000
POLYGOPREP 1000-50 C_{18}	yes	~ 1 % C	1000 Å	40–63 µm	711029.100	711029.1000
Octyl phases $-(CH_2)_7-CH_3$						
POLYGOPREP 60-12 C_8	no*	7 % C	60 Å	10–15 µm	711007.100	711007.1000
POLYGOPREP 60-20 C_8	no*	7 % C	60 Å	15–25 µm	711008.100	711008.1000
POLYGOPREP 60-30 C_8	no*	7 % C	60 Å	25–40 µm	711470.100	711470.1000
POLYGOPREP 60-50 C_8	no*	7 % C	60 Å	40–63 µm	711490.100	711490.1000
* On request, these POLYGOPREP RP phases can be endcapped at surcharge.						
Butyl phases $-(CH_2)_3-CH_3$						
POLYGOPREP 300-12 C_4	yes	~ 1 % C	300 Å	10–15 µm	711022.100	711022.1000
POLYGOPREP 300-20 C_4	yes	~ 1 % C	300 Å	15–25 µm	711023.100	711023.1000
POLYGOPREP 300-30 C_4	yes	~ 1 % C	300 Å	25–40 µm	711690.100	711690.1000
POLYGOPREP 300-50 C_4	yes	~ 1 % C	300 Å	40–63 µm	711700.100	711700.1000
POLYGOPREP 1000-30 C_4	yes	< 1 % C	1000 Å	25–40 µm	711026.100	711026.1000
POLYGOPREP 1000-50 C_4	yes	< 1 % C	1000 Å	40–63 µm	711027.100	711027.1000
Cyano phases (nitrile) $-(CH_2)_3-CN$						
POLYGOPREP 60-12 CN		~ 4.5 % C	60 Å	10–15 µm	711015.100	711015.1000
POLYGOPREP 60-20 CN		~ 4.5 % C	60 Å	15–25 µm	711016.100	711016.1000
POLYGOPREP 60-30 CN		~ 4.5 % C	60 Å	25–40 µm	711017.100	711017.1000
Amino phases $-(CH_2)_3-NH_2$						
POLYGOPREP 60-12 NH_2		~ 3 % C	60 Å	10–15 µm	711012.100	711012.1000
POLYGOPREP 60-20 NH_2		~ 3 % C	60 Å	15–25 µm	711013.100	711013.1000
POLYGOPREP 60-30 NH_2		~ 3 % C	60 Å	25–40 µm	711014.100	711014.1000

Ordering information

Phase	Pore size	Particle size	Pack of 100 g	Pack of 1 kg	Pack of 5 kg
Unmodified POLYGOPREP silica SiOH					
POLYGOPREP 60-12	60 Å	10–15 µm		711001.1000	711001.5000
POLYGOPREP 60-20	60 Å	15–25 µm		711240.1000	711240.5000
POLYGOPREP 60-30	60 Å	25–40 µm		711250.1000	711250.5000
POLYGOPREP 60-50	60 Å	40–63 µm		711260.1000	711260.5000
POLYGOPREP 60-80	60 Å	63–100 µm		711270.1000	711270.5000
POLYGOPREP 60-130	60 Å	63–200 µm		711037.1000	711037.5000
POLYGOPREP 100-12	100 Å	10–15 µm		711002.1000	711002.5000
POLYGOPREP 100-20	100 Å	15–25 µm		711003.1000	711003.5000
POLYGOPREP 100-30	100 Å	25–40 µm		711540.1000	711540.5000
POLYGOPREP 100-50	100 Å	40–63 µm		711550.1000	711550.5000
POLYGOPREP 100-80	100 Å	63–100 µm		711033.1000	711033.5000
POLYGOPREP 100-130	100 Å	63–200 µm		711034.1000	711034.5000
POLYGOPREP 300-12	300 Å	10–15 µm	711004.100	711004.1000	
POLYGOPREP 300-20	300 Å	15–25 µm	711610.100	711610.1000	
POLYGOPREP 300-30	300 Å	25–40 µm	711620.100	711620.1000	
POLYGOPREP 300-50	300 Å	40–63 µm	711630.100	711630.1000	
POLYGOPREP 1000-12	1000 Å	10–15 µm	711035.100	711035.1000	
POLYGOPREP 1000-20	1000 Å	15–25 µm	711036.100	711036.1000	
POLYGOPREP 1000-30	1000 Å	25–40 µm	711005.100	711005.1000	
POLYGOPREP 1000-50	1000 Å	40–63 µm	711006.100	711006.1000	

Adsorbents for column chromatography

Silica adsorbents for low pressure column chromatography

- Silica 60; pore size ~ 60 Å; pore volume ~ 0.75 mL/g; spec. surface BET ~ 500 m^2/g highly porous, amorphous silicic acid in the form of hard, opalescent particles, prepared by precipitation of water glass with sulfuric acid
- For higher demands on the performance of column packings we recommend our high-purity irregular POLYGOPREP silicas (see before).
- Silica FIA for the fluorescence indicator adsorption procedure for the determination of hydrocarbon groups in the testing of liquid fuels in accordance with DIN 51791 and ASTM D 1319-58T
- The FIA method determines saturated hydrocarbons, olefins and aromatic hydrocarbons of a sample chromatographically by adsorption and desorption in a column filled with FIA silica, in the presence of a fluorescent dye mixture.

Ordering information

Description	Particle size	1 kg	5 kg	25 kg
Silica 60, 0.015–0.04 mm	–	815650.1	815650.5	815650.25
Silica 60, 0.025–0.04 mm	–	815300.1	815300.5	815300.25
Silica 60, 0.04–0.063 mm	230–400 mesh	815380.1	815380.5	815380.25
Silica 60 M, 0.04–0.063 mm	230–400 mesh	815381.1	815381.5	815381.25
Silica 60, 0.05–0.1 mm	130–270 mesh	815390.1	815390.5	815390.25
Silica 60, 0.05–0.2 mm	70–270 mesh	815320.1	815320.5	815320.25
Silica 60, 0.063–0.2 mm	70–230 mesh	815330.1	815330.5	815330.25
Silica 60, < 0.063 mm	+230 mesh	815400.1	815400.5	815400.25
Silica 60, < 0.08 mm	+190 mesh	815310.1	815310.5	815310.25
Silica 60, 0.1–0.2 mm	70–130 mesh	815340.1	815340.5	815340.25
Silica 60, 0.2–0.5 mm	35–70 mesh	815350.1	815350.5	815350.25
Silica 60, 0.5–1.0 mm	18–35 mesh	815360.1	815360.5	815360.25
Silica FIA fine	0.071–0.16 mm	815410.1		
Silica FIA coarse	0.071–0.63 mm	815430.1		

Aluminum oxide

- Aluminum oxides produced by dehydration of different aluminum hydroxides, e.g., hydrargillite between 400 and 500 °C.
- Activity grade I, particle size 50–200 µm, specific surface (BET) ~ 130 m^2/g

Ordering information

Description	pH	1 kg	5 kg	25 kg
Aluminum oxide 90 basic	pH 9.5 ± 0.3	815010.1	815010.5	815010.25
Aluminum oxide 90 neutral	pH 7 ± 0.5	815020.1	815020.5	815020.25
Aluminum oxide 90 acidic	pH 4 ± 0.3	815030.1	815030.5	815030.25

Kieselguhr

- Naturally occurring amorphous silicic acids of fossil origin, also known as diatomaceous earth or diatomite purified for chromatographic applications
- Compared to silica, kieselguhr has a small surface of low activity → application in partition chromatography; impregnated with various substances (paraffin, silicone oil, undecane) it can be used for reversed phase chromatography
- The following grades of kieselguhr are manufactured by Johns-Manville. They are narrowly classified with homogeneous particle size distributions and high purity.
- For columns packed with kieselguhr please see CHROMABOND® XTR for liquid-liquid extraction, page 63.

Ordering information

Description	Rel. purification factor	Rel. flow rate	1 kg	5 kg
Filter-Cel®	100	100	815510.1	815510.5
Hyflo® Super-Cel®	58	534	815530.1	815530.5
Celite® 503	42	910	815540.1	815540.5
Celite® 535	35	1269	815550.1	815550.5
Celite® 545	32	1830	815560.1	815560.5

Florisil®

- Hard granular magnesia silica gel:
  MgO 15.5 ± 0.5 % · SiO_2 84.0 ± 0.5 % · Na_2SO_4 ≤ 1.0 %;
  60/100 mesh
- Recommended application
  Sample preparation (see chapter “Solid phase extraction”, page 16)
- Clean-up of pesticide residues, separation of chlorinated pesticides, extraction of steroids, sex hormones, antibiotics, lipids etc.

Ordering information

Description	Particle size	1 kg	5 kg
Florisil standard 60/100 mesh	0.15/0.25 mm	815710.1	815710.5

Adsorbents for column chromatography

Polyamide

- Polyamide 6 = ε-polycaprolactam
- The separation mechanism mainly based on hydrogen bonds
- Recommended application
  Separation of phenolic compounds (e.g., isolation of natural products) carboxylic acids, aromatic nitro compounds
- For SPE columns packed with polyamide see CHROMABOND® PA page 44.

Ordering information

Description	Particle size	1 kg	5 kg
Polyamide SC 6, < 0.07 mm	< 0,07 mm	815610.1	815610.5
Polyamide SC 6, 0.05–0.16 mm	0.05–0.16 mm	815620.1	815620.5
Polyamide SC 6, 0.10–0.30 mm	0.10–0.30 mm	815600.1	815600.5

Unmodified cellulose

- Cellulose MN 100:
  native fibrous cellulose, standard grade average degree of polymerization 620–680, fiber length (85 %) 20–100 µm, specific surface acc. to Blaine ~ 6500 cm^2/g; residue on ignition at 850 °C < 10000 ppm, < 20 ppm Fe, < 5 ppm Cu, < 7 ppm P, CH_2Cl_2 extract < 0.20 %
- Cellulose MN 2100:
  native fibrous cellulose, purified grade (washed with different eluents) average degree of polymerization 620–680, fiber length (85 %) 20–75 µm, specific surface acc. to Blaine ~ 5500 cm^2/g residue on ignition at 850 °C < 1000 ppm, < 2 ppm Fe, < 1 ppm Cu, < 2 ppm P, CH_2Cl_2 extract < 0.15 %
- Grade MN 2100ff is a defatted cellulose MN 2100 with a CH_2Cl_2 extract < 0.02 %

Ordering information

Description	1 kg	5 kg	25 kg
Cellulose MN 100	815050.1	815050.5	815050.25
Cellulose MN 2100	815060.1	815060.5	815060.25
Cellulose MN 2100ff (Cellulose MN 2100 defatted)	815070.1		

MACHEREY-NAGEL
optimal autosampler vials for your sample

Vials and closures

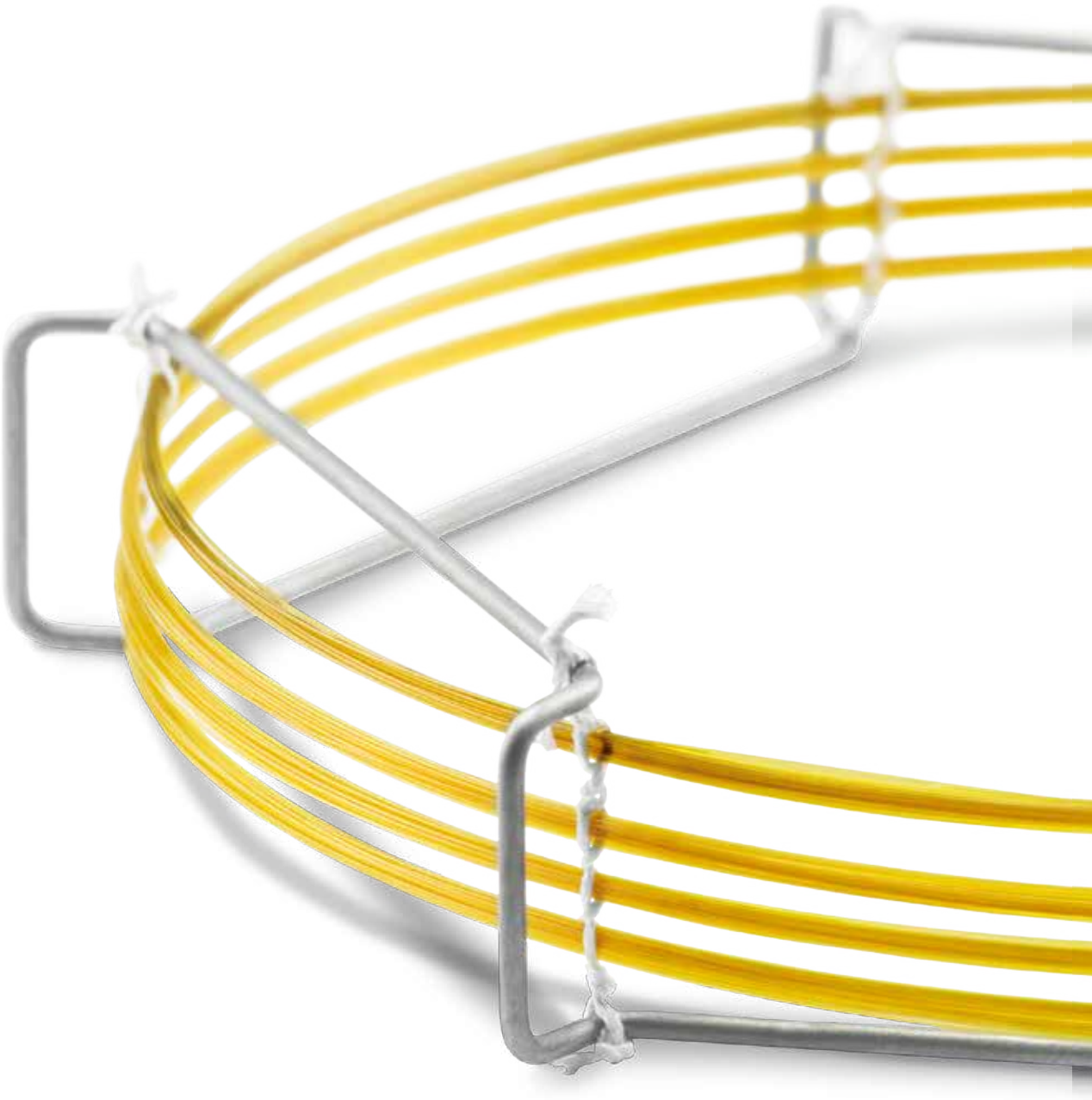
For reliable and reproducible analysis the correct storage of sample solutions is important. MACHEREY-NAGEL offers diverse vials and suitable closures.

Our product range includes

- Different vial types from N 8 to N 24
  - Crimp neck
  - Screw neck
  - Snap ring
- Clear glass, amber glass and polypropylene vials, with or without scale and label
- Diverse inserts for small sample volumes
- Variety of closures and septa of different material
- Suitable accessories like crimping tools and vial containers
- Compatibility with different autosamplers from page 136 onwards

Our broad range of vials and closures can be found from page 97 onwards.

Also use our VialFinder on *www.mn-net.com/VialFinder*



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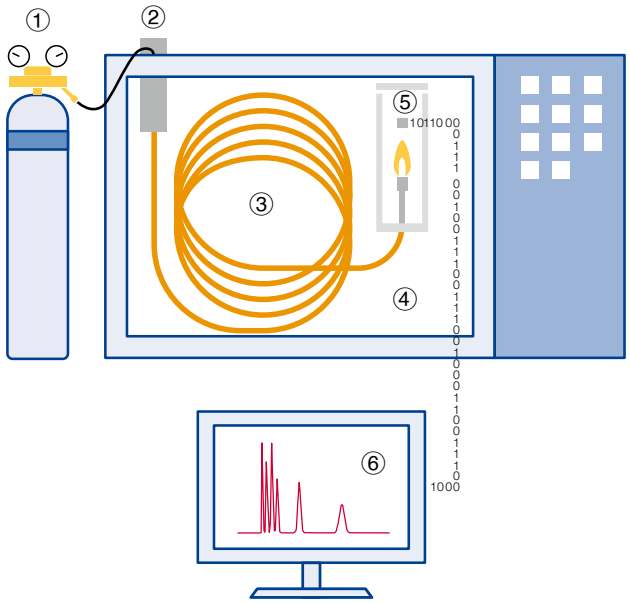
The GC system



Configuration of a gas chromatograph

① Gas supply: carrier gas and - if necessary - detector gases e.g., for FID detector

② Sample injector: During direct injection, the sample is applied to the column without touching any other parts made from glass or metal (on-column injection). During indirect injection, the sample is brought into an evaporator and is then transferred onto the column either completely, or partially (split technique). Both techniques allow working at low temperatures, high temperatures and the use of temperature programming.

③ Capillary column: the heart of the GC system

④ Temperature-controlled oven

⑤ Detector: indicates a substance by generating an electrical signal (response). Some detectors are specific for certain classes of substances or for certain elements (e.g., P, N).

⑥ Data station for configuration of a gas chromatograph

The separation process

Chromatographic separation is achieved through continuous distribution of each sample component between the mobile and the stationary phase:

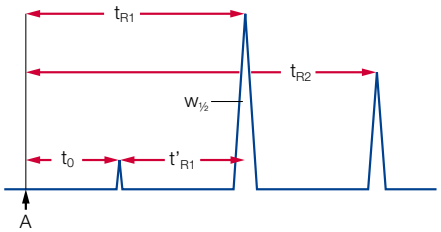
In GC, the mobile phase is always a gas, mostly either He, N_2 or H_2.

The stationary phase is often a viscous, gum-like liquid adhered to the inner wall of a capillary column (WCOT = Wall Coated Open Tubular).

Transport of the components occurs exclusively in the mobile phase, while separation only takes place in the stationary phase. The quality of a separation (resolution) depends on the residence time of the components within the stationary phase and on the rate of interactions. The type of interaction between component and phase (selectivity) is determined by the functional groups of the stationary phase. The polarity of the phase is a function of its substituents.

The chromatogram

A chromatogram consists of a base line and a number of peaks. The area of a peak allows quantitative determinations:



A: starting point of a chromatogram = time of injection of a dissolved solute

A component can be identified by its retention time (qualitative determination):

$$t_{Ri} = t_0 + t'_{Ri}$$

t_0: dead time = residence time of a solute in the mobile phase (time required by a component to migrate through the chromatographic system without any interaction with the stationary phase)

t_{Ri}: retention time = time interval between peak i and the point of injection

t'_{Ri}: net retention time = difference between total retention time and dead time t_0. It indicates how long a substance stays in the stationary phase.

Other terms characterizing a separation:

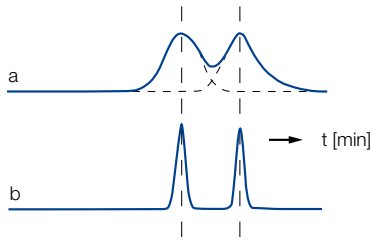
k': retention factor: a measure for the position of a sample peak in the chromatogram. The retention factor is specific for a given compound and constant under constant conditions.

$$k'_i = \frac{t_{Ri} - t_0}{t_0}$$

α: relative retention, also called separation factor or selectivity coefficient, is the ratio of two capacity factors. The reference substance is always in the denominator.

$$\alpha = \frac{k'_2}{k'_1}$$

The relative retention does not provide any information on the quality of a separation. For equal values of α two very broad peaks may overlap (as shown in a), or may be completely resolved (as in b), if they are accordingly narrow.



R: resolution: a measure for the quality of a separation, taking ($w_{1/2}$) into account according to:

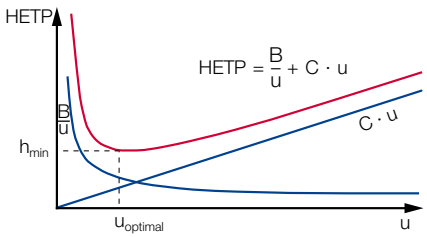
$$R = 1.18 \cdot \frac{t_{R2} - t_{R1}}{(w_{1/2})_2 + (w_{1/2})_1}$$

N: number of theoretical plates: characterizes the quality of a column (should be determined for k' > 5). The height equivalent to a theoretical plate (h, HETP) is calculated by dividing the length L of the column by the number of theoretical plates Nth. The smaller this value the more efficient the column.

$$N = 5.54 \cdot \frac{(t_{Ri})}{(w_{1/2})} \qquad h = HETP = \frac{L}{N}$$

The Golay equation shows how the plate height h depends on the flow velocity u:

B: molecular axial diffusion; B is a function of the diffusion coefficient of the component in the respective carrier gas



C: resistance to mass transfer

In practice often higher velocities than $u_{opt.}$ are chosen, if separation efficiency is sufficient. Higher carrier velocities mean shorter retention times.

Parameters characterizing a capillary column

OPTIMA® 5	1.0 µm film	30 m x	0.32 mm ID
A	B	C	D

A. Stationary phase

Different chemical structures of stationary phases are responsible for the type of interaction (selectivity) between the phase and the analytes. The stationary phase also limits the temperature range for chromatography. For a detailed summary of MN phases for GC please see the following chapter.

B. Film thickness

MACHEREY-NAGEL offers ranges from 0.1 to 5.0 µm. The standard film thickness is 0.25 µm. Thin films (0.1–0.2 µm) are very well suited for high-boiling, temperature-sensitive or almost contemporaneously eluting substances.

Increasing the film thickness will increase the capacity, the retention for low-boiling substances and the inertness of the column. This is especially helpful for samples with a broad range of concentrations, or the separation of volatile polar substances.

A better coverage of the column wall by a thicker film and a reduced column surface due to a shorter column have a positive impact on the separation of very active substrates, that may cause noticeable tailing when they come in contact with non-coated spots of the column wall.

Thick films, however, always mean more stationary phase in the column, hence increased column bleeding. Therefore, maximum operating temperatures for thick-film columns are reduced. In addition, thick-film columns may have a lesser separating capacity.

C. Column length

The separating efficiency (better the number of plates N) of a column is directly proportional to its length. Most routine separations are carried out on 25 or 30 m columns, while more complex samples may require 50 or 60 m. 10 m columns are common for Fast GC (see page 340).

D. Inner diameter (ID)

The lower the ID, the higher is the theoretically possible number of plates per meter.

0.1–0.2 mm ID:
for high resolution and short retention times at low carrier gas flow

0.25 mm ID:
for analysis of complex mixtures

0.32 mm ID:
for routine analysis with short retention times, but increased capacity

0.53 mm ID:
for rapid separations with inert surface and highest capacity

USP listing

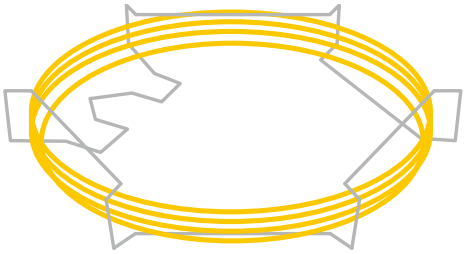
USP listing of MN GC phases

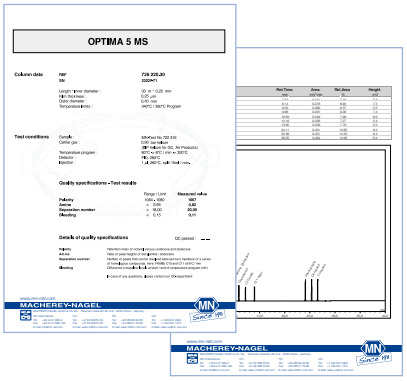
Code	Specifications	MN GC phases	Page
USP G1 / G2	dimethylpolysiloxane oil	OPTIMA® 1	310
		OPTIMA® 1 MS	312
		OPTIMA® 1 MS Accent	312
		OPTIMA® 1-TG	348
		PERMABOND® SE-30	336
		PERMABOND® P-100	352
USP G3	50 % phenyl - 50 % methylpolysiloxane	OPTIMA® 17	327
		OPTIMA® 17 MS	328
		OPTIMA® 17-TG	348
USP G6	trifluoropropylmethylpolysiloxane	OPTIMA® 210	329
USP G7	50 % 3-cyanopropyl - 50 % phenylmethylpolysiloxane	OPTIMA® 225	330
USP G16	polyethylene glycol (average molecular weight ~ 15 000); high molecular weight compound of polyethylene glycol and diepoxide	OPTIMA® WAX	332
		OPTIMA WAXplus®	333
		PERMABOND® CW 20 M	337
		PERMABOND® CW 20 M-DEG	354
		FS-CW 20 M-AM	351
USP G19	25 % phenyl – 25 % cyanopropyl – 50 % methylsiloxane	OPTIMA® 225	330
USP G25	high molecular weight compound of polyethylene glycol and diepoxide, which is esterified with terephthalic acid	OPTIMA® FFAP	334
		OPTIMA® FFAPplus	335
		PERMABOND® FFAP	338
USP G27	5 % phenyl – 95 % methylpolysiloxane	OPTIMA® 5	314
		OPTIMA® 5 Amine	350
		OPTIMA® 5 HT	349
		OPTIMA® 5 MS	315
		OPTIMA® 5 MS Accent	316
		PERMABOND® SE-52	336
USP G28	25 % phenyl – 75 % methylpolysiloxane	OPTIMA® 35 MS	326
USP G32	20 % phenylmethyl – 80 % dimethylpolysiloxane	OPTIMA® 35 MS	326
USP G35	high molecular weight compound of polyethylene glycol and diepoxide, which is esterified with nitroterephthalic acid	OPTIMA® FFAP	334
		OPTIMA® FFAPplus	335
		PERMABOND® FFAP	338
USP G36	1 % vinyl – 5 % phenylmethylpolysiloxane	OPTIMA® 5	314
		OPTIMA® 5 Amine	350
		OPTIMA® 5 HT	349
		OPTIMA® 5 MS	315
		OPTIMA® 5 MS Accent	316
		PERMABOND® SE-54 HKW	352
USP G38	dimethylpolysiloxane oil	OPTIMA® 1	310
		OPTIMA® 1 MS	312
		OPTIMA® 1 MS Accent	312
		OPTIMA® 1-TG	348
		PERMABOND® SE-30	336
		PERMABOND® P-100	352
USP G42	35 % phenyl – 65 % dimethylpolysiloxane	OPTIMA® 35 MS	326
USP G43	6 % cyanopropylphenyl – 94 % dimethylpolysiloxane	OPTIMA® 1301	321
		OPTIMA® 1301 MS	322
		OPTIMA® 624	323
		OPTIMA® 624 LB	323
USP G46	14 % cyanopropylphenyl – 86 % methylpolysiloxane	OPTIMA® 1701	324
		OPTIMA® 1701 MS	325
USP G49	proprietary derivatized phenyl groups on a polysiloxane backbone	OPTIMA® δ-3	319

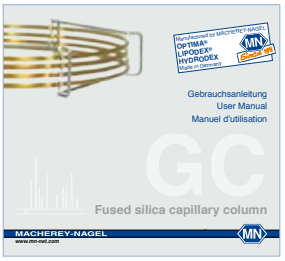
Additional information for GC columns

Scope of delivery

Each column is individually tested and supplied with test certificate and test chromatogram, but without fittings or ferrules. Columns have fused ends or are sealed with septa, to protect them from atmospheric oxygen. Further more an instruction leaflet is enclosed.

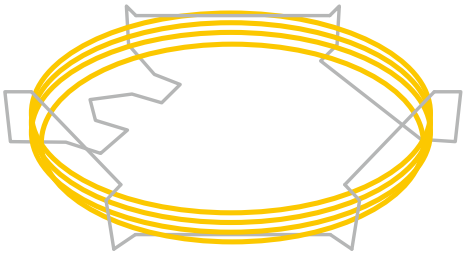




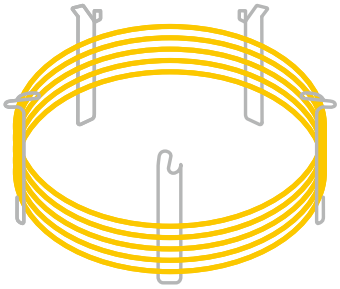


GC cages

The standard size of a GC cage is 7 inches. On request, all columns can be supplied on a 5 inch (13 cm) cage e.g., for the Agilent GC 6850. To order, please add an E at the end of the REF number (e.g., 726470.30E)



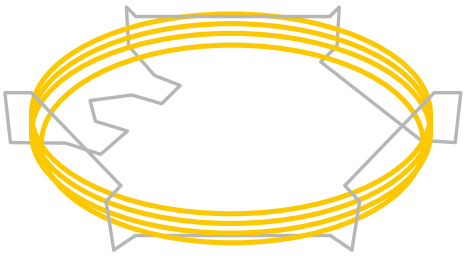
7 inches standard size e.g., REF 726600.30



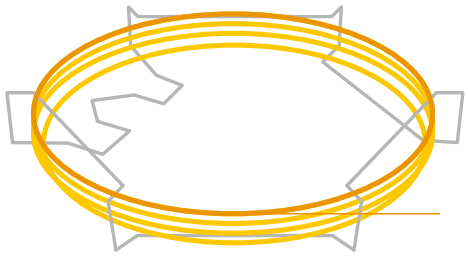
5 inches special cage e.g., REF 726600.30E

Integrated guard column

To prolong column life, even at highly contaminated or matrix-containing samples, MN offers the option to add an integrated guard column. All capillary columns are available with a 10 m guard column with respective deactivation. To order, please add V1 at the end of the REF number (e.g., 726600.30V1). Guard column combinations with other lengths, IDs or different deactivation are available on request.



Without integrated guard column e.g., REF 726600.30



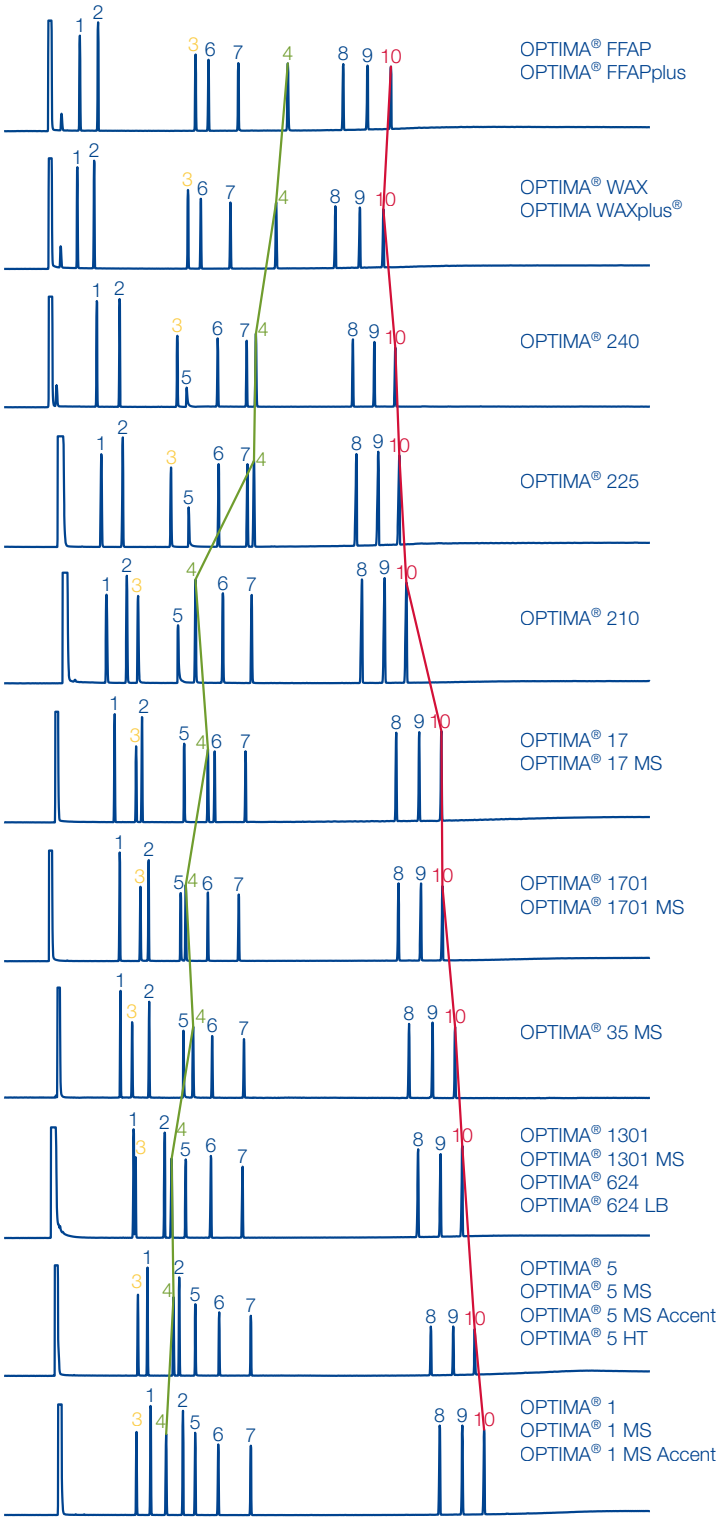
With integrated guard column e.g., REF 726600.30V1

MACHEREY-NAGEL derivatization reagents

Purpose of derivatization

- Improved volatility, better thermal stability or a lower limit of detection in gas chromatography
- Prerequisite: quantitative, rapid and reproducible formation of only one derivative
- Halogen atoms inserted by derivatization (e.g., trifluoroacetates) for specific detection (ECD) with the advantage of high sensitivity
- Influence of elution orders and fragmentation patterns in MS by a specific derivatization
- We provide reagents for
  - Silylation
  - Alkylation (methylation)
  - Acylaction
- For 1 x 10 mL, 1 x 50 mL and 6 x 50 mL also as screw neck vial
- Product range from page 357 onwards

Separation properties of OPTIMA® phases



Peaks:

1. Undecane
2. Dodecane
3. Octanol
4. Dimethylaniline
5. Decylamine
6. Methyl decanoate
7. Methyl undecanoate
8. Henicosane
9. Docosane
10. Tricosane

All columns:	0.25 µm film, 30 m x 0.25 mm ID
Sample:	MN OPTIMA® test mixture (REF 722316)
Injection:	1.0 µL, split 15 mL/min
Carrier gas:	0.80 bar He
Temperature:	80 °C T_{max} (isothermal), 8 °C/min (20 min T_{max})
Detector:	FID 260–280 °C

Summary of MN phases for GC

Overview of OPTIMA® MN phases

Phase	Composition	Page	Relative polarity ①	Maximum temperature ②
OPTIMA® 1	100 % dimethylpolysiloxane	310		
OPTIMA® 1 MS		312		340 / 360 °C
OPTIMA® 1 MS Accent		312		
OPTIMA® 5	5 % phenyl – 95 % methylpolysiloxane	314		340 / 360 °C
OPTIMA® 5 MS	5 % diphenyl – 95 % dimethylpolysiloxane	315		340 / 360 °C
OPTIMA® 5 MS Accent	silarylene phase with selectivity similar to 5 % diphenyl – 95 % dimethylpolysiloxane	316		340 / 360 °C
OPTIMA® XLB	silarylene phase like above, optimized silarylene content for low bleeding	317		340 / 360 °C
OPTIMA® δ-3	phase with autoselectivity ④	319		340 / 360 °C
OPTIMA® δ-6	phase with autoselectivity ④	320		340 / 360 °C
OPTIMA® 1301	6 % cyanopropylphenyl – 94 % dimethylpolysiloxane	321		300 / 320 °C
OPTIMA® 1301 MS	silarylene phase with low bleeding: polarity similar to 6 % cyanopropylphenyl – 94 % dimethylpolysiloxane	322		300 / 320 °C
OPTIMA® 624	6 % cyanopropylphenyl – 94 % dimethylpolysiloxane	323		280 / 300 °C
OPTIMA® 624 LB	like above, phase with low bleeding	323		
OPTIMA® 1701	14 % cyanopropylphenyl – 86 % dimethylpolysiloxane	324		280 / 300 °C
OPTIMA® 1701 MS	silarylene phase with low bleeding: polarity similar to 14 % cyanopropylphenyl – 86 % dimethylpolysiloxane	325		280 / 300 °C

① = nonpolar, = polar

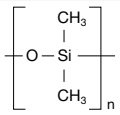
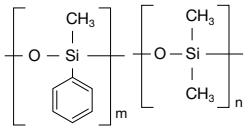
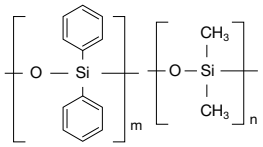
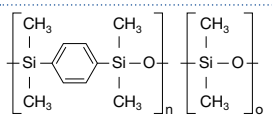
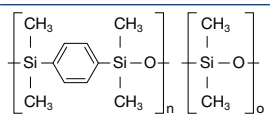
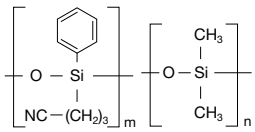
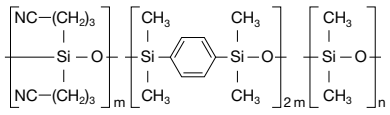
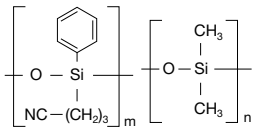
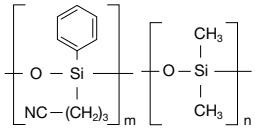
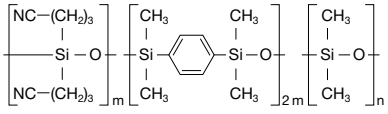
② First temperature (long term temperature) for isothermal operation, second value for the max. temperature (short term temperature) in a temperature program. Please note th
For details refer to the description of individual phases.

③ Phases which provide a similar selectivity based on chemical and physical properties

④ See description on page 318

GC columns for special separations can be found from page 339 onwards.

Summary of MN phases for GC

Structure	USP	Similar phases ③
	G1 / G2 / G38	PERMABOND® SE-30, OV-1, DB-1, SE-30, HP-1, SPB™-1, CP-Sil 5 CB, Rtx®-1, 007-1, BP1, MDN-1, AT™-1, ZB-1, OV-101
		5 % diphenyl – 95 % dimethylpolysiloxane
	G27 / G36	PERMABOND® SE-52, SE-54, SE-52, HP-5, SPB™-5, CP-Sil 8, Rtx®-5, 007-5, BP5, MDN-5, AT™-5, ZB-5
	G27 / G36	DB-5, DB-5MS, HP-5MS, Ultra-2, Equity™-5, CP-Sil 8CB low bleed/MS, Rxi®-5MS, Rtx®-5SIL-MS, Rtx®-5MS, 007-5MS, BPX™5, MDN-5S, AT™-5MS, VF-5MS
	G27 / G36	
	–	DB-XLB, Rxi®-XLB, Rtx®-XLB, MDN-12, VF-XMS
see description page 318	G49	no similar phases
see description page 318	–	no similar phases
	G43	HP-1301, DB-1301, SPB™-1301, Rtx®-1301, CP-1301, 007-1301
	G43	VF-1301ms, Rxi®-1301Sil MS, TG-1301MS
	G43	HP-624, HP-VOC, DB-624, DB-VRX, SPB™-624, CP-624, Rtx®-624, Rtx®-Volatiles, 007-624, BP624, VOCOL
	G46	OV-1701, DB-1701, CP-Sil 19 CB, HP-1701, Rtx®-1701, SPB™-1701, 007-1701, BP10, ZB-1701
	G46	VF-1701ms, TG-1701MS, OV-1701, DB-1701, HP-1701, Rtx®-1701, SPB™-1701, CP Sil 19 CB, 007-1701, BP10, ZB-1701

at for columns with 0.53 mm ID and for columns with thicker films temperature limits are generally lower.

Summary of MN phases for GC

Phase	Composition	Page	Relative polarity ①	Maximum temperature ②
OPTIMA® 35 MS	silarylene phase with selectivity similar to 35 % diphenyl – 65 % dimethylpolysiloxane	326		360 / 370 °C
OPTIMA® 17	phenylmethylpolysiloxane, 50 % phenyl	327		320 / 340 °C
OPTIMA® 17 MS	silarylene phase with selectivity similar to 50 % phenyl – 50 % methylpolysiloxane	328		340 / 360 °C
OPTIMA® 210	trifluoropropylmethylpolysiloxane (50 % trifluoropropyl)	329		260 / 280 °C
OPTIMA® 225	50 % cyanopropylmethyl – 50 % phenylmethylpolysiloxane	330		260 / 280 °C
OPTIMA® 240	33 % cyanopropylmethyl – 67 % dimethylpolysiloxan	331		260 / 280 °C
OPTIMA® WAX	polyethylene glycol 20 000 Da	332		240 / 250 °C
OPTIMA WAXplus®	polyethylene glycol with optimized cross-linking	333		260 / 270 °C
OPTIMA® FFAP	polyethylene glycol 2-nitroterephthalate	334		250 / 260 °C
OPTIMA® FFAPplus	polyethylene glycol 2-nitroterephthalate with optimized cross-linking	335		250 / 260 °C

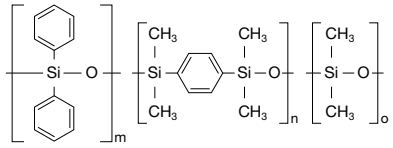
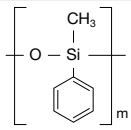
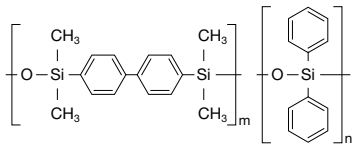
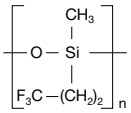
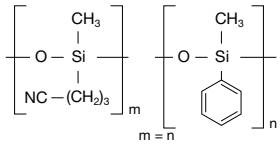
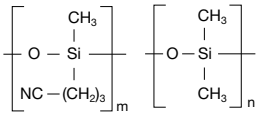
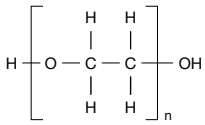
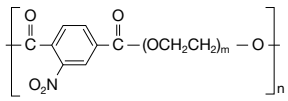
① = nonpolar, = polar

② First temperature (long term temperature) for isothermal operation, second value for the max. temperature (short term temperature) in a temperature program. Please note t
For details refer to the description of individual phases.

③ Phases which provide a similar selectivity based on chemical and physical properties

GC columns for special separations can be found from page 339 onwards.

Summary of MN phases for GC

Structure	USP	Similar phases ③
	G28 / G32 / G42	DB-35 MS, HP-35, SPB™-35, Rxi®-35SIL MS, Rtx-35, 007-35, BPX™-35, MDN-35, AT™-35 MS, ZB-35, OV-11, VF-35 MS
	G3	OV-17, DB-17, HP-50+, HP-17, SPB™-50, SP-2250, Rxi®-17, Rtx®-50, CP-Sil 24 CB, 007-17, ZB-50
	G3	OV-17, AT™-50, BPX™-50, DB-17, DB-17ms, HP-50+, HP-17, SPB™-50, SPB™-17, SP-2250, Rtx®-50, CP-Sil 24 CB, 007-17, VF-17ms, ZB-50
	G6	OV-210, DB-210, Rtx®-200, 007-210
	G7 / G19	DB-225, HP-225, OV-225, Rtx®-225, CP-Sil 43, 007-225, BP225
	–	no similar phases
	G16	PERMABOND® CW 20 M, DB-Wax, Supelcowax, HP-Wax, HP-INNOWAX, Rtx-Wax, CP-Wax 52 CB, Stabilwax, 007-CW, BP20, AT-Wax, ZB-Wax DB-Wax, Supelcowax, HP-Wax, HP-INNOWAX, Rtx-Wax, CP-Wax 52 CB, Stabilwax, 007-CW, BP20, AT-Wax, ZB-Wax
	G35 / G25	PERMABOND® FFAP, DB-FFAP, HP-FFAP, CP-Wax 58 FFAP CB, 007-FFAP, CP-FFAP CB, Nukol™, AT-1000, SPB-1000, BP21, OV-351 DB-FFAP, HP-FFAP, CP-SIL 58 CB, 007-FFAP, CP-FFAP CB, Nukol™

hat for columns with 0.53 mm ID and for columns with thicker films temperature limits are generally lower.

OPTIMA® 1 100 % dimethylpolysiloxane · USP G1 / G2 / G38

Key features
- Nonpolar phase
- Structure see page 307

Recommended application
- Separation of components according to boiling points
- Thick film columns ≥ 3 µm film are especially recommended for solvent analysis.

Temperature
- Columns with 0.1–0.32 mm ID and films < 3 µm: T_{max} 340 °C (long-term temperature), T_{max} 360 °C (short-term max. temperature in a temperature program)
- 0.53 mm ID, films < 3 µm: T_{max} 320 and 340 °C, resp.
- Thick film columns with films ≥ 3 µm: max. temperatures 300 and 320 °C, resp.

Similar phases
- PERMABOND® SE-30 (see page 336), OV-1, DB-1, SE-30, HP-1, SPB™-1, CP-Sil 5 CB, Rtx®-1, 007-1, BP1, MDN-1, AT™-1, ZB-1, OV-101

Ordering information

OPTIMA® 1

	Length → 10 m	12 m	15 m	20 m	25 m	30 m	50 m	60 m
0.1 mm ID (0.4 mm OD)								
0.10 µm film	726024.10			726024.20				
0.40 µm film				726025.20				
0.2 mm ID (0.4 mm OD)								
0.10 µm film					726832.25			
0.20 µm film		726834.12			726834.25		726834.50	
0.35 µm film		726837.12			726837.25		726837.50	
0.50 µm film							726839.50	
0.25 mm ID (0.4 mm OD)								
0.10 µm film	726038.10		726038.15		726038.25	726038.30		726038.60
0.25 µm film	726050.10		726050.15		726050.25	726050.30	726050.50	726050.60
0.50 µm film	726081.10				726081.25	726081.30	726081.50	726081.60
1.00 µm film					726802.25	726802.30	726802.50	726802.60
0.32 mm ID (0.5 mm OD)								
0.10 µm film	726301.10				726301.25	726301.30	726301.50	726301.60
0.25 µm film	726302.10		726302.15		726302.25	726302.30	726302.50	726302.60
0.35 µm film					726821.25	726821.30	726821.50	726821.60
0.50 µm film	726304.10				726304.25	726304.30	726304.50	726304.60
1.00 µm film	726323.10		726323.15		726323.25	726323.30	726323.50	726323.60
3.00 µm film					726805.25	726805.30	726805.50	726805.60
5.00 µm film	726931.10				726931.25	726931.30	726931.50	
0.53 mm ID (0.8 mm OD)								
0.50 µm film			726519.15		726519.25	726519.30		
1.00 µm film	726529.10		726529.15		726529.25	726529.30		
2.00 µm film	726521.10				726521.25	726521.30	726521.50	
5.00 µm film	726926.10				726926.25	726926.30	726926.50	

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

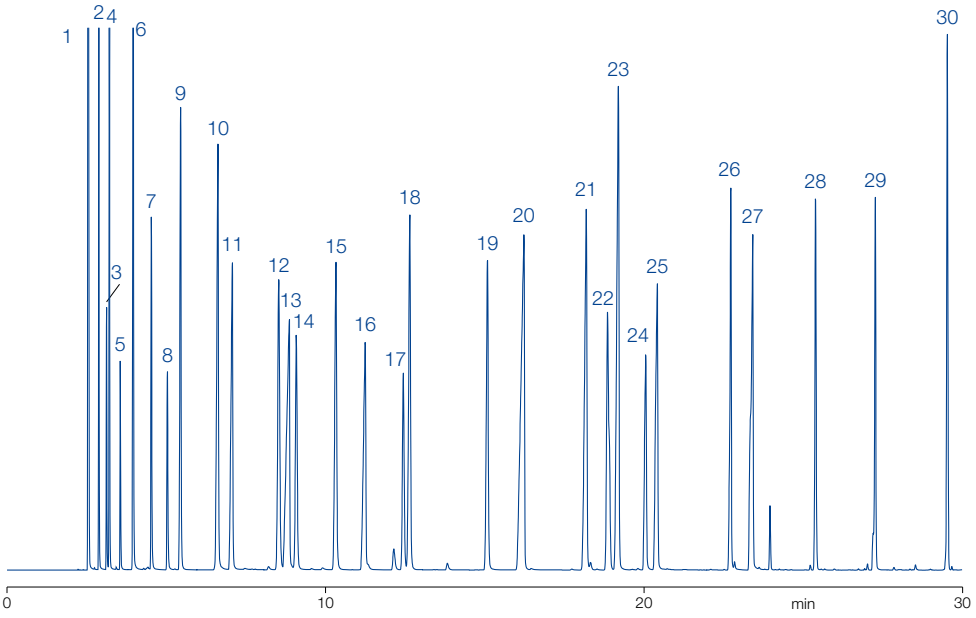
Solvent analysis

MN Appl. No. 201390

Column:	OPTIMA® 1, 60 m x 0.32 mm ID, 1.0 µm film
Sample:	solvent mixture, courtesy of J. Lutz, Alcan Rorschach, Switzerland
Injection:	0.4 µL, split 1:60
Carrier gas:	H_2, 120 kPa
Temperature:	50 °C (9 min) → 90 °C, 4 °C/min → 280 °C (2 min), 14 °C/min
Detector:	FID 300 °C

Peaks:

1. Methanol
2. Ethanol
3. Acetone
4. 2-Propanol
5. Methyl acetate
6. *n*-Propanol
7. Methyl ethyl ketone
8. Ethyl acetate
9. Isobutanol
10. *n*-Butanol
11. 1-Methoxy-2-propanol
12. Isooctane
13. Ethyl glycol
14. Isoheptane
15. Methyl isobutyl ketone
16. 1-Ethoxy-2-propanol
17. Toluene
18. Isobutyl acetate
19. Butyl acetate
20. 4-Hydroxy-4-methyl-2-pentanone
21. 1-Methoxy-2-propyl acetate
22. Xylene
23. Cyclohexanone
24. Ethyl glycol acetate
25. Butyl glycol
26. Heptanol
27. Ethyl diglycol
28. Butyl diglycol
29. Butyl glycol acetate
30. Butyl diglycol acetate



OPTIMA® · nonpolar capillary columns

OPTIMA® 1 MS 100 % dimethylpolysiloxane · USP G1 / G2 / G38

Key features
- Selectivity identical to OPTIMA® 1, Phase with low bleeding
- Structure see page 307

Recommended application
- GC/MS and ECD, general analysis at trace level

Temperature
- T_{max} 340 °C (long-term temperature), T_{max} 360 °C (short-term max. temperature in a temperature program)

Similar phases
- Ultra-1, DB-1MS, HP-1MS, Rxi®-1MS, Rtx®-1MS, Equity™-1, AT™-1MS, VF-1MS, CP-Sil 5 CB MS

Ordering information

OPTIMA® 1 MS

Length →	12 m	15 m	25 m	30 m	50 m	60 m
0.2 mm ID (0.4 mm OD)						
0.20 µm film			726201.25		726201.50	
0.35 µm film	726203.12					
0.25 mm ID (0.4 mm OD)						
0.25 µm film		726205.15		726205.30		726205.60
0.32 mm ID (0.5 mm OD)						
0.25 µm film				726202.30		726202.60

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

OPTIMA® 1 MS Accent 100 % dimethylpolysiloxane · USP G1 / G2 / G38

Key features
- Selectivity identical to OPTIMA® 1, nonpolar phase
- Lowest column bleed
- Solvent rinsing for removal of impurities applicable
- Increased sensitivity due to an unmatched low background level
- Structure see page 307

Recommended application
- Ideal for ion trap and quadrupole MS detectors
- Perfect inertness for basic compounds
- All-round phase for environmental analysis, trace analysis, EPA methods, pesticides, PCB, food and drug analysis

Temperature
- T_{max} 340 °C (long-term temperature), T_{max} 360 °C (short-term max. temperature in a temperature program)

Similar phases
- Ultra-1, DB-1MS, HP-1MS, Rxi®-1MS, Rtx®-1MS, Equity™-1, AT™-1MS, VF-1MS, CP-Sil 5 CB MS

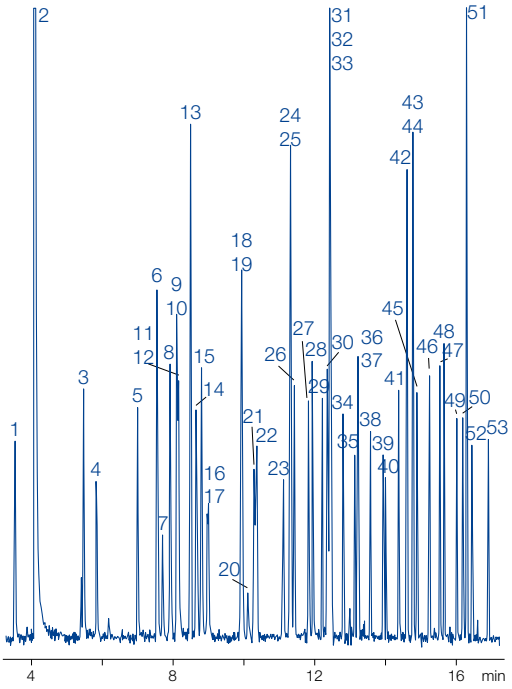
EPA 8140 / 8141 / 8141 A Organophosphorus pesticides

MN Appl. No. 213030

Column:	OPTIMA® 1 MS Accent, 30 m x 0.32 mm ID, 0.50 µm film
Sample:	0.2 µg/mL in hexane, 8140 / 8141 OP pesticides calibration mix A and 8141 OP pesticides calibration mix B; IS triphenyl phosphate and tributyl phosphate
Injection:	250 °C, splitless (hold 1 min)
Carrier gas:	He, 1 mL/min, constant pressure
Temperature:	100 °C → 180 °C, 10 °C/min (2 min) → 300 °C, 18 °C/min (3 min)
Detector:	FPD (Flame Photometric Detector), 280 °C

Peaks:

1. Dichlorvos
2. Hexamethylphosphoramide
3. Mevinphos
4. Trichlorfon
5. TEPP
6. Thionazin
7. Demeton-O
8. Ethoprop
9. Tributyl phosphate (IS)
10. Dicrotophos
11. Monocrotophos
12. Naled
13. Sulfotepp
14. Phorate
15. Dimethoate
16. Demeton-S
17. Dioxathion
18. Terbufos
19. Fonophos
20. Phosphamidon isomer
21. Diazinon
22. Disulfoton
23. Phosphamidon
24. Dichlorofenthion
25. Parathion-methyl
26. Chlorpyrifos-methyl
27. Ronnel
28. Fenitrothion
29. Malathion
30. Fenthion
31. Aspon
32. Parathion-ethyl
33. Chlorpyrifos
34. Trichloronate
35. Chlorfenvinphos
36. Merphos
37. Crotoxyphos
38. Stirofos
39. Tokuthion
40. Merphos oxidation product
41. Fensulfothion
42. Famphur
43. Ethion
44. Bolstar
45. Carbophenothion
46. Triphenyl phosphate (IS)
47. Phosmet
48. EPN
49. Azinphos-methyl
50. Leptophos
51. Tri-o-cresyl phosphate
52. Azinphos-ethyl
53. Coumaphos



Ordering information

OPTIMA® 1 MS Accent

	Length → 15 m	25 m	30 m	50 m	60 m
0.2 mm ID (0.4 mm OD)					
0.20 µm film		725801.25		725801.50	
0.25 mm ID (0.4 mm OD)					
0.25 µm film	725805.15		725805.30		725805.60
0.50 µm film			725806.30		725806.60
0.32 mm ID (0.5 mm OD)					
0.25 µm film			725802.30		725802.60
0.50 µm film			725807.30		725807.60

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

OPTIMA® · weakly polar capillary columns

OPTIMA® 5 5 % phenyl – 95 % methylpolysiloxane · USP G27 / G36

Key features
- Nonpolar phase
- Structure see page 307

Recommended application
- Standard phase with large range of application

Temperature
- Columns with 0.1–0.32 mm ID and films < 3 µm: T_{max} 340 °C (long-term temperature), T_{max} 360 °C (short-term max. temperature in a temperature program)
- 0.53 mm ID, films < 3 µm: T_{max} 320 and 340 °C, resp.
- Thick film columns with films ≥ 3 µm: max. temperatures 300 and 320 °C, resp.

Similar phases
- PERMABOND® SE-52 (see page 336), SE-54, SE-52, HP-5, SPB™-5, CP-Sil 8, Rtx®-5, 007-5, BP5, MDN-5, AT™-5, ZB-5

Ordering information

OPTIMA® 5

	Length → 10 m	15 m	25 m	30 m	50 m	60 m
0.1 mm ID (0.4 mm OD)						
0.10 µm film	726846.10					
0.2 mm ID (0.4 mm OD)						
0.10 µm film			726854.25			
0.20 µm film			726857.25		726857.50	
0.35 µm film			726860.25		726860.50	
0.50 µm film			726863.25		726863.50	
0.25 mm ID (0.4 mm OD)						
0.10 µm film			726911.25	726911.30	726911.50	726911.60
0.25 µm film	726056.10	726056.15	726056.25	726056.30	726056.50	726056.60
0.35 µm film			726623.25	726623.30	726623.50	726623.60
0.50 µm film			726099.25	726099.30	726099.50	726099.60
1.00 µm film			726807.25	726807.30	726807.50	726807.60
0.32 mm ID (0.5 mm OD)						
0.10 µm film	726313.10	726313.15	726313.25	726313.30	726313.50	726313.60
0.25 µm film		726314.15	726314.25	726314.30	726314.50	726314.60
0.35 µm film			726628.25	726628.30	726628.50	726628.60
0.50 µm film			726316.25	726316.30	726316.50	726316.60
1.00 µm film		726325.15	726325.25	726325.30	726325.50	726325.60
3.00 µm film			726809.25	726809.30	726809.50	726809.60
5.00 µm film		726934.15	726934.25	726934.30	726934.50	
0.53 mm ID (0.8 mm OD)						
0.50 µm film	726523.10		726523.25	726523.30		
1.00 µm film	726541.10	726541.15	726541.25	726541.30		
2.00 µm film	726525.10		726525.25	726525.30	726525.50	726525.60
5.00 µm film	726916.10		726916.25	726916.30	726916.50	

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

Further applications can be found online in our application database at *www.mn-net.com/apps*

OPTIMA® 5 MS 5 % diphenyl – 95 % dimethylpolysiloxane · USP G27 / G36

Key features
- Selectivity identical to OPTIMA® 5
- Phase with low bleeding
- Structure see page 307

Recommended application
- GC/MS and ECD, applications and general analysis at trace level
- Perfect inertness for basic compounds

Temperature
- T_{max} 340 °C (long-term temperature), T_{max} 360 °C (short-term max. temperature in a temperature program)

Similar phases
- DB-5, DB-5MS, HP-5MS, Ultra-2, Equity™-5, CP-Sil 8CB low bleed/MS, Rxi®-5MS, Rtx®-5SIL-MS, Rtx®-5MS, 007-5MS, BPX™5, MDN-5S, AT™-5MS, VF-5MS

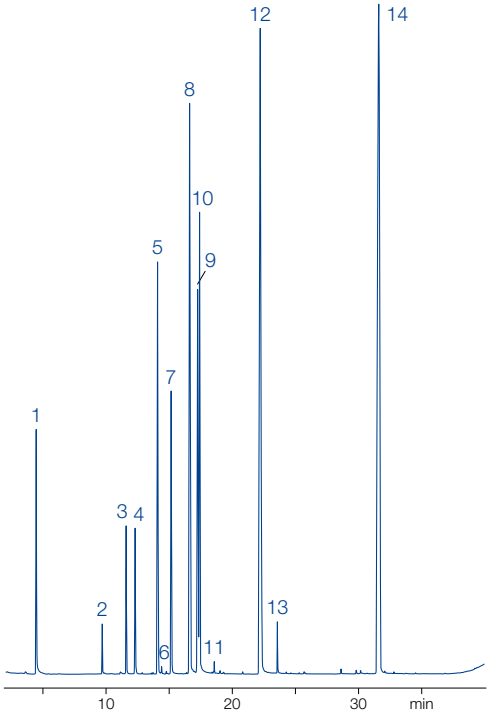
Analysis of various phenols

MN Appl. No. 210110

Column: OPTIMA® 5 MS, 30 m x 0.25 mm ID, 0.25 µm film
Sample: 5 ppm of each compound except *N-i*-propylaniline (9.4 ppm)
Methode: SPME
Temperature: 40 °C (2 min) → 240 °C, 6 °C/min → 320 °C, 20 °C/min
Detector: MSD

Peaks:
1. Toluene-D_8
2. Phenol
3. 2-Methylphenol (o-Cresol)
4. Nitrobenzene-D_5
5. *N-i*-Propylaniline
6. 2,4-Dichlorophenol
7. 4-Chlorophenol
8. 4-Bromo-2-chlorophenol
9. 3-Bromophenol
10. 4-Chloro-3-methylphenol
11. 2,4-Dibromophenol
12. 2-Hydroxybiphenyl
13. 2-Cyclohexylphenol
14. Hexafluorobisphenol A

Courtesy of Riedel-de-Haën, Seelze, Germany



Ordering information

OPTIMA® 5 MS

	Length → 12 m	15 m	25 m	30 m	50 m	60 m
0.2 mm ID (0.4 mm OD)						
0.20 µm film	726210.12		726210.25		726210.50	
0.35 µm film	726215.12		726215.25		726215.50	
0.25 mm ID (0.4 mm OD)						
0.25 µm film		726220.15		726220.30		726220.60
0.50 µm film				726225.30		726225.60
1.00 µm film				726226.30		726226.60
0.32 mm ID (0.5 mm OD)						
0.25 µm film				726211.30		
0.50 µm film				726213.30		
1.00 µm film			726212.25		726212.50	726212.60

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

OPTIMA® · weakly polar capillary columns

OPTIMA® 5 MS Accent silarylene phase · USP G27 / G36

Key features
- Chemically bonded, cross-linked silarylene phase with polarity similar to a 5 % diphenyl - 95 % dimethylpolysiloxane phase
- Lowest column bleed, nonpolar phase, solvent rinsing for removal of impurities applicable
- Structure see page 307

Recommended application
- Ideal for ion trap and quadrupole MS detectors
- Perfect inertness for basic compounds
- All-round phase for environmental analysis, trace analysis, EPA methods, pesticides, PCB, food and drug analysis

Temperature
- T_{max} 340 °C (long-term temperature), T_{max} 360 °C (short-term max. temperature in a temperature program)
- Film thickness > 0.5 µm: T_{max} 320 and 340 °C, resp.

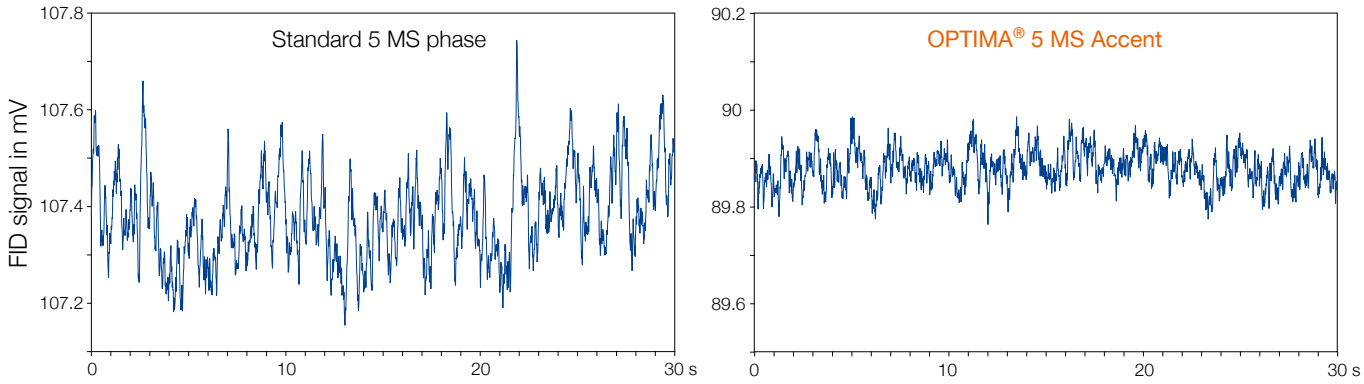
Similar phases
- DB-5, DB-5MS, HP-5MS, Ultra-2, Equity™-5, CP-Sil 8CB low bleed/MS, Rxi®-5MS, Rtx®-5SIL-MS, Rtx®-5MS, 007-5MS, BPX™5, MDN-5S, AT™-5MS, VF-5MS

Increased sensitivity due to an unmatched low background level

The bleed comparison test of OPTIMA® 5 MS Accent with a conventional 5 MS phase shows the outstanding performance of the silarylene phase.

The unmatched low background level of the OPTIMA® 5 MS Accent, which is approximately three times lower compared to a 5 MS brand column, provides significantly increased sensitivity and allows its application in trace analysis particularly of high-boiling compounds.

Background noise at 340 °C



Ordering information

OPTIMA® 5 MS Accent

	Length → 12 m	15 m	25 m	30 m	50 m	60 m
0.2 mm ID (0.4 mm OD)						
0.20 µm film			725810.25		725810.50	
0.35 µm film	725815.12				725815.50	
0.25 mm ID (0.4 mm OD)						
0.25 µm film		725820.15		725820.30		725820.60
0.50 µm film				725825.30		725825.60
1.00 µm film				725826.30		725826.60
0.32 mm ID (0.5 mm OD)						
0.25 µm film				725811.30		725811.60
0.50 µm film				725813.30		
1.00 µm film			725812.25			725812.60

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

OPTIMA® XLB silarylene phase

Key features

- Chemically bonded, cross-linked silarylene phase, optimized silarylene content for lowest column bleed, nonpolar phase, perfect inertness for basic compounds, solvent rinsing for removal of impurities applicable
- Structure see page 307

Recommended application

- Ideal for ion trap and quadrupole MS detectors, ultra low bleed phase, highly selective for environmental and trace analysis, pesticides, recommended phase for PCB separations

Temperature

- T_{max} 340 °C (long-term temperature), T_{max} 360 °C (short-term max. temperature in a temperature program)

Similar phases

- DB-XLB, Rxi®-XLB, Rtx®-XLB, MDN-12, VF-XMS

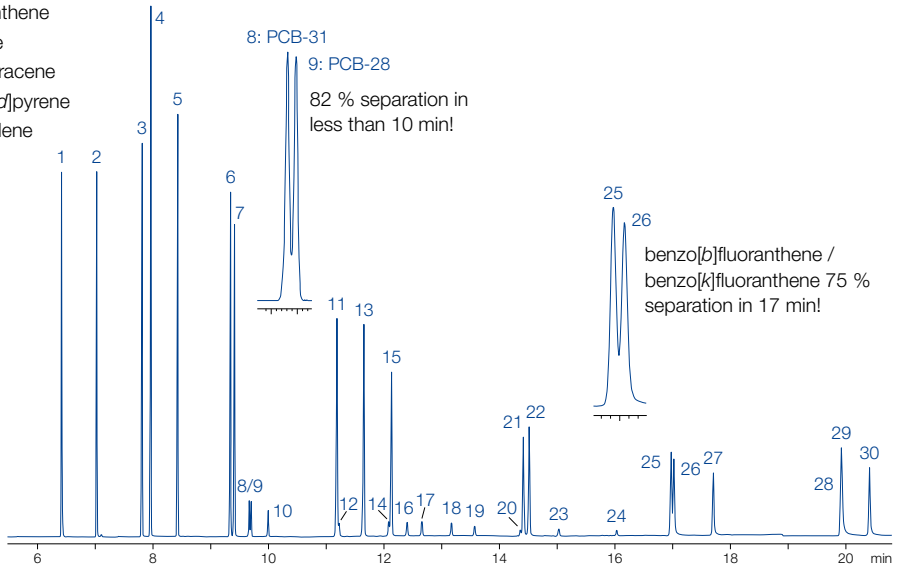
Rapid separation of PCB and PAH

MN Appl. No. 212920

Column:	OPTIMA® XLB, 30 m x 0.25 mm ID, 0.25 µm film
Injection:	1 µL, Standard 0.005 ng/µL, 250 °C, pulsed, splitless, pulse 1.38 bar in 1 min
Carrier gas:	60 mL/min He
Temperature:	40 °C (2 min) → 240 °C (2 min), 30 °C/min → 340 °C (5 min), 10 °C/min
Detektion:	MS source 230 °C, interface 280 °C, quadrupole 150 °C

Peaks:

1. Naphthalene
2. 2-Methylnaphthalene
3. Acenaphthylene
4. Acenaphthene
5. Fluorene
6. Phenanthrene
7. Anthracene
8. PCB-31
9. PCB-28
10. PCB-52
11. Fluoranthene
12. PCB-101
13. Pyrene
14. PCB-77
15. 2-Methylfluoranthene
16. PCB-118
17. PCB-153
18. PCB-138
19. PCB-126
20. PCB-180
21. Benz[*a*]anthracene
22. Chrysene
23. PCB-169
24. PCB-194
25. Benzo[*b*]fluoranthene
26. Benzo[*k*]fluoranthene
27. Benzo[*a*]pyrene
28. Dibenz[*ah*]anthracene
29. Indeno[1,2,3-*cd*]pyrene
30. Benzo[*ghi*]perylene



Courtesy of Centre d'Analyses de Recherche, Lab. d'Hydrologie, 65400 Illkirch, France

Ordering information

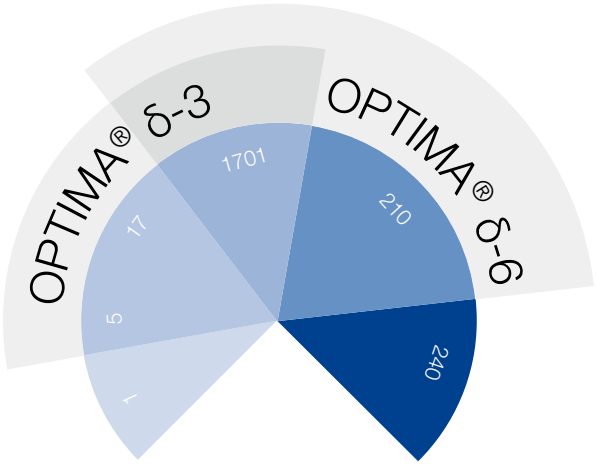
OPTIMA® XLB

	Length → 30 m	60 m
0.25 mm ID (0.4 mm OD)		
0.25 µm film	725850.30	725850.60

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

OPTIMA® δ · phases with autoselectivity

Range of polarities covered by OPTIMA® δ phases



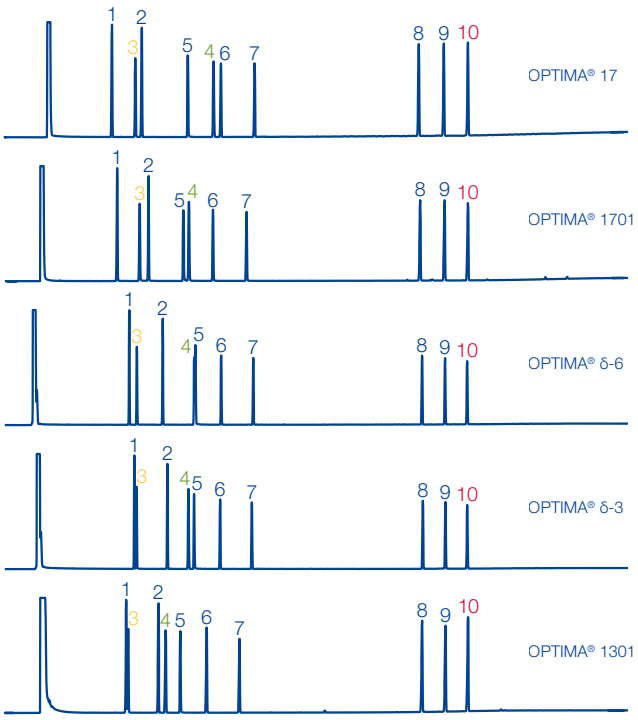
All stationary GC phases can be classified by their polarities. While the selectivity of common GC phases is generally determined by permanent dipole-dipole interactions, OPTIMA® δ-3 and OPTIMA® δ-6 show an additional feature. Large, polarizable groups in the polymer chain of the stationary phase enable the analyte to induce a further dipole moment that increases with the polarity of said analyte. We call this phenomenon "Autoselectivity", because the column adjusts itself to the polarity of the analyte. The implemented polymers consist of cross-linked polysiloxanes with a defined composition and an extremely narrow distribution of molecular weight.

OPTIMA® δ phases cover broad ranges of polarities. Compared with conventional phases, OPTIMA® δ-3 polarity ranges from approximately the nonpolar OPTIMA® 5 to the midpolar OPTIMA® 1701, while for OPTIMA® δ-6 the polarity covers a range from about the midpolar OPTIMA® 17 to the polar OPTIMA® 210.

OPTIMA® δ phases show high temperature limits (340 / 360 °C), as well as low bleed levels, which makes them ideal for the use with mass selective (MSD) or phosphorus/nitrogen detectors (PND) in the field of environmental trace analysis.

Isomeric phenols, such as chloro- and nitrophenols, are difficult to analyze with standard GC phases (e.g., OPTIMA® 5 or OPTIMA® 17) because of co-elutions. The autoselective OPTIMA® δ-3 is able to separate all 22 phenols due to stronger interactions occurring with more polar molecules, because polar analytes induce a dipole moment in the phase of the OPTIMA® δ-3 (see chromatogram page 319).

Separation characteristics of OPTIMA® δ phases



Conditions and peaks (see page 305)

Key features of OPTIMA® δ phases

- Wide range of application due to autoselectivity
- Outstanding thermal stability similar to nonpolar phases
- Low bleed levels
- Medium polar without CN groups

Ordering information about OPTIMA® δ phases can be found on page 319 and page 320.

OPTIMA® δ-3 polysiloxane phase with autoselectivity · USP G49

Key features

- Medium polar without CN groups
- Autoselectivity resulting in a polarity range from approximately the nonpolar OPTIMA® 5 to the midpolar OPTIMA® 1701 (see page 318)
- Analytes determine the polarity of the phase

Recommended application

- Ideal for MSD and PND detectors

Temperature

- 0.1–0.32 mm ID:
  T_{max} 340 °C (long-term temperature),
  T_{max} 360 °C (short-term max. temperature in a temperature program)
- 0.53 mm ID:
  T_{max} 320 and 340 °C, resp.

Similar phases

- Exclusive from MN

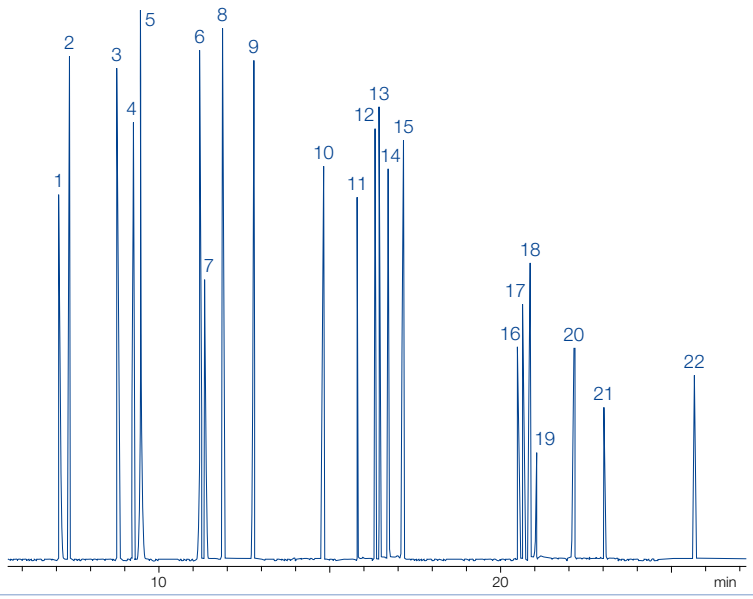
Analysis of isomeric phenols

MN Appl. No. 250060

Column: OPTIMA® δ-3, 60 m x 0.25 mm ID, 0.25 µm film
Injection: 1.0 µL, split 1:80
Carrier gas: He, 1.3 bar
Temperature: 60 °C (3 min) → 320 °C, 6 °C/min
Detector: MSD HP 5971

Peaks:

1. Phenol
2. 2-Chlorophenol
3. 2-Methylphenol
4. 4-Methylphenol
5. 3-Methylphenol
6. 2,4-Dimethylphenol
7. 2-Nitrophenol
8. 2,4-Dichlorophenol
9. 2,6-Dichlorophenol
10. 4-Chloro-3-methylphenol
11. 2,3,5-Trichlorophenol
12. 2,4,6-Trichlorophenol
13. 2,4,5-Trichlorophenol
14. 2,3,4-Trichlorophenol
15. 2,3,6-Trichlorophenol
16. 2,3,5,6-Tetrachlorophenol
17. 2,3,4,5-Tetrachlorophenol
18. 2,3,4,6-Tetrachlorophenol
19. 2,4-Dinitrophenol
20. 3,4,5-Trichlorophenol
21. 2-Methyl-4,6-dinitrophenol
22. 2-Isopropyl-4,6-dinitrophenol



Ordering information

OPTIMA® δ-3

Length → / 10 m	20 m	25 m	30 m	50 m	60 m
0.1 mm ID (0.4 mm OD)					
0.10 µm film 726410.10	726410.20				
0.2 mm ID (0.4 mm OD)					
0.20 µm film		726400.25		726400.50	
0.25 mm ID (0.4 mm OD)					
0.25 µm film			726420.30		726420.60
0.50 µm film			726421.30		
0.32 mm ID (0.5 mm OD)					
0.25 µm film			726440.30		726440.60
0.35 µm film			726441.30		726441.60
1.00 µm film			726442.30		726442.60
0.53 mm ID (0.8 mm OD)					
1.00 µm film			726443.30		

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

OPTIMA® δ · phases with autoselectivity

OPTIMA® δ-6 polysiloxane phase with autoselectivity

Key features

- Medium polar without CN groups Autoselectivity resulting in a polarity range from approximately the midpolar OPTIMA® 17 to the polar OPTIMA® 210 (see page 318)
- Analytes determine the polarity of the phase

Recommended application

- Ideal for MSD and PND detectors

Temperature

- 0.1–0.32 mm ID: T_{max} 340 °C (long-term temperature), T_{max} 360 °C (short-term max. temperature in a temperature program)
- 0.53 mm ID: T_{max} 320 and 340 °C, resp.

Similar phases

- Exclusive from MN

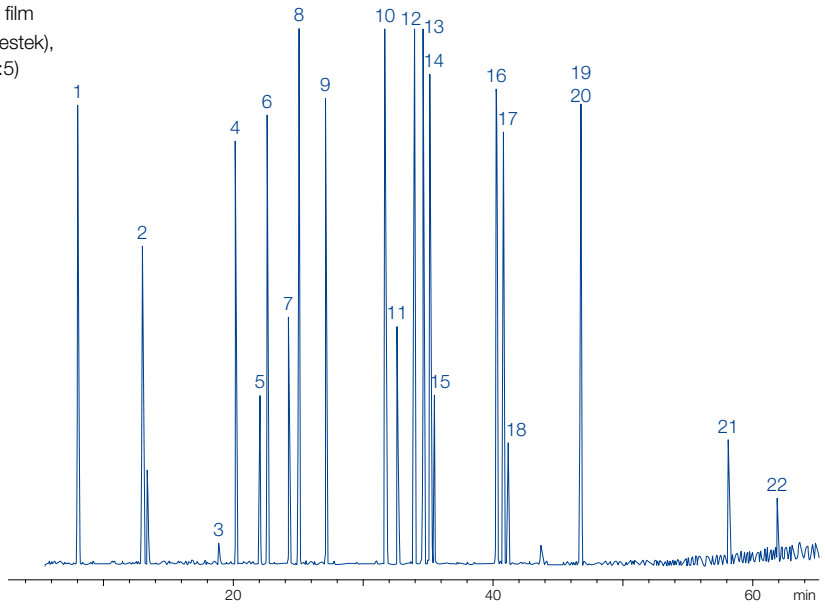
Separation of organophosphorus pesticides (EPA 8140 / 8141)

MN Appl. No. 250420

Column:	OPTIMA® δ-6, 50 m x 0.2 mm ID, 0.2 µm film
Sample:	EPA 8140 OP pesticide calibration mix (Restek), 200 µg/mL each in hexane – acetone (95:5)
Injection:	1 µL, split 1:30
Carrier gas:	2.0 bar He
Temperature:	150 °C → 300 °C (10 min), 2.5 °C/min
Detector:	MSD HP 5971

Peaks:

1. Dichlorvos
2. Mevinphos
3. Demeton-S
4. Ethoprop
5. Naled
6. Phorate
7. Demeton-O
8. Diazinon
9. Disulfoton
10. Ronnel
11. Parathion-methyl
12. Chlorpyrifos
13. Trichloronate
14. Fenthion
15. Merphos
16. Stirofos
17. Tokuthion
18. Merphos oxidation product
19. Fensulfothion
20. Bolstar
21. Azinphos-methyl
22. Coumaphos



Ordering information

OPTIMA® δ-6

Length → / 10 m	25 m	30 m	50 m	60 m
0.1 mm ID (0.4 mm OD)				
0.10 µm film	726490.10			
0.2 mm ID (0.4 mm OD)				
0.20 µm film		726465.25		726465.50	
0.25 mm ID (0.4 mm OD)				
0.25 µm film			726470.30		726470.60
0.32 mm ID (0.5 mm OD)				
0.25 µm film			726480.30		726480.60
0.35 µm film			726481.30		726481.60
1.00 µm film			726482.30		726482.60
0.53 mm ID (0.8 mm OD)				
1.00 µm film			726483.30		

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

OPTIMA® 1301 6 % cyanopropyl-phenyl – 94 % dimethylpolysiloxane · USP G43

Key features
- Midpolar phase
- Structure see page 307

Recommended application
- Pesticide analysis
- For corresponding columns with higher film thickness see OPTIMA® 624

Temperature
- T_{max} 300 °C (long-term temperature), T_{max} 320 °C (short-term max. temperature in a temperature program)

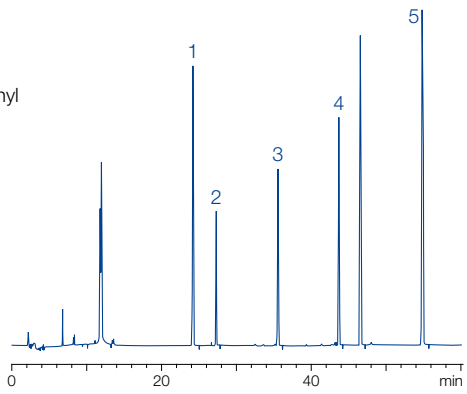
Similar phases
- HP-1301, DB-1301, SPB™-1301, Rtx®-1301, CP-1301, 007-1301

Analysis of a pesticide mixture

MN Appl. No. 210620

Column:	OPTIMA® 1301, 60 m x 0.25 mm ID, 0.25 µm film
Injection:	3 µL (0.1 ng/µL), 80 °C (1 min) → 250 °C (1 min) pulsed splitless
Carrier gas:	He, 54 mL/min
Temperature:	80 °C (2 min) → 190 °C, 20 °C/min (12 min) → 240 °C, 2 °C/min (23 min) → 260 °C, 10 °C/min (20 min)
Detector:	ECD

Peaks :
1. Propyzamide
2. Vinclozolin
3. Bromophos-ethyl
4. 2,4-DDT
5. Brompropylate

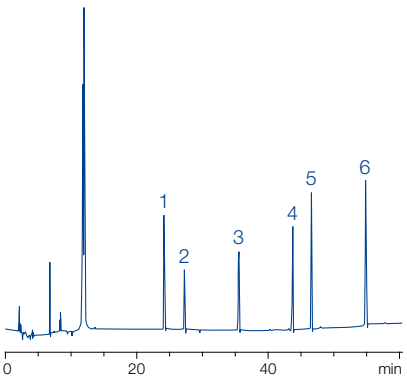


Analysis of a PCB mixture

MN Appl. No. 210650

Column:	OPTIMA® 1301, 60 m x 0.25 mm ID, 0.25 µm film
Injection:	3 µL (0.1 ng/µL), 80 °C (1 min) → 250 °C (1 min) pulsed splitless
Carrier gas:	He, 54 mL/min
Temperature:	80 °C (2 min) → 190 °C, 20 °C/min (12 min) → 240 °C, 2 °C/min (23 min) → 260 °C, 10 °C/min (20 min)
Detector:	ECD

Peaks :
1. PCB-28
2. PCB-52
3. PCB-128
4. PCB-153
5. PCB-138
6. PCB-180



Ordering information

OPTIMA® 1301

	Length → 25 m	30 m	50 m	60 m
0.25 mm ID (0.4 mm OD)				
0.25 µm film	726771.25	726771.30	726771.50	726771.60
0.32 mm ID (0.5 mm OD)				
0.25 µm film	726777.25	726777.30		726777.60
1.00 µm film		726780.30	726780.50	726780.60
0.53 mm ID (0.8 mm OD)				
1.00 µm film	726783.25			

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

Further applications can be found online in our application database at *www.mn-net.com/apps*

OPTIMA® 1301 MS 6 % cyanopropyl-phenyl – 94 % dimethylpolysiloxane · USP G43

Key features

- Chemically bonded, cross-linked silarylene phase with selectivity similar to 6 % cyanopropyl-phenyl – 94 % dimethylpolysiloxane, symmetric substituted cyanopropylsilanes and integrated phenyl rings (silarylene)
- Midpolar phase with very low bleed
- Perfect deactivation
- Structure see page 307

Recommended application

- Specially suitable for sophisticated environmental analysis (e.g., EPA methods for PAHs, PCBs and pesticides)
- 100 % ion trap and quadrupol MS compatibility

Temperature

- T_{max} 300 °C (long-term temperature), T_{max} 320 °C (short-term max. temperature in a temperature program)

Similar phases

- VF-1301ms, Rxi®-1301Sil MS, TG-1301MS

Ordering information

OPTIMA® 1301 MS

	Length → 30 m	60 m
0.25 mm ID (0.4 mm OD)		
0.25 µm film	726640.30	726640.60
0.32 mm ID (0.5 mm OD)		
0.25 µm film	726641.30	726641.60
1.00 µm film	726642.30	726642.60
0.53 mm ID (0.8 mm OD)		
1.00 µm film	726643.30	726643.60

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

OPTIMA® 624 6 % cyanopropyl-phenyl – 94 % dimethylpolysiloxane · USP G43

Key features
- Midpolar phase
- Structure see page 307

Recommended application
- Environmental analysis
- For corresponding columns with lower film thickness see OPTIMA® 1301

Temperature
- T_{max} 280 °C (long-term temperature), T_{max} 300 °C (short-term max. temperature in a temperature program)

Similar phases
- HP-624, HP-VOC, DB-624, DB-VRX, SPB™-624, CP-624, Rtx®-624, Rtx®-Volatiles, 007-624, BP624, VOCOL

OPTIMA® 624 LB 6 % cyanopropyl-phenyl – 94 % dimethylpolysiloxane

Key features
- Midpolar phase with low bleeding
- Structure see page 307

Recommended application
- Halogenated hydrocarbons, volatiles, aromatic compounds, solvents etc.

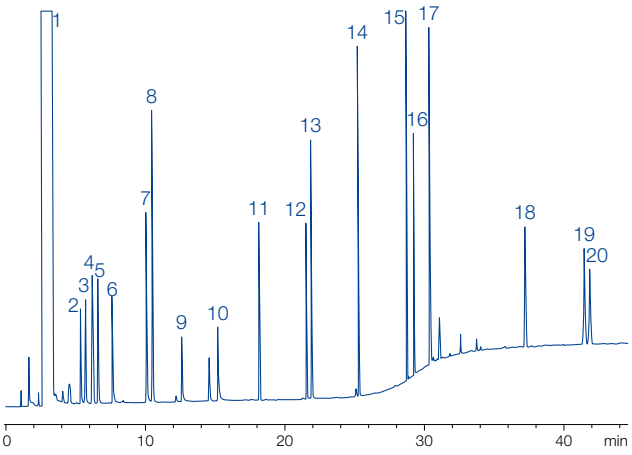
Solvents and semi-volatiles

MN Appl. No. 212520

Column:	OPTIMA® 624 LB, 30 m x 0.32 mm ID, 1.8 µm film; retention gap Phe-Sil 0.5 m x 0.53 mm
Injection:	1 µL (10 ppm per substance in acetone), cold on-column
Carrier gas:	1.1 bar He
Temperature:	45 °C (3 min) → 150 °C (6 °C/min) → 300 °C (18 °C/min), 20 min 300 °C
Detector:	FID 280 °C

Peaks:

1. Acetone
2. Ethyl acetate
3. Tetrahydrofuran
4. Cyclohexane
5. 2-Methyl-2-butanol
6. 1-Butanol
7. Pyridine
8. Toluene
9. Dimethylformamide
10. Dimethylsulfoxide
11. Decane
12. 1-Octanol
13. Acetophenone
14. Butyrophenone
15. Heptanophenone
16. 5-Methoxyindole
17. Dibenzylamine
18. Methyl eicosanoate
19. Methyl *cis*-13-docosenoate
20. Methyl docosanoate



Ordering information

	Length → 25 m	30 m	50 m	60 m
OPTIMA® 624				
0.2 mm ID (0.4 mm OD)				
1.10 µm film	726784.25			
0.25 mm ID (0.4 mm OD)				
1.40 µm film	726785.25	726785.30	726785.50	726785.60
0.32 mm ID (0.5 mm OD)				
1.80 µm film	726787.25	726787.30	726787.50	726787.60
0.53 mm ID (0.8 mm OD)				
3.00 µm film	726789.25	726789.30		
OPTIMA® 624 LB				
0.32 mm ID (0.5 mm OD)				
1.80 µm film		726786.30	726786.50	

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

OPTIMA® · medium polar capillary columns

OPTIMA® 1701 14 % cyanopropyl-phenyl – 86 % dimethylpolysiloxane · USP G46

Key features
- Midpolar phase, special selectivity due to high cyanopropyl content
- Structure see page 307

Recommended application
- Reference column for structure identification, e.g., in combination with OPTIMA® 5
- Film thickness ≥ 1 µm for solvent analysis

Temperature
- T_{max} 280 °C (long-term temperature), T_{max} 300 °C (short-term max. temperature in a temperature program)
- 0.53 mm ID: T_{max} 280 and 300 °C, resp.

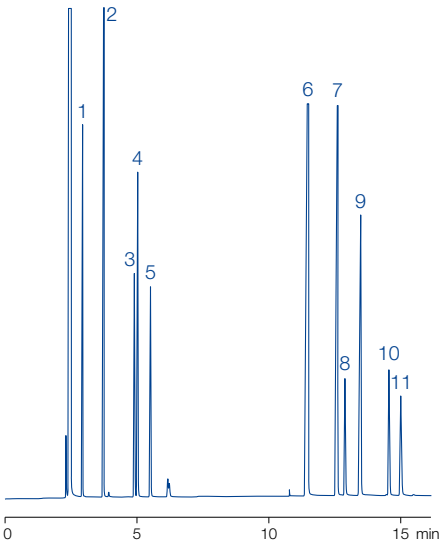
Similar phases
- OV-1701, DB-1701, CP-Sil 19 CB, HP-1701, Rtx®-1701, SPB™-1701, 007-1701, BP10, ZB-1701

Analysis of aromatic hydrocarbons

MN Appl. No. 200400

Column: OPTIMA® 1701, 25 m x 0.32 mm ID, 0.25 µm film
Injection: 1 µL, split 1:40
Carrier gas: 0,6 bar N_2
Temperature: 60 °C → 120 °C, 4 °C/min
Detector: FID 260 °C

Peaks:
1. Benzene
2. Toluene
3. Ethylbenzene
4. *p*-Xylene
5. *o*-Xylene
6. Phenol
7. 2-Methylphenol
8. 2,6-Dimethylphenol
9. 4-Methylphenol
10. 2,4-Dimethylphenol
11. 2,4,6-Trimethylphenol



Ordering information

OPTIMA® 1701

Length →	10 m	15 m	25 m	30 m	50 m	60 m
0.2 mm ID (0.4 mm OD)						
0.20 µm film			726841.25		726841.50	
0.25 mm ID (0.4 mm OD)						
0.25 µm film	726058.10	726058.15	726058.25	726058.30	726058.50	726058.60
0.50 µm film				726064.30		726064.60
1.00 µm film				726965.30		
0.32 mm ID (0.5 mm OD)						
0.25 µm film	726318.10	726318.15	726318.25	726318.30	726318.50	726318.60
0.35 µm film			726824.25	726824.30	726824.50	726824.60
0.50 µm film			726320.25	726320.30	726320.50	726320.60
1.00 µm film			726929.25	726929.30	726929.50	726929.60
0.53 mm ID (0.8 mm OD)						
1.00 µm film	726545.10	726545.15	726545.25	726545.30		
2.00 µm film		726735.15	726735.25	726735.30	726735.50	

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

Further applications can be found online in our application database at *www.mn-net.com/apps*

OPTIMA® 1701 MS silarylene phase · USP G46

Key features

- Chemically bonded, cross-linked silarylene phase with selectivity similar to 14 % cyanopropyl-phenyl – 86 % dimethylpolysiloxane, symmetric substituted cyanopropylsilanes and integrated phenyl rings (silarylene)
- Midpolar phase with very low bleed
- Perfect deactivation
- Structure see page 307

Recommended application

- Environmental analysis (e.g., PAHs, PCBs, pesticides)
- Reference column for structure identification, e.g., in combination with OPTIMA® 5 MS
- 100 % ion trap and quadrupole MS compatibility

Temperature

- T_{max} 280 °C (long-term temperature), T_{max} 300 °C (short-term max. temperature in a temperature program)

Similar phases

- VF-1701ms, TG-1701MS, OV-1701, DB-1701, HP-1701, Rtx®-1701, SPB™-1701, CP Sil 19 CB, 007-1701, BP10, ZB-1701

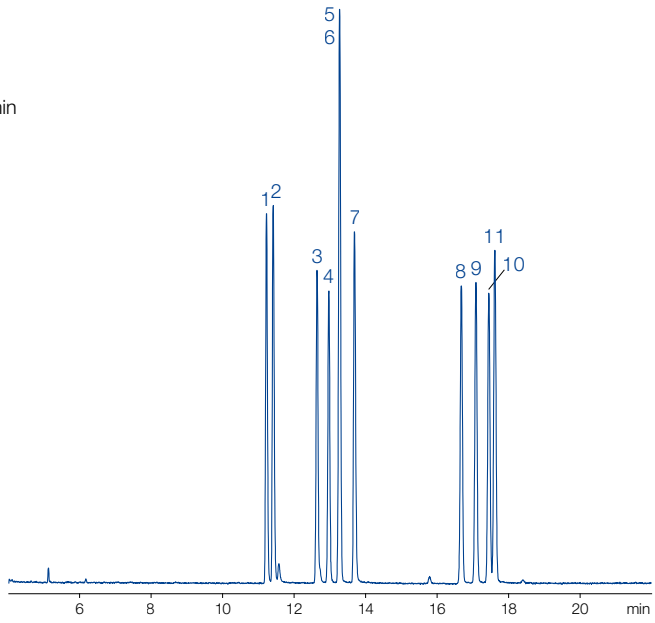
Separation of triazine pesticides (EPA 619)

MN Appl. No. 215080

Column:	OPTIMA® 1701 MS, 30 m x 0.25 mm ID, 0.25 µm film
Injection:	1 µL, 250 °C, split 1:100
Carrier gas:	42 cm/s He
Temperature:	160 °C (1 min) → 180 °C, 15 °C/min → 220 °C, 2 °C/min
Detector:	MSD

Peaks:

1. Prometon
2. Atraton
3. Propazine
4. Atrazine
5. Simazine
6. Terbuthylazine
7. Secbumeton
8. Prometryn
9. Ametryn
10. Simetryn
11. Terbutryn



Ordering information

OPTIMA® 1701 MS

	Length → 30 m	60 m
0.25 mm ID (0.4 mm OD)		
0.25 µm film	726630.30	726630.60
0.50 µm film	726631.30	726631.60
1.00 µm film	726632.30	726632.60
0.32 mm ID (0.5 mm OD)		
0.25 µm film	726633.30	726633.60
0.50 µm film	726634.30	726634.60
1.00 µm film	726635.30	726635.60

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

OPTIMA® 35 MS silarylene phase · USP G42 / close equivalent to USP G28 / G32

Key features

- Chemically bonded cross-linked silarylene phase with selectivity similar to 35 % phenyl – 65 % methyl polysiloxane, midpolar phase, polymer without CN groups
- Very low column bleeding
- Structure see page 309

Recommended application

- Ideal for ion trap detectors
- Optimum column for confirmation of analytical results in combination with a 1 MS or 5 MS
- All-round phase for environmental analysis, ultra trace analysis, EPA methods, pesticides, PCB, food and drug analysis

Temperature

- T_{max} 360 °C (long-term temperature), T_{max} 370 °C (short-term max. temperature in a temperature program)

Similar phases

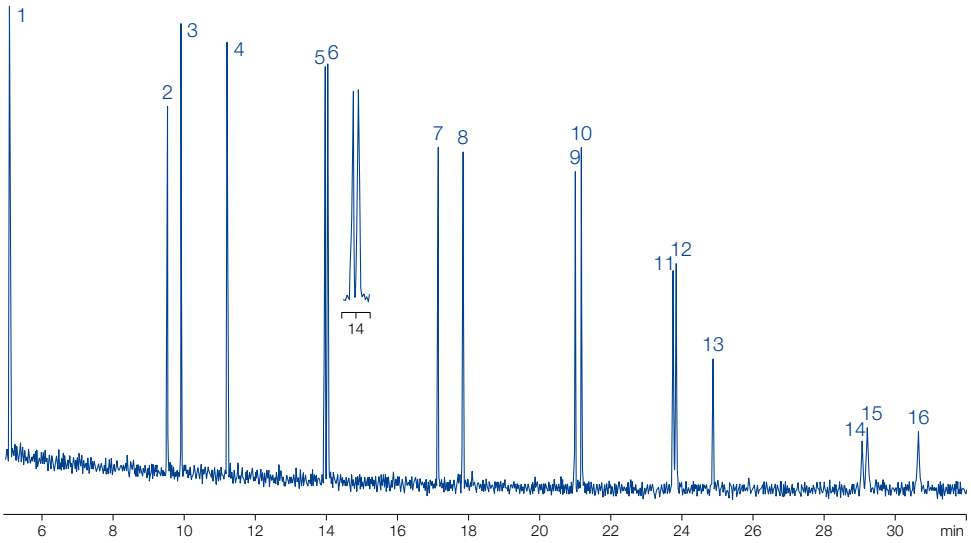
- DB-35 MS, HP-35, SPB™-35, Rxi®-35SIL MS, Rtx-35, 007-35, BPX™-35, MDN-35, AT™-35 MS, ZB-35, OV-11, VF-35 MS

PAH in accordance with EPA 610

MN Appl. No. 213190

Column: OPTIMA® 35 MS, 30 m x 0.25 mm ID, 0.25 µm film
Injection: 1 µL, split 1:10
Carrier gas: 0.6 bar H_2
Temperature: 100 °C (3 min) → 300 °C (10 min), 6 °C/min
Detector: MSD

Peaks
1. Naphthalene
2. Acenaphthylene
3. Acenaphthene
4. Fluorene
5. Phenanthrene
6. Anthracene
7. Fluoranthene
8. Pyrene
9. Benz[*a*]anthracene
10. Chrysene
11. Benzo[*b*]fluoranthene
12. Benzo[*k*]fluoranthene
13. Benzo[*a*]pyrene
14. Indeno[1,2,3-*cd*]pyrene
15. Dibenz[*ah*]anthracene
16. Benzo[*ghi*]perylene



Ordering information

OPTIMA® 35 MS

	Length → 30 m	60 m
0.25 mm ID (0.4 mm OD)		
0.25 µm film	726154.30	726154.60
0.32 mm ID (0.5 mm OD)		
0.25 µm film	726157.30	726157.60

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

Further applications can be found online in our application database at *www.mn-net.com/apps*

OPTIMA® 17 phenylmethylpolysiloxane (50 % phenyl) · USP G3

Key features
- Midpolar phase
- Structure see page 309

Recommended application
- Steroids, pesticide, drug analysis

Temperature
- T_{max} 320 °C (long-term temperature), T_{max} 340 °C (short-term max. temperature in a temperature program)
- 0.53 mm ID: T_{max} 300 and 320 °C resp.

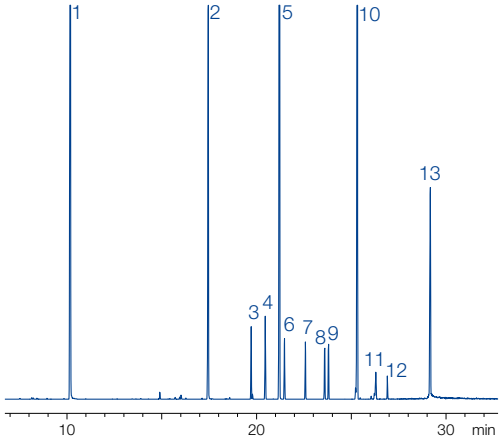
Similar phases
- OV-17, DB-17, HP-50+, HP-17, SPB™-50, SP-2250, Rxi®-17, Rtx®-50, CP-Sil 24 CB, 007-17, ZB-50

Analysis of pesticides

MN Appl. No. 200930

Column: OPTIMA® 17, 25 m x 0.2 mm ID, 0.20 µm film
Sample: pesticides, standard of the cantonal laboratory Schaffhausen (Switzerland), 0.1 mg/mL or 0.01 mg/mL each
Injection: 1.0 µL, 3 s without split
Carrier gas: He, 25 cm/s
Temperature: 100 °C (3 min), 8 °C/min → 250 °C, 10 °C/min → 320 °C
Detector: MSD HP 5971

Peaks:
1. Dichlorphos
2. Naled
3. Vinclozolin
4. Chlorthalonil
5. Chlorpyrifos
6. Dichlofluanid
7. Procymidon
8. Captan
9. Folpet
10. Carbophenothion
11. Iprodion
12. Captafol
13. Coumaphos



Ordering information

OPTIMA® 17

	Length → 10 m	12 m	15 m	25 m	30 m	50 m	60 m
0.1 mm ID (0.4 mm OD)							
0.10 µm film	726848.10						
0.2 mm ID (0.4 mm OD)							
0.20 µm film		726065.12		726065.25		726065.50	
0.50 µm film				726066.25		726066.50	
0.25 mm ID (0.4 mm OD)							
0.15 µm film				726742.25	726742.30	726742.50	726742.60
0.25 µm film			726022.15	726022.25	726022.30	726022.50	726022.60
0.50 µm film				726067.25	726067.30	726067.50	726067.60
0.32 mm ID (0.5 mm OD)							
0.15 µm film					726755.30		
0.25 µm film				726351.25	726351.30	726351.50	726351.60
0.35 µm film				726757.25	726757.30	726757.50	726757.60
0.50 µm film				726744.25	726744.30	726744.50	726744.60
0.53 mm ID (0.8 mm OD)							
1.00 µm film	726747.10		726747.15	726747.25	726747.30		

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

OPTIMA® 17 MS silarylene phase · USP G3

Key features

- Medium polar silarylene phase with selectivity analogue to 50 % phenyl – 50 % methylpolysiloxane, no CN groups in the polymer
- Structure see page 309

Recommended application

- Ideal for ion trap detectors
- Optimum reference column in combination with a 1 MS or 5 MS
- All-round phase for environmental analysis, ultra-trace analysis, EPA methods, pesticide, PCBs, food and drug analysis

Temperature

- T_{max} 340 °C (long-term temperature),
- T_{max} 360 °C (short-term max. temperature in a temperature program)

Similar phases

- OV-17, AT™-50, BPX™-50, DB-17, DB-17ms, HP-50+, HP-17, SPB™-50, SPB™-17, SP-2250, Rtx®-50, CP-Sil 24 CB, 007-17, VF-17ms, ZB-50

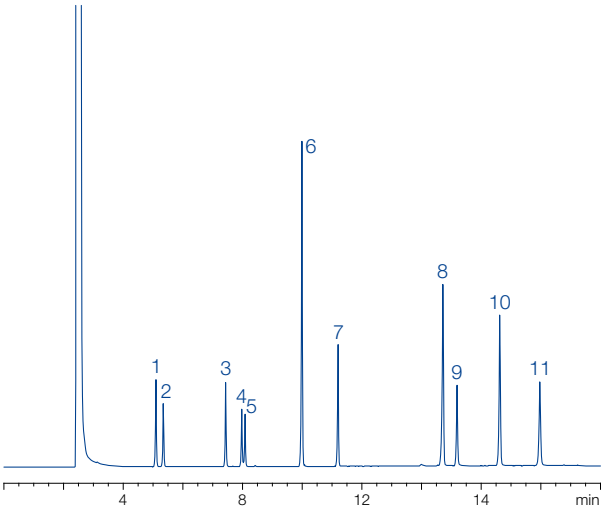
Analysis of phenols

MN Appl. No. 213600

Column:	OPTIMA® 17 MS, 30 m x 0.25 mm ID, 0.25 µm film
Sample:	phenol mix 604
Injection:	1.0 µL, 230 °C, split 1:30
Carrier gas:	0.8 bar He
Temperature:	100 °C, 10 °C/min → 250 °C
Detector:	FID 280 °C

Peaks:

1. Phenol
2. 2-Chlorophenol
3. 2,4-Dimethylphenol
4. 2-Nitrophenol
5. 2,4-Dichlorophenol
6. 4-Chloro-3-methylphenol
7. 2,4,6-Trichlorophenol
8. 4-Nitrophenol
9. 2,4-Dinitrophenol
10. 2-Methyl-4,6-dinitrophenol
11. Pentachlorophenol



Ordering information

OPTIMA® 17 MS

	Length → 30 m	60 m
0.25 mm ID (0.4 mm OD)		
0.25 µm film	726162.30	726162.60
0.32 mm ID (0.5 mm OD)		
0.25 µm film	726165.30	726165.60

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

Further applications can be found online in our application database at *www.mn-net.com/apps*

OPTIMA® 210 trifluoropropyl-methylpolysiloxane (50 % trifluoropropyl) · close equivalent to USP G6

Key features
- Midpolar phase
- Structure see page 309

Recommended application
- Environmental analysis, especially for *o*-, *m*- and *p*-substituted aromatic hydrocarbons

Temperature
- T_{max} 260 °C (long-term temperature), T_{max} 280 °C (short-term max. temperature in a temperature program)

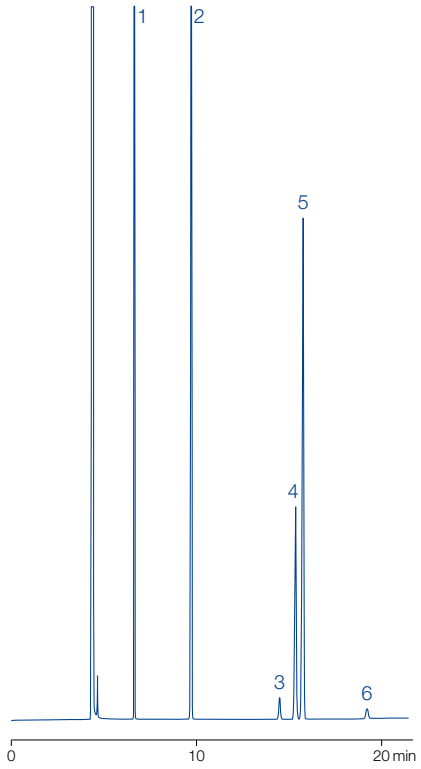
Similar phases
- OV-210, DB-210, Rtx®-200, 007-210

Aromatic hydrocarbons (BTX)

MN Appl. No. 200230

Column: OPTIMA® 210, 50 m x 0.25 mm ID, 0.5 µm film
Injection: 0.5 µL, split 105 mL/min
Carrier gas: 130 kPa N_2 (1.1 mL/min)
Temperature: 50 °C
Detector: FID 250 °C

Peaks:
1. Benzene
2. Toluene
3. Ethylbenzene
4. *p*-Xylene
5. *m*-Xylene
6. *o*-Xylene



Ordering information

OPTIMA® 210

	Length → 15 m	25 m	30 m	50 m	60 m
0.25 mm ID (0.4 mm OD)					
0.25 µm film	726871.15	726871.25	726871.30	726871.50	726871.60
0.50 µm film			726874.30	726874.50	726874.60
0.32 mm ID (0.5 mm OD)					
0.25 µm film	726877.15		726877.30	726877.50	726877.60
0.50 µm film		726880.25	726880.30	726880.50	726880.60

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

OPTIMA® 225 50 % cyanopropyl-methyl – 50 % phenylmethylpolysiloxane · Close equivalent to USP G7 / G19

Key features
- Midpolar phase
- Structure see page 309

Recommended application
- Fatty acid analysis

Temperature
- T_{max} 260 °C (long-term temperature), T_{max} 280 °C (short-term max. temperature in a temperature program)

Similar phases
- OV-210, DB-210, Rtx®-200, 007-210

Analysis of FAME in porcine fat

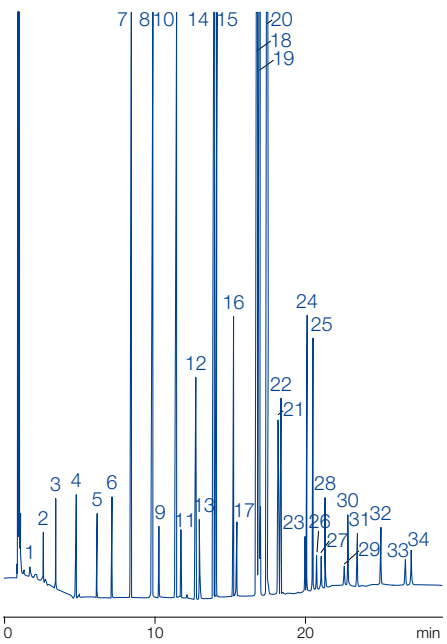
MN Appl. No. 210060

Column: OPTIMA® 225, 25 m x 0.32 mm ID, 0.25 µm film
Injection: 1 µL, split 1:40
Carrier gas: 60 kPa H_2
Temperature: 50 °C (2 min) → 125 °C, 30 °C/min → 160 °C, 5 °C/min → 180 °C, 20 °C/min → 200 °C, 3 °C/min → 220 °C, 20 °C/min (10 min)
Detector: FID 260 °C

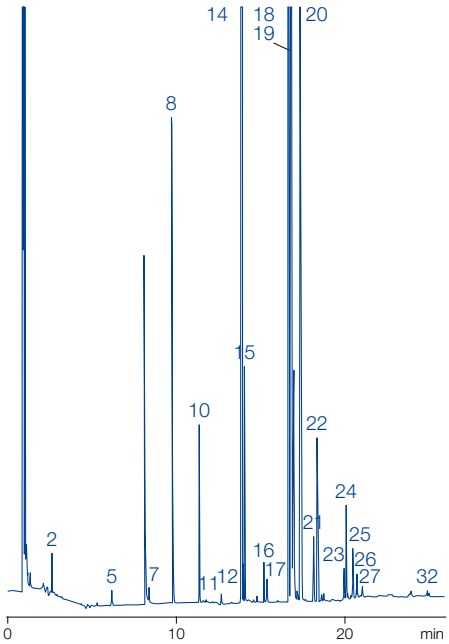
Peaks:

1. C4:0	18. C18:0
2. C5:0	19. C18:1
3. C6:0	20. C18:2
4. C8:0	21. C18:3
5. C10:0	22. C19:0
6. C11:0	23. C20:0
7. C12:0	24. C20:1
8. C13:0	25. C20:2
9. C13:1	26. C20:4
10. C14:0	27. C20:3
11. C14:1	28. C20:5
12. C15:0	29. C22:0
13. C15:1	30. C22:1
14. C16:0	31. C22:2
15. C16:1	32. C22:6
16. C17:0	33. C24:0
17. C17:1	34. C24:1

FAME Standard



FAME in porcine fat



Courtesy of Dr. Bantleon, Mr. Leusche, Mr. Hagemann, VFG-Labor, Versmold, Germany

Ordering information

OPTIMA® 225

Length →	10 m	15 m	25 m	30 m	50 m	60 m
0.1 mm ID (0.4 mm OD)						
0.10 µm film	726080.10					
0.25 mm ID (0.4 mm OD)						
0.25 µm film		726118.15	726118.25	726118.30	726118.50	726118.60
0.32 mm ID (0.5 mm OD)						
0.25 µm film			726352.25	726352.30	726352.50	726352.60

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

OPTIMA® 240 33 % cyanopropyl-methyl – 67 % dimethylpolysiloxane

Key features
- Midpolar phase
- Structure see page 309

Recommended application
- FAMEs, dioxins

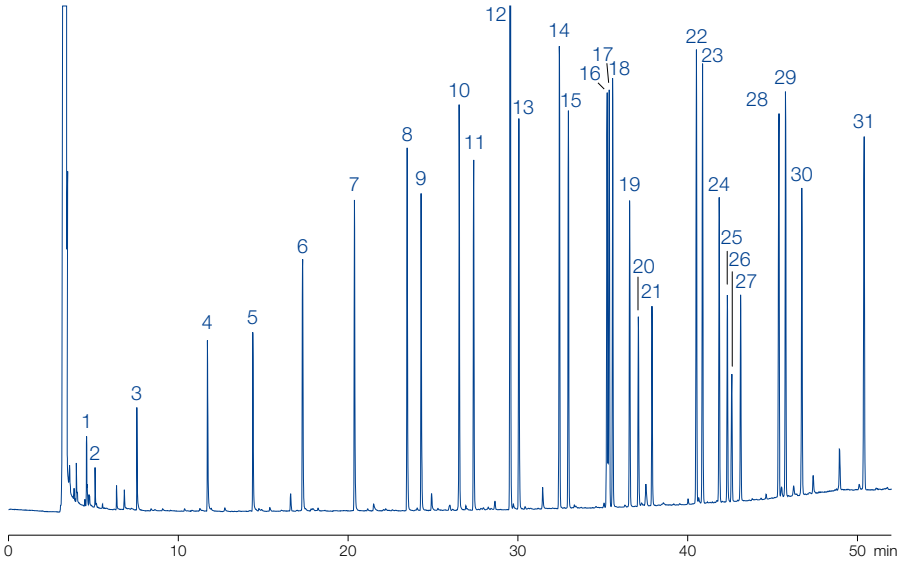
Temperature
- T_{max} 260 °C (long-term temperature), T_{max} 280 °C (short-term max. temperature in a temperature program)

Fatty acid methyl esters *cis/trans* C18:1 (FAME)

MN Appl. No. 201620

Column: OPTIMA® 240, 60 m x 0.25 mm ID, 0.25 µm film
Sample: FAME mixture
Injection: 1.0 µL, split 1:25
Carrier gas: 150 kPa H_2
Temperature: 80 °C → 120 °C, 20 °C/min → 260 °C (10 min), 3 °C/min
Detector: FID 280 °C

Peaks:
1. C4:0
2. C5:0
3. C8:0
4. C10:0
5. C11:0
6. C12:0
7. C13:0
8. C14:0
9. C14:1
10. C15:0
11. C15:1
12. C16:0
13. C16:1
14. C17:0
15. C17:1
16. C18:0
17. *trans*-C18:1
18. *cis*-C18:1
19. C18:2
20. C18:3
21. C18:3
22. C20:0
23. C20:1
24. C20:2
25. C20:3
26. C20:4
27. C20:3
28. C22:0
29. C22:1
30. C22:3
31. C24:1



Ordering information

OPTIMA® 240

	Length → 25 m	30 m	50 m	60 m
0.25 mm ID (0.4 mm OD)				
0.25 µm film		726089.30	726089.50	726089.60
0.50 µm film		726090.30		726090.60
0.32 mm ID (0.5 mm OD)				
0.25 µm film	726091.25	726091.30	726091.50	726091.60
0.35 µm film		726095.30		726095.60
0.50 µm film		726096.30		726096.60

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

Further applications can be found online in our application database at *www.mn-net.com/apps*

OPTIMA® WAX polyethylene glycol 20 000 Da · USP G16

Key features
- Polar phase
- Structure see page 309

Recommended application
- Solvent analysis and alcohols, suitable for aqueous solutions

Temperature
- T_{max} 240 °C (long-term temperature), T_{max} 250 °C (short-term max. temperature in a temperature program)
- 0.53 mm ID: T_{max} 220 and 240 °C resp.

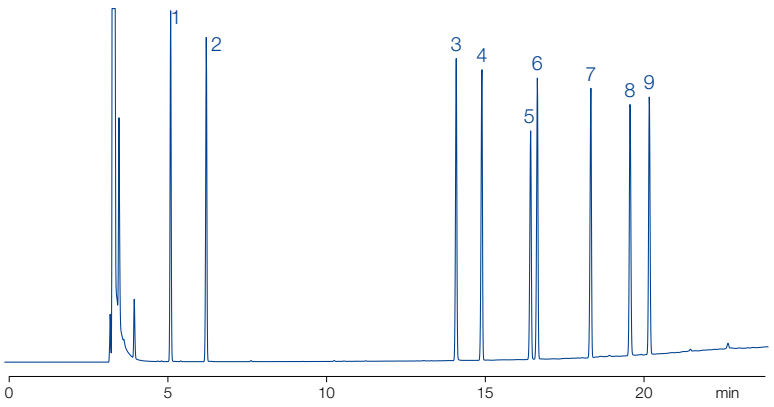
Similar phases
- PERMABOND® CW 20 M (see page 337), DB-Wax, Supelcowax, HP-Wax, HP-INNOWAX, Rtx-Wax, CP-Wax 52 CB, Stabilwax, 007-CW, BP20, AT-Wax, ZB-Wax

Modified Grob test

MN Appl. No. 211170

Column: OPTIMA® WAX, 50 m x 0.32 mm ID, 0.5 µm film
Injection: 1 µL, split 1:20
Carrier gas: 1,2 bar He
Temperature: 80 °C → 250 °C, 8 °C/min
Detector: FID 250 °C

Peaks:
1. Decane
2. Undecane
3. Octanol
4. Methyl decanoate
5. Dicyclohexylamine
6. Methyl undecanoate
7. Methyl dodecanoate
8. 2,6-Dimethylaniline
9. 2,6-Dimethylphenol



Ordering information

OPTIMA® WAX

	Length → 25 m	30 m	50 m	60 m
0.25 mm ID (0.4 mm OD)				
0.25 µm film	726600.25	726600.30	726600.50	726600.60
0.32 mm ID (0.5 mm OD)				
0.25 µm film	726321.25	726321.30	726321.50	726321.60
0.50 µm film	726296.25	726296.30	726296.50	726296.60
0.53 mm ID (0.8 mm OD)				
1.00 µm film	726549.25	726549.30		
2.00 µm film		726548.30		

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

Further applications can be found online in our application database at *www.mn-net.com/apps*

OPTIMA WAXplus® cross-linked polyethylene glycol · USP G16

Key features
- Polar phase with improved cross-linking for lower column bleed and better temperature stability
- Structure see page 309

Recommended application
- Broad range of application, e.g., for solvents and alcohols, suitable for aqueous solutions

Temperature
- T_{max} 260 °C (long-term temperature), T_{max} 270 °C (short-term max. temperature in a temperature program)

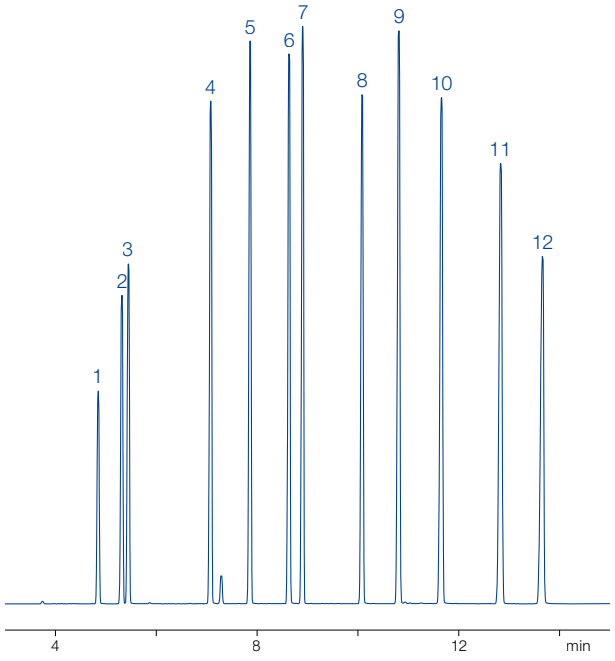
Similar phases
- DB-Wax, Supelcowax, HP-Wax, HP-INNOWAX, Rtx-Wax, CP-Wax 52 CB, Stabilwax, 007-CW, BP20, AT-Wax, ZB-Wax

Alcohols

MN Appl. No. 214160

Column: OPTIMA WAXplus®, 30 m x 0.25 mm ID, 0.5 µm film
Injection: 0.1 µL, split 1:80
Carrier gas: 1.3 bar He
Temperature: 40 °C → 260 °C, 12 °C/min (15 min)
Detector: FID 260 °C

Peaks:
1. Methanol
2. 2-Propanol
3. Ethanol
4. 1-Propanol
5. 2-Methyl-1-propanol
6. 1-Butanol
7. 4-Methyl-2-pentanol
8. 1-Pentanol
9. 2-Methyl-1-pentanol
10. 1-Hexanol
11. Cyclohexanol
12. 1-Heptanol



Ordering information

OPTIMA WAXplus®

	Length → 30 m	60 m
0.25 mm ID (0.4 mm OD)		
0.25 µm film	726380.30	726380.60
0.50 µm film	726381.30	726381.60
0.32 mm ID (0.5 mm OD)		
0.25 µm film	726382.30	726382.60
0.50 µm film	726383.30	726383.60

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

OPTIMA® FFAP polyethylene glycol 2-nitroterephthalate · USP G35 / close equivalent to USP G25

Key features
- Polar phase (FFAP = Free Fatty Acid Phase)
- Structure see page 309

Recommended application
- Fatty acid methyl esters (FAMEs), free carboxylic acids

Temperature
- 0.10–0.32 mm ID: T_{max} 250 °C (long-term temperature), T_{max} 260 °C (short-term max. temperature in a temperature program)
- 0.53 mm ID: T_{max} 220 and 240 °C, resp.

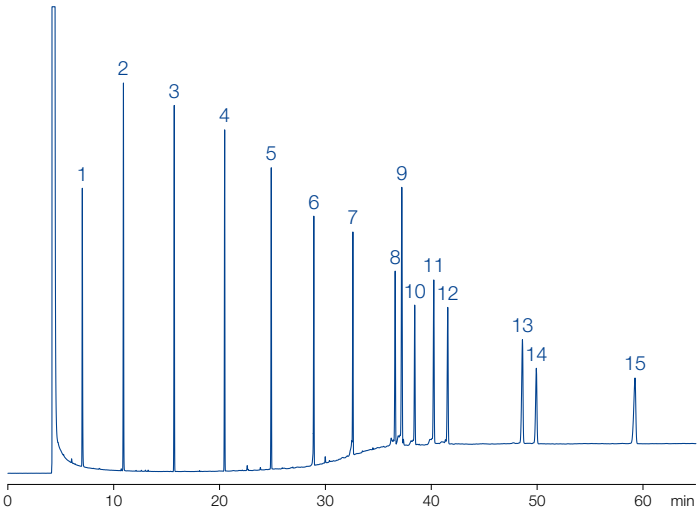
Similar phases
- PERMABOND® FFAP (see page 338), DB-FFAP, HP-FFAP, CP-Wax 58 FFAP CB, 007-FFAP, CP-FFAP CB, Nukol™, AT-1000, SPB-1000, BP21, OV-351

FAME test

MN Appl. No. 211140

Column: OPTIMA® FFAP, 60 m x 0.32 mm ID, 0.25 µm film
Injection: 1.0 µL, 220 °C, split 1:40
Carrier gas: 1.2 bar He
Temperature: 55 °C → 250 °C, 6 °C/min
Detector: FID 220 °C

Peaks:
1. C4
2. C6
3. C8
4. C10
5. C12
6. C14
7. C16
8. C18
9. C18:1 *cis/trans*
10. C18:2
11. C18:3
12. C20
13. C22
14. C22:1
15. C24



Ordering information

OPTIMA® FFAP

	Length → 10 m	25 m	30 m	50 m	60 m
0.1 mm ID (0.4 mm OD)					
0.10 µm film	726180.10				
0.25 mm ID (0.4 mm OD)					
0.25 µm film		726116.25	726116.30	726116.50	726116.60
0.32 mm ID (0.5 mm OD)					
0.25 µm film		726341.25	726341.30	726341.50	726341.60
0.50 µm film		726344.25	726344.30	726344.50	
0.53 mm ID (0.8 mm OD)					
0.50 µm film			726345.30		
1.00 µm film		726346.25			

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

OPTIMA® FFAPplus polyethylene glycol 2-nitroterephthalate · USP G35 / close equivalent to G25

Key features
- Polar phase
- Structure see page 309

Recommended application
- FAMEs, free carboxylic acids

Temperature
- T_{max} 250 °C (long-term temperature), T_{max} 260 °C (short-term max. temperature in a temperature program)

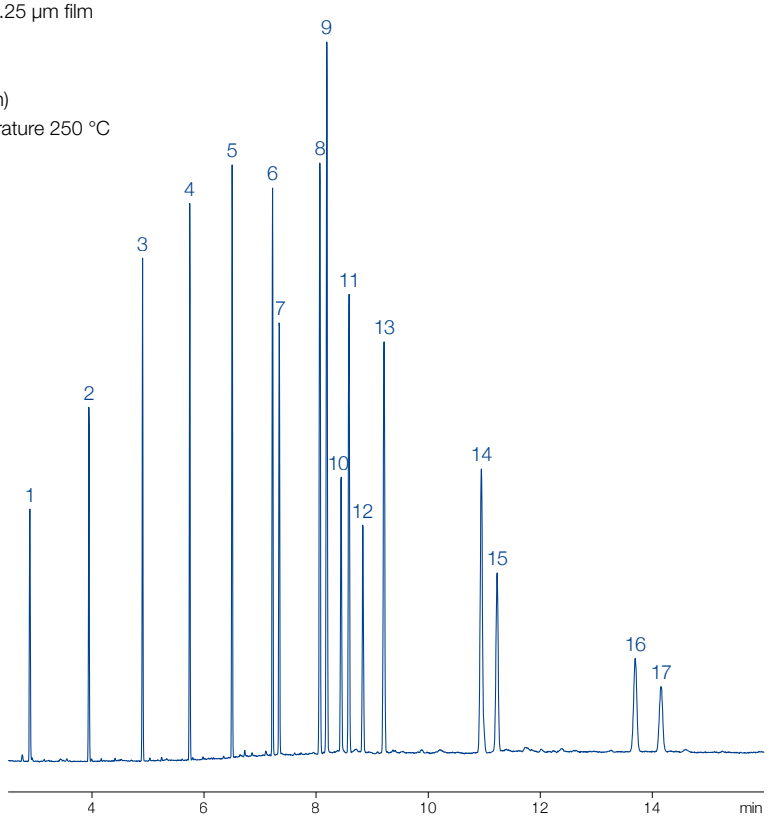
Similar phases
- DB-FFAP, HP-FFAP, CP-SIL 58 CB, 007-FFAP, CP-FFAP CB, Nukol™

FAMEs from biodiesel

MN Appl. No. 214590

Column: OPTIMA® FFAPplus, 30 m x 0.25 mm ID, 0.25 µm film
Injection: 1 µL, 260 °C, split 1:15
Carrier gas: 40 cm/s He
Temperature: 70 °C (1 min) → 240 °C, 30 °C/min (10 min)
Detector: MS-EI, ion source 200 °C, interface temperature 250 °C

Peaks:
Methyl esters of:
1. Caproic acid (C6:0)
2. Caprylic acid (C8:0)
3. Capric acid (C10:0)
4. Lauric acid (C12:0)
5. Myristic acid (C14:0)
6. Palmitic acid (C16:0)
7. Palmitoleic acid (C16:1)
8. Stearic acid (C18:0)
9. Oleic acid (C18:1 *cis*)
10. Linoleic acid (C18:2 *cis*)
11. Nonadecanoic acid (C19:0)
12. Linolenic acid (C18:3)
13. Arachidic acid (C20:0)
14. Behenic acid (C22:0)
15. Erucic acid (C22:1 *cis*)
16. Lignoceric acid (C24:0)
17. Nervonic acid (C24:1 *cis*)



Ordering information

OPTIMA® FFAPplus

	Length → 30 m	60 m
0.25 mm ID (0.4 mm OD)		
0.25 µm film	726241.30	726241.60
0.50 µm film	726242.30	726242.60
0.32 mm ID (0.5 mm OD)		
0.25 µm film	726243.30	726243.60
0.50 µm film	726246.30	726246.60

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

Further applications can be found online in our application database at *www.mn-net.com/apps*

PERMABOND® SE-30 100 % dimethylpolysiloxane · USP G1 / G2 / G38

Key features

· Nonpolar phase

Temperature

· T_{max} 300 °C (long-term temperature),
T_{max} 320 °C (short-term max. temperature in a temperature program)

Similar phases

· OPTIMA® 1 (see page 310)

Ordering information

PERMABOND® SE-30

	Length → 25 m	50 m
0.25 mm ID (0.4 mm OD)		
0.25 µm film	723052.25	723052.50
0.32 mm ID (0.5 mm OD)		
0.25 µm film	723306.25	
0.50 µm film		723308.50

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

PERMABOND® SE-52 5 % phenyl – 95 % dimethylpolysiloxane · USP G27

Key features

· Nonpolar phase

Temperature

· T_{max} 300 °C (long-term temperature),
T_{max} 320 °C (short-term max. temperature in a temperature program)

Similar phases

· OPTIMA® 5 (see page 314)

Ordering information

PERMABOND® SE-52

	Length → 25 m
0.25 mm ID (0.4 mm OD)	
0.25 µm film	723054.25
0.32 mm ID (0.5 mm OD)	
0.25 µm film	723310.25
0.50 µm film	723312.25

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

PERMABOND® CW 20 M polyethylene glycol 20 000 Dalton · USP G16

Key features
- Polar phase

Recommended application
- Solvent analysis and alcohols, suitable for aqueous solutions

Temperature
- 0.1–0.32 mm ID: T_{max} 220 °C (long-term temperature), T_{max} 240 °C (short-term max. temperature in a temperature program)
- 0.53 mm ID: T_{max} 200 and 220 °C, resp.

Similar phases
- See OPTIMA® WAX (see page 332)

Ordering information

PERMABOND® CW 20 M

	Length → 10 m	25 m	30 m	50 m	60 m
0.1 mm ID (0.4 mm OD)					
0.10 µm film	723064.10				
0.25 mm ID (0.4 mm OD)					
0.25 µm film	723060.10	723060.25	723060.30	723060.50	723060.60
0.32 mm ID (0.5 mm OD)					
0.25 µm film	723321.10	723321.25	723321.30	723321.50	723321.60
0.35 µm film	723827.10	723827.25		723827.50	
0.50 µm film	723296.10	723296.25	723296.30	723296.50	723296.60
0.53 mm ID (0.8 mm OD)					
0.50 µm film	723515.10	723515.25			
1.00 µm film	723549.10	723549.25	723549.30		
2.00 µm film	723517.10	723517.25	723517.30		

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

Further applications can be found online in our application database at *www.mn-net.com/apps*

PERMABOND® FFAP polyethylene glycol 2-nitroterephthalate · USP G35 / close equivalent to G25

Key features
- Polar phase

Recommended application
- FAMEs, free carboxylic acids

Temperature
- 0.1–0.32 mm ID: T_{max} 220 °C (long-term temperature), T_{max} 240 °C (short-term max. temperature in a temperature program)
- 0.53 mm ID: T_{max} 200 and 220 °C, resp.

Similar phases
- See OPTIMA® FFAP (see page 334)

Ordering information

PERMABOND® FFAP

	Length → 10 m	20 m	25 m	30 m	50 m	60 m
0.1 mm ID (0.4 mm OD)						
0.10 µm film	723180.10	723180.20				
0.25 µm film	723181.10					
0.25 mm ID (0.4 mm OD)						
0.10 µm film			723936.25		723936.50	
0.25 µm film	723116.10		723116.25	723116.30	723116.50	723116.60
0.32 mm ID (0.5 mm OD)						
0.10 µm film			723356.25		723356.50	
0.25 µm film			723341.25	723341.30	723341.50	723341.60
0.35 µm film	723830.10		723830.25		723830.50	
0.50 µm film	723344.10		723344.25	723344.30	723344.50	723344.60
0.53 mm ID (0.8 mm OD)						
1.00 µm film	723555.10		723555.25		723555.50	

In addition to this standard program we will be happy to supply columns custom-made to your specifications. Information about scope of delivery, special cages and integrated guard columns see additional information for GC columns on page 303.

Further applications can be found online in our application database at *www.mn-net.com/apps*

Capillary columns for special GC separations

Certain analytical separations can be accomplished more easily with chromatographic columns, that have been especially developed for that task, compared with standard columns. The following table summarizes our program of GC speciality capillaries, the individual columns will be described in detail on the following pages.

Overview

Separation/special application	Recommended capillary column	Page
Fast GC column with 0.10 mm ID		
	OPTIMA® 1, OPTIMA® 5, OPTIMA® δ-3, OPTIMA® δ-6 OPTIMA® 17, OPTIMA® 225, OPTIMA® FFAP PERMABOND® CW 20 M, PERMABOND® FFAP	340
Enantiomer separation cyclodextrin phases		
	FS-LIPODEX® A, FS-LIPODEX® B, FS-LIPODEX® C FS-LIPODEX® D, FS-LIPODEX® E, FS-LIPODEX® G	342
	FS-HYDRODEX β-PM, FS-HYDRODEX β-3 P, FS-HYDRODEX β-6TBDM, FS-HYDRODEX β-6TBDE, FS-HYDRODEX β-6TBDE, FS-HYDRODEX β-TBDAc, FS-HYDRODEX γ-DiMOM	344
Biodiesel		
Methanol analysis	OPTIMA® BioDiesel M	346
FAME analysis	OPTIMA® BioDiesel F	346
Glycerol and triglycerides	OPTIMA® BioDiesel G	346
Triglycerides		
	OPTIMA® 1-TG	348
	OPTIMA® 17-TG	348
High temperature GC		
	OPTIMA® 5 HT	349
Amines		
Polyfunctional amines	OPTIMA® 5 Amine	350
Amine separations	FS-CW 20 M-AM	351
Petrochemical products (complex hydrocarbon mixtures)		
	PERMABOND® P-100	352
Environmental analysis of volatile halogenated hydrocarbons		
	PERMABOND® SE-54 HKW	352
Silanes (monomeric, e.g., chlorosilanes)		
	PERMABOND® Silane	354
Diethylene glycol, e.g., for the quality control of wine		
	PERMABOND® CW 20 M-DEG	354

Capillary columns for Fast GC

Fast GC

Key features

- Decreased column diameters, high heating rates and decreased column lengths for faster GC separations with high resolution efficiency
- Small inner diameters combined with very fast temperature programs can reduce the analysis time by up to 80 %
- High sensitivity detectors with small volume and very short response time, as well as very rapid data acquisition and processing
- Small inner diameters result in high column inlet pressures and a lower volume flow of the mobile phase: very fast injection of very small samples against a high pressure
- Amount of sample, which can be injected, is limited by the inner diameter and the thin film

Temperature

- High heating rates place special demands on stationary phases. OPTIMA® columns meet exactly this requirement: very low bleeding, long lifetimes, even for continuous high heating rates

Comparison of a separation on a 50 m standard capillary with separation on a 10 m fast GC column

MN Appl. No. 211260

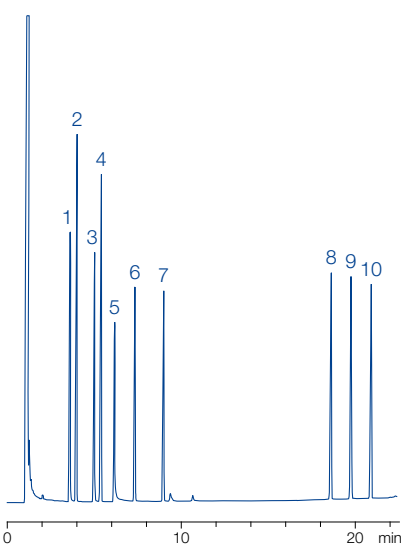
Peaks:

1. Octanol
2. Undecane
3. Dimethylaniline
4. Dodecane
5. Decylamine
6. Methyl decanoate
7. Methyl undecanoate
8. Henicosane
9. Docosane
10. Tricosane

A) Fast GC column

Column: OPTIMA® 5, 10 m x 0.1 mm ID, 0.1 µm film

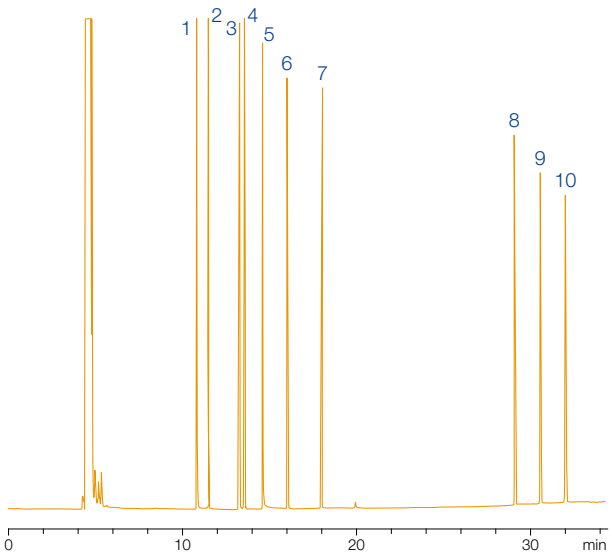
Injection 1 µL, split 1:40,
Carrier gas 0.75 bar He



B) standard GC column

Column: OPTIMA® 5, 50 m x 0.25 mm ID, 0.25 µm film

Injection 1 µL, split 1:35,
Carrier gas 1.5 bar He



Both separations:

Temperature: 80 °C → 320 °C (10 min), 8 °C/min

Detector: FID

While maintaining the temperature program and halving the pressure a time saving of 30 % results with identical separation efficiency.

Ordering information

Columns for Fast GC

Phase	Maximum temperature	ID [mm]	Film thickness [µm]	REF (10 m)	REF (20 m)
OPTIMA® 1					
	340 / 360 °C	0.10	0.10	726024.10	726024.20
		0.10	0.40		726025.20
OPTIMA® 5					
	340 / 360 °C	0.10	0.10	726846.10	
OPTIMA® δ-3					
	340 / 360 °C	0.10	0.10	726410.10	726410.20
OPTIMA® δ-6					
	340 / 360 °C	0.10	0.10	726490.10	
OPTIMA® 17					
	320 / 340 °C	0.10	0.10	726848.10	
OPTIMA® 225					
	260 / 280 °C	0.10	0.10	726080.10	
OPTIMA® FFAP					
	250 / 260 °C	0.10	0.10	726180.10	
PERMABOND® CW 20 M					
	220 / 240 °C	0.10	0.10	723064.10	
PERMABOND® FFAP					
	220 / 240 °C	0.10	0.10	723180.10	723180.20
		0.10	0.25	723181.10	
OPTIMA® 5 Amine					
	300 / 320 °C	0.10	0.40	726361.10	
FS-CW 20 M-AM					
	220 / 240 °C	0.10	0.25	733111.10	
FS-LIPODEX® E					
	200 / 220 °C	0.10	0.10	723382.10	
FS-HYDRODEX β-6TBDM					
	230 / 250 °C	0.10	0.10	723383.10	
In addition to this standard program, all MN GC phases can be custom-made as fast GC columns					

Further applications can be found online in our application database at *www.mn-net.com/apps*

Capillary columns for enantiomer separation

LIPODEX® cyclodextrin phases for enantiomer separation

Key features

- Base material: cyclic oligosaccharides consisting of six (α-cyclodextrin), seven (β-cyclodextrin) or eight (γ-cyclodextrin) glucose units bonded through 1,4-linkages
- Regioselective alkylation and / or acylation of the hydroxyl groups leads to lipophilic phases with varying enantioselectivity, which are well suited for GC enantiomer analysis
- Important advantage: many compounds can be analyzed without derivatization (however, for certain substances enantioselectivity can be favorably influenced by formation of derivatives)

Recommended application

- A large number of separations have been achieved, however, it is not possible to make a general prediction, which phase could solve a given separation task. Even for compounds with small structural differences or within homologous series the enantiodifferentiation can be quite different. The following table shows typical applications.

Note:

- Water as solvent is strictly forbidden for all cyclodextrin phases
- Dry the sample with our CHROMAFIX® Dry (Na_2SO_4) cartridges (see page 61)
- Use suitable nonpolar solvent

Phase	Cyclodextrin derivate	T_{max} [°C]	Recommended application
LIPODEX® A			
	hexakis-(2,3,6-tri-O-pentyl)-α-CD	200 / 220	carbohydrates, polyols, diols, hydroxycarboxylic acid esters, (epoxy-) alcohols, glycerol derivatives, spiroacetals, ketones, alkyl halides
LIPODEX® B			
	hexakis-(2,6-di-O-pentyl-3-O-acetyl)-α-CD	200 / 220	lactones, diols (cyclic carbonates), aminols, aldols (O-TFA), glycerol derivatives (cyclic carbonates)
LIPODEX® C			
	heptakis-(2,3,6-tri-O-pentyl)-β-CD	200 / 220	Alcohols, cyanhydrins, olefins, hydroxycarboxylic acid esters, alkyl halides
LIPODEX® D			
	heptakis-(2,6-di-O-pentyl-3-O-acetyl)-β-CD	200 / 220	aminols (TFA), β-amino acid esters, trans-cycloalkane-1,2-diols, trans-cycloalkane-1,2- diols, trans-cycloalkane-1,3-diols (TFA)
LIPODEX® E			
	octakis-(2,6-di-O-pentyl-3-O-butyryl)-γ-CD	200 / 220	α-amino acids, α- and β-hydroxycarboxylic acid esters, alcohols (TFA), diols (TFA), ketones, pheromones (cyclic acetals), amines, alkyl halides, lactones
LIPODEX® G			
	octakis-(2,3-di-O-pentyl-6-O-methyl)-γ-CD	220 / 240	menthol isomers, ketones, alcohols, carboxylic acid esters, terpenes

Ordering information

LIPODEX®

Length →	10 m 0.10 mm ID	25 m 0.25 mm ID	50 m 0.25 mm ID
FS-LIPODEX® A			
		723360.25	723360.50
FS-LIPODEX® B			
		723362.25	723362.50
FS-LIPODEX® C			
		723364.25	723364.50
FS-LIPODEX® D			
		723366.25	723366.50
FS-LIPODEX® E			
	723382.10	723368.25	723368.50
FS-LIPODEX® G			
		723379.25	723379.50
All columns with 0.4 mm OD			

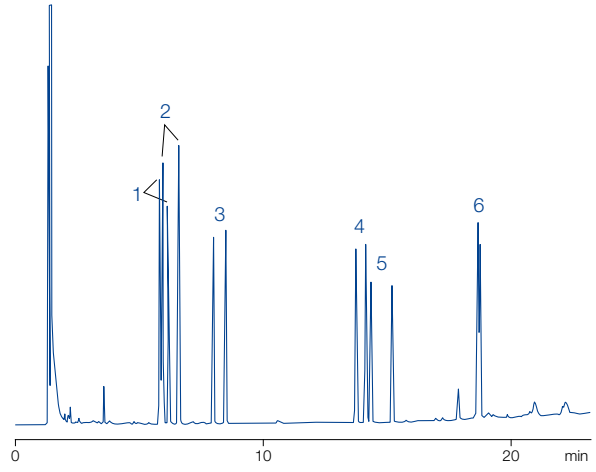
Capillary columns for enantiomer separation

Enantiomer separation of amino acid methyl esters (TFA)

MN Appl. No. 202592

Column:	FS-LIPODEX® E, 25 m x 0.25 mm ID
Injection:	1 µL, split ~ 1: 100
Carrier gas:	60 kPa H_2
Temperature:	90 → 190 °C, 4 °C/min
Detector:	FID 250 °C

Peaks:
(D is eluted before L except for proline: L before D)
1. Alanine
2. Valine
3. Leucine
4. Proline
5. Aspartic acid
6. Phenylalanine



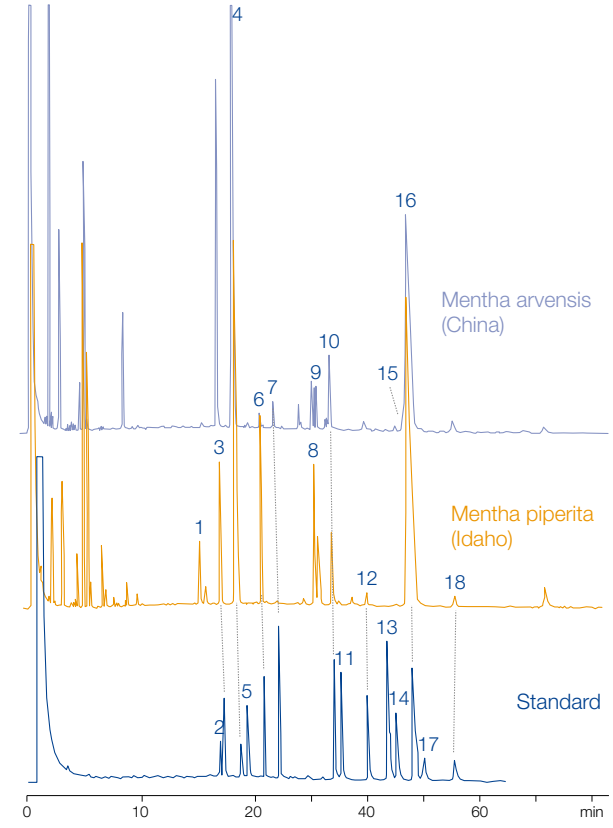
Separation of chiral constituents of peppermint oil

MN Appl. No. 250410

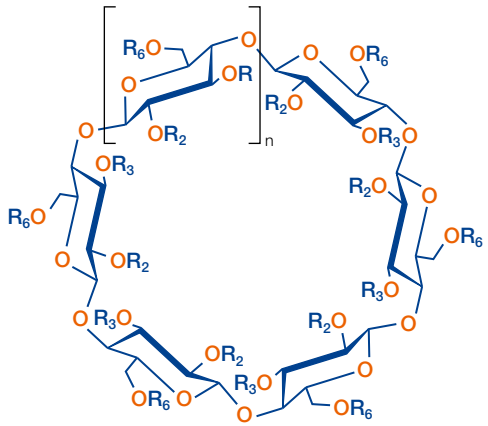
W. A. König et al., High Resol. Chromatogr. 20 (1997) 55–61

Column:	FS-LIPODEX® G, 25 m x 0.25 mm ID
Carrier gas:	50 kPa H_2
Temperature:	75 °C, isothermal
Detector:	FID

Peaks:
1. (+)-trans-Sabinene hydrate
2. (+)-Menthone
3. (+)-Isomenthone
4. (–)-Menthone
5. (–)-Isomenthone
6. (+)-Menthofuran
7. (–)-Isopulegol
8. (–)-Menthyl acetate
9. (+)-Pulegone
10. (+)-Neomenthol
11. (–)-Neomenthol
12. (+)-Neoisomenthol
13. (+)-Menthol
14. (–)-Neoisomenthol
15. (+)-Piperitone
16. (–)-Menthol
17. (+)-Isomenthol
18. (–)-Isomenthol



Cyclodextrin derivates



Further applications can be found online in our application database at *www.mn-net.com/apps*

Capillary columns for enantiomer separation

HYDRODEX cyclodextrin phases for enantiomer separation

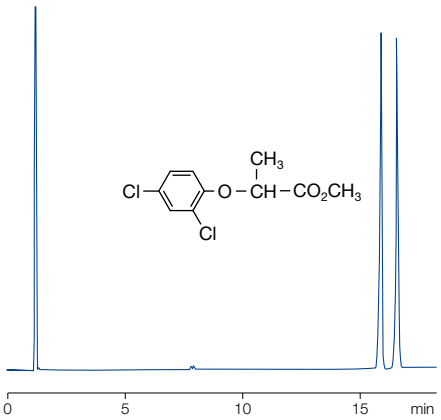
Recommended application

- Cyclodextrin derivatives (see page 343) with high melting point: for GC enantiomer separation diluted with polysiloxanes

Enantiomer separation of dichlorprop methyl ester

MN Appl. No. 202542

Column:	FS-HYDRODEX β-3P, 25 m x 0.25 mm ID
Injection:	0.1 µL (~1 % in CH_2Cl_2), split 130 mL/min
Carrier gas:	60 kPa H_2 (1.9 mL/min)
Temperature:	160 °C
Detector:	FID 250 °C



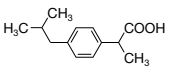
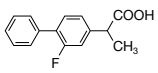
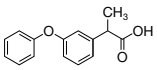
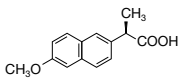
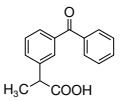
Separation of isomeric antiinflammatory drugs

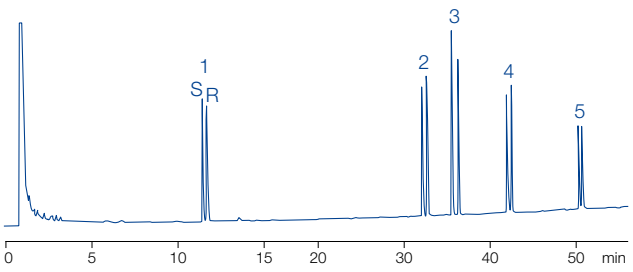
MN Appl. No. 210150

Courtesy of Prof. W.A. König, Hamburg, Germany

Column:	FS-HYDRODEX β-6TBDM, 25 m x 0.25 mm ID
Carrier gas:	He
Temperature:	135 °C → 200 °C, 1 °C/min
Detector:	FID

Peaks:

1. Ibuprofen 
2. Flurbiprofen 
3. Fenoprofen 
4. Naproxen 
5. Ketoprofen 



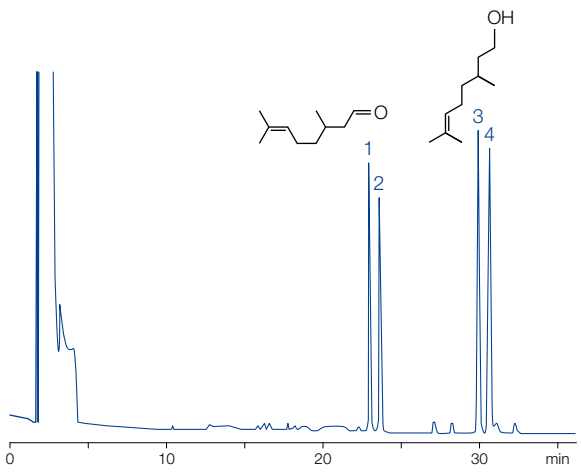
Phase	Cyclodextrin derivative (diluted with optimized polysiloxane)	T_{max} [°C]	Recommended application
HYDRODEX β-PM			
	heptakis-(2,3,6-tri-O-methyl)-β-CD	230 / 250	hydroxycarboxylic acid esters, alcohols, diols, olefins, lactones, acetals
HYDRODEX β-3P			
	heptakis-(2,6-di-O-methyl-3-O-pentyl)-β-CD	230 / 250	terpenes, dienes, allenes, terpene alcohols, 1,2-epoxy-alkanes, carboxylic acids (esters), hydroxycarboxylic acid esters, pharmaceuticals, pesticides
HYDRODEX β-6TBDM			
	heptakis-(2,3-di-O-methyl-6-O-t-butyldimethyl-silyl)-β-CD	230 / 250	γ-lactones, cyclopentanones, terpenes, esters, tartrates
HYDRODEX β-6TBDE			
	heptakis-(2,3-di-O-ethyl-6-O-t-butyldimethyl-silyl)-β-CD	230 / 250	essential oils
HYDRODEX β-TBDAc			
	heptakis-(2,3-di-O-acetyl-6-O-t-butyldimethyl-silyl)-β-CD	220 / 240	alcohols, esters, ketones, aldehydes, δ-lactones
HYDRODEX γ-TBDAc			
	octakis-(2,3-di-O-acetyl-6-O-t-butyldimethyl-silyl)-γ-CD	220 / 240	cyclic ketones, aromatic ketones, oxiranes, aromatic esters, aromatic amides
HYDRODEX γ-DIMOM			
	octakis-(2,3-di-O-methoxymethyl-6-O-t-butyldimethyl-silyl)-γ-CD	220 / 240	ketones, terpenes, cyclic ethers, alcohols, amines

Separation of (R/S) citronellol + citronellal

MN Appl. No. 212440

Column:	FS-HYDRODEX β-TBDAc, 50 m x 0.25 mm ID
Injection:	1 µL, 1:1000 in CH_2Cl_2, split 25 mL/min
Carrier gas:	1.5 bar H_2
Temperature:	100 °C
Detector:	FID 220 °C

Peaks:
1. (R)/(S)-Citronellal
2. (S)/(R)-Citronellal
3. (S)-Citronellol
4. (R)-Citronellol

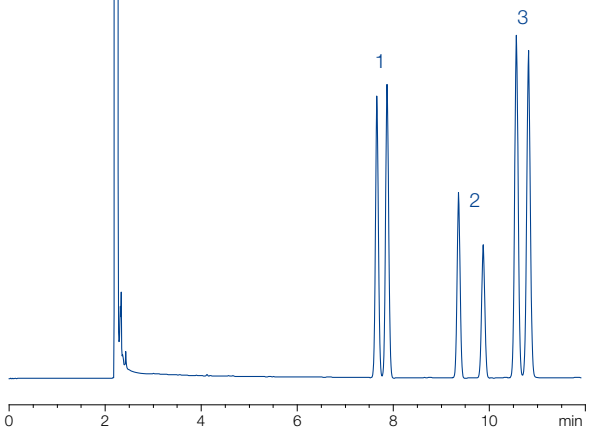


Separation of essential oils

MN Appl. No. 212980/212990/213000

Column:	FS-HYDRODEX γ-TBDAc, 50 m x 0.25 mm ID
Injektor:	220 °C
Carrier gas:	1.2 bar H_2
Temperature:	125 °C
Detector:	FID 220 °C

Peaks:
1. Fenchone (1.5 mg/mL)
2. Menthone (0.5 mg/mL)
3. Menthol (2 mg/mL)



Ordering information

HYDRODEX

Length →	10 m 0.10 mm ID	25 m 0.25 mm ID	50 m 0.25 mm ID
FS-HYDRODEX β-PM			
		723370.25	723370.50
FS-HYDRODEX β-3P			
		723358.25	723358.50
FS-HYDRODEX β-6TBDM			
	723383.10	723381.25	723381.50
FS-HYDRODEX β-6TBDE			
		723386.25	
FS-HYDRODEX β-TBDAc			
		723384.25	723384.50
FS-HYDRODEX γ-TBDAc			
		723387.25	723387.50
FS-HYDRODEX γ-DiMOM			
		723388.25	723388.50
All columns with 0.4 mm OD			

Further applications can be found online in our application database at *www.mn-net.com/apps*

Capillary columns for biodiesel analysis

OPTIMA® BioDiesel for the analysis of biodiesel (DIN EN 14214 / ASTM D 6751)

OPTIMA® BioDiesel M for analysis of methanol in accordance with DIN EN 14110

Key features

- The methanol content in biodiesel as specified in DIN EN 14110 must not exceed 0.2 %. The column OPTIMA® Bio-Diesel M allows the GC headspace analysis of the methanol content in biodiesel in the concentration range from 0.01 to 0.5 % with 2-propanol as internal standard.

Temperature

- T_{max} 340 °C (long-term temperature), T_{max} 360 °C (short-term max. temperature in a temperature program)

Similar phases

- Select™ Biodiesel for Methanol, Trace TR-BioDiesel (M)

OPTIMA® BioDiesel F for analysis of FAMEs in accordance with DIN EN 14103:2011

Key features

- The analysis of biodiesel requires separation of typical FAMEs between myristic acid (C_{14}) and nervonic acid (C_{24}:1) methyl esters. This analysis is possible on OPTIMA® BioDiesel F in only 22 min. Additionally, linolenic acid methyl ester can be determined due to the good resolution. The extended standard DIN EN 14103:2011 also covers smaller FAMEs starting from C_6 (see application 214510 on opposite page). Change of the internal standard from C_{17} to C_{19} also allows the analysis of animal fats.

Temperature

- T_{max} 240 °C (long-term temperature), T_{max} 250 °C (short-term max. temperature in a temperature program)

Similar phases

- Select™ Biodiesel for FAME, Trace TR-BioDiesel (F)

OPTIMA® BioDiesel G for analysis of glycerol and glycerides in accordance with DIN EN 14105

Key features

- The capillary column OPTIMA® BioDiesel G allows determination of free glycerol and residues of mono-, di- and triglycerides in FAMEs intended as additives for mineral oils. The procedure can be applied for FAMEs from rapeseed oil, sunflower oil and soy bean oil. Glycerol as well as mono- and diglycerides are derivatized to more volatile substances by addition of MSTFA in the presence of pyridine (see page 363).

Temperature

- T_{max} 380 °C (long-term temperature), T_{max} 400 °C (short-term max. temperature in a temperature program)

Similar phases

- Select™ Biodiesel for Glycerides, Trace TR-BioDiesel (G), MET-Biodiesel

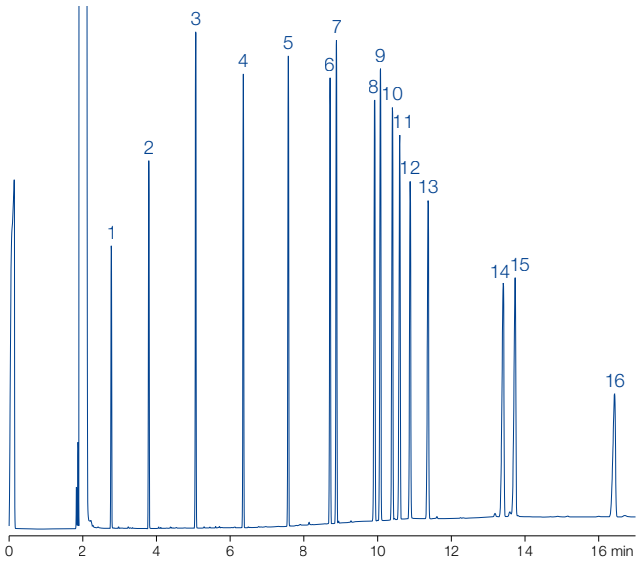
Analysis of FAMEs from biodiesel in accordance with DIN EN 14103:2011

MN Appl. No. 214510

Column:	OPTIMA® BioDiesel F, 30 m x 0.25 mm ID
Sample:	50 µg/mL each in dichloromethane
Injection:	10 µL, 250 °C, split 1:20
Carrier gas:	1.2 bar He
Temperature:	80 °C → 250 °C (8.5 min), 20 °C/min
Detector:	FID 260 °C

Peaks:

1. C6:0
2. C8:0
3. C10:0
4. C12:0
5. C14:0
6. C16:0
7. C16:1
8. C18:0
9. C18:1
10. C18:2
11. C19:0, int. st.
12. C18:3
13. C20:0
14. C22:0
15. C22:1
16. C24:0



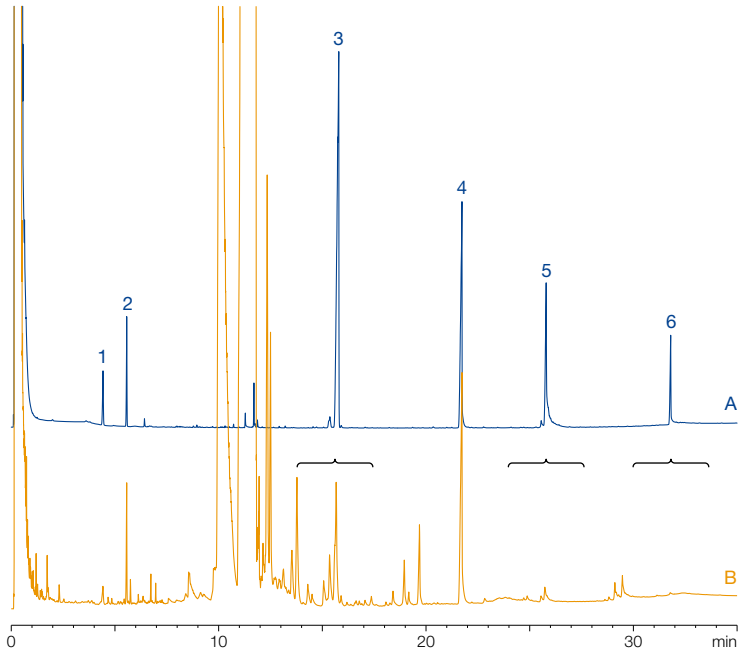
Analysis of glycerol and glycerides from biodiesel

MN Appl. No. 213640

Column:	OPTIMA® BioDiesel G, 10 m x 0.25 mm ID
Sample:	A) standard in *n*-heptane B) biodiesel
Injection:	2 µL, 350 °C, CIS (15 °C → 350 °C, 12 °C/s)
Carrier gas:	0.8 bar H_2, split 1: 2.6
Temperature:	50 °C (3.5 min) → 180 °C, 15 °C/min → 280 °C, 7 °C/min → 370 °C (10 min), 10 °C/min
Detector:	FID 380 °C

Peaks:

1. Glycerol (TMS)
2. Butanetriol (TMS), IS
3. Monoolein = glycerol monooleate (TMS)
   + monoacylglycerides
4. Tricaprin (glycerol tricaprate), IS
5. Diolein = glycerol dioleate (TMS)
   + diacylglycerides
6. Triolein = glycerol trioleate
   + triacylglycerides



Ordering information

OPTIMA® BioDiesel

	Length → 10 m	30 m
OPTIMA® BioDiesel M		
0.32 mm ID (0.5 mm OD)		726905.30
OPTIMA® BioDiesel F		
0.25 mm ID (0.4 mm OD)		726900.30
OPTIMA® BioDiesel G		
0.25 mm ID (0.4 mm OD)	726903.10	

Capillary columns for triglyceride analysis

OPTIMA® 1-TG · 17-TG for triglyceride analysis · USP G1 / G2 / G38 (1-TG) · USP G3 (17-TG)

Key features

- Short capillary columns (max. 25 m and 0.32 mm ID) with low-bleeding stationary phases thermally stable with optimized deactivation

Recommended application

- OPTIMA® 1-TG
  100 % dimethylpolysiloxane offers separation according to carbon number
- OPTIMA® 17-TG
  phenyl-methyl-polysiloxane (50 % phenyl) for separation according to degree of unsaturation

Temperature

- T_{max} 370 °C (both phases)

Similar phases der OPTIMA® 1-TG:

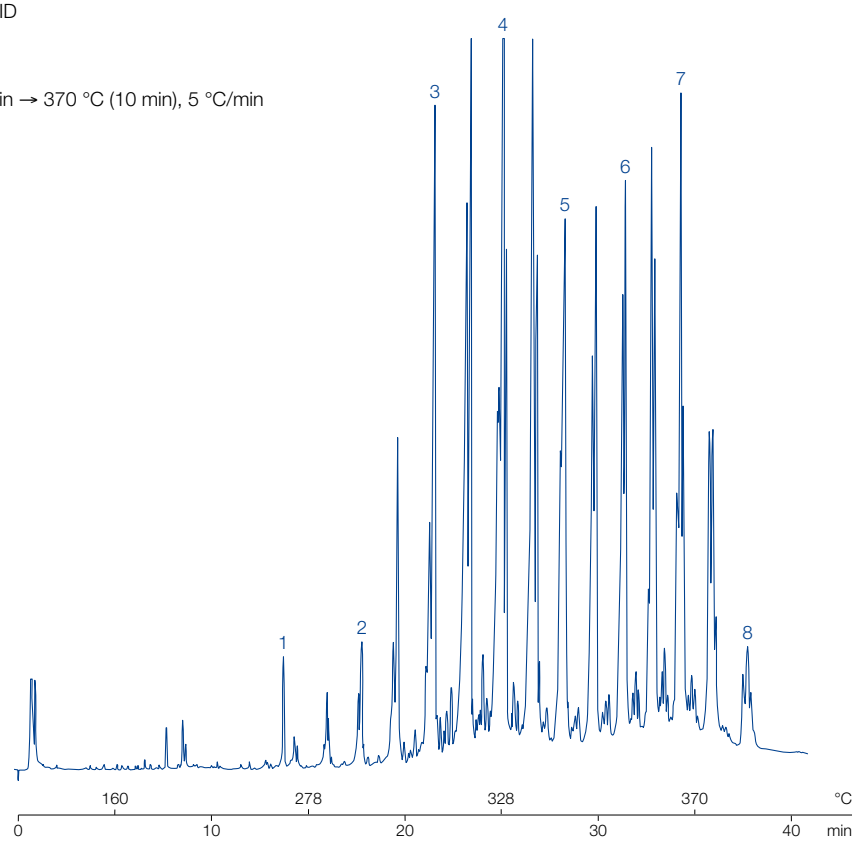
- SPB-1 TG, DB-1 HT, 400-1 HT, HT-5

Triglycerides (from butter)

MN Appl. No. 201790

Column: OPTIMA® 1-TG, 25 m x 0.32 mm ID
Injection: 0.5 µL
Carrier gas: 80 kPa H_2
Temperature: 80 °C (1 min) → 250 °C, 20 °C/min → 370 °C (10 min), 5 °C/min
Detector: FID 380 °C

Peaks:
1. Cholesterol
2. T-30
3. T-34
4. T-38
5. T-42
6. T-46
7. T-50
8. T-54



Ordering information

OPTIMA® 1-TG · OPTIMA® 17-TG

	Length → 10 m	25 m
OPTIMA® 1-TG		
0.25 mm ID (0.4 mm OD)	726133.10	726133.25
0.32 mm ID (0.5 mm OD)	726132.10	726132.25
OPTIMA® 17-TG		
0.32 mm ID (0.5 mm OD)	726131.10	726131.25

OPTIMA® 5 HT for high temperature GC · USP G27 / G36

Key features
- Chemically bonded, cross-linked silarylene phase with polarity similar to a 5 % diphenyl - 95 % dimethylpolysiloxane phase
- Nonpolar phase, low bleeding

Recommended application
- Ideal for MS detectors, can be rinsed with solvents
- For simulated distillation, hydrocarbon, fuel and oil analysis, high-boiling analytes

Temperature
- T_{max} 380 °C (long-term temperature), T_{max} 400 °C (short-term max. temperature in a temperature program)

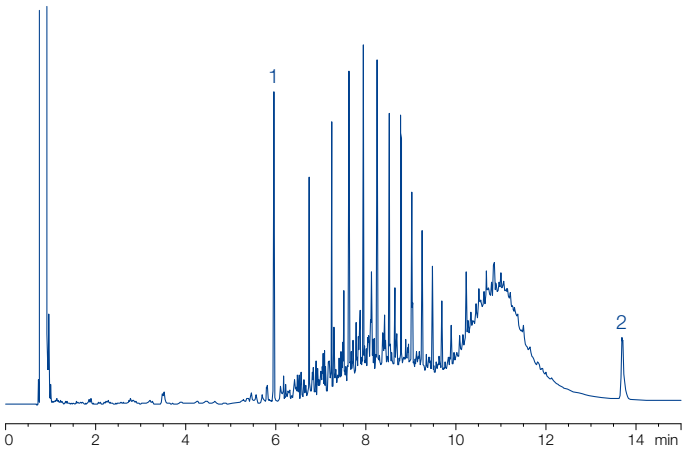
Similar phases
- DB-5HT, VF-5HT, HT-5, XTI-5HT, ZB-5HT

Separation of motor oil / mineral oil (type A + B),
rapid determination in accordance with DIN H-53 / ISO DIS

MN Appl. No. 213400

Column:	OPTIMA® 5 HT, 15 m x 0.32 mm ID, 0.25 µm film
Sample:	mineral oil type A + B (hydrocarbon index kit acc. to EN ISO 9377-2) in hexane
Injection:	1 µL, splitless, 300 °C
Carrier gas:	0.6 bar He
Temperature:	40 °C (5 min) → 390 °C, 50 °C/min
Detector:	FID 280 °C

Peaks:
1. Decane (C10)
2. Tetracontane (C40)



Ordering information

OPTIMA® 5 HT

	Length → 15 m	30 m
0.25 mm ID (0.4 mm OD)		
0.10 µm film	726102.15	726102.30
0.25 µm film	726106.15	726106.30
0.32 mm ID (0.5 mm OD)		
0.10 µm film	726104.15	726104.30
0.25 µm film	726108.15	726108.30

Further applications can be found online in our application database at *www.mn-net.com/apps*

Capillary columns for amine separation

OPTIMA® 5 Amine special column for analysis of amines · USP G27 / G36

Key features

- Nonpolar phase
- Improved linearity for analysis of active components at trace levels: no amine absorptions even for aliphatic and aromatic amines at concentrations of 100 pg/peak
- Tested with the OPTIMA® Amine test mixture (REF 722317), which contains, amongst others, diethanolamine and propanol-pyridine (this test mixture is supplied with each column)

Recommended application

- Especially deactivated for the analysis of polyfunctional amines such as ethanolamines, amino-functionalized diols and similar compounds, which are important base materials in industrial chemistry, and show strong tailing on standard-deactivated columns

Temperature

- T_{max} 300 °C (long-term temperature), T_{max} 320 °C (short-term max. temperature in a temperature program)

Similar phases

- Rtx®-5 Amine, PTA-5

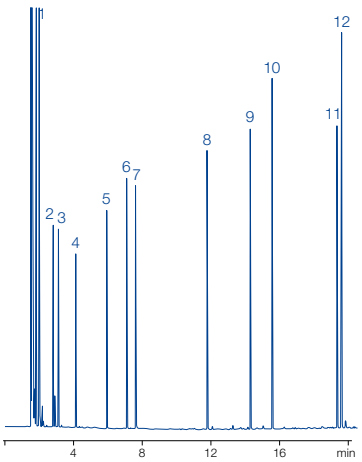
Separation of secondary and tertiary amines

MN Appl. No. 210280

Column:	OPTIMA® 5 Amine, 30 m x 0.25 mm ID, 1.0 µm film
Injection:	1 µL, split 1:100
Carrier gas:	0.6 bar H_2
Temperature:	100 °C (3 min) → 280 °C, 10 °C/min
Detector:	FID 280 °C

Peaks:

1. Diethylamine
2. Di-isopropylamine
3. Triethylamine
4. Di-*n*-propylamine
5. Di-*n*-butylamine
6. Tri-*n*-propylamine
7. Di-isobutylamine
8. Tri-*n*-butylamine
9. Di-isohexylamine
10. Dicyclohexylamine
11. Dibenzylamine
12. Tri-*n*-hexylamine



Ordering information

OPTIMA® 5 Amine

	Length → 10 m	25 m	30 m
0.1 mm ID (0.4 mm OD)			
0.40 µm film	726361.10		
0.2 mm ID (0.4 mm OD)			
0.35 µm film		726355.25	
0.25 mm ID (0.4 mm OD)			
0.50 µm film			726354.30
1.00 µm film			726358.30
0.32 mm ID (0.5 mm OD)			
0.25 µm film			726360.30
1.00 µm film			726353.30
1.50 µm film			726356.30
0.53 mm ID (0.8 mm OD)			
1.00 µm film			726359.30
3.00 µm film			726357.30

FS-CW 20 M-AM polyethylene glycol 20 000, non-immobilized · USP G16

Key features

- Polyethylene glycol, basic for amine separations

Temperature

- T_{max} 220 °C (long-term temperature), T_{max} 240 °C (short-term max. temperature in a temperature program)

Similar phases

- Carbowax™ Amine, CP-Wax 51, CAM, Stabilwax® DB

Ordering information

FS-CW 20 M-AM

	Length → 10 m	25 m	50 m
0.1 mm ID (0.4 mm OD)			
0.25 µm film	733111.10		
0.25 mm ID (0.4 mm OD)			
0.25 µm film		733110.25	733110.50
0.32 mm ID (0.5 mm OD)			
0.25 µm film		733299.25	733299.50
0.35 µm film			733442.50
0.53 mm ID (0.8 mm OD)			
1.00 µm film		733551.25	

Further applications can be found online in our application database at *www.mn-net.com/apps*

Capillary columns for hydrocarbons

PERMABOND® P-100 for analysis of petrochemical products · USP G1 / G2 / G38

Key features
- Extra long column with nonpolar dimethylpolysiloxane phase

Recommended application
- High resolution and sufficient capacity for analysis of complex mixtures of hydrocarbons

Temperature
- T_{max} 300 °C (long-term temperature), T_{max} 320 °C (short-term max. temperature in a temperature program)

Ordering information

PERMABOND® P-100

	Length → 100 m
0.25 mm ID (0.4 mm OD)	
0.50 µm film	723890.100

PERMABOND® SE-54-HKW for volatile halogenated hydrocarbons · USP G36

Recommended application
- SE-54 optimized for volatile halogenated hydrocarbons

Temperature
- T_{max} 300 °C (long-term temperature), T_{max} 320 °C (short-term max. temperature in a temperature program)

For the analysis of halogenated hydrocarbons, we recommend our optimized column PERMABOND® SE-54-HKW at 25 or 50 m length with our approved polysiloxane phase SE-54.

As an alternative, or to verify analytical results, the OPTIMA® 624 has proven itself as advantageous, especially for the determination of 1,1,2-trichlorotrifluoroethane (F 113) along with dichloromethane.

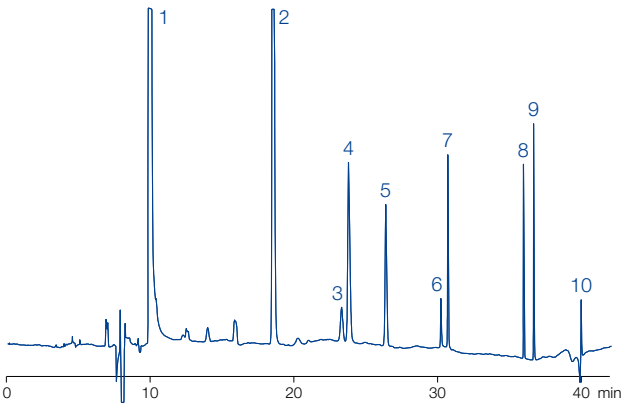
Both phases are also suited for the determination of vinyl chloride as well as for the separation of cis/trans isomers of 1,2-dichloroethene. The high film thickness secures a high capacity and an outstanding resolution. For GC/MS coupling, we recommend OPTIMA® 624 LB or OPTIMA® 624 with 0.2 or 0.25 mm ID

Volatile halogenated hydrocarbons

MN Appl. No. 212480

Column: PERMABOND® SE-54-HKW, 50 m x 0.32 mm ID
Injection: 1 µL, split ~ 1:30
Carrier gas: 0.9 bar He
Temperature: 35 °C (25 min) → 160 °C (5 min), 10 °C/min
Detector: ECD 300 °C

Peaks:
1. Dichloromethane (795 ng/mL)
2. Trichloromethane (75 ng/mL)
3. 1,1,1-Trichloroethane (67 ng/mL)
4. 1,2-Dichloroethane (100 ng/mL)
5. Tetrachloromethane (15.9 ng/mL)
6. Trichloroethene (14.6 ng/mL)
7. Bromodichloromethane (20 ng/mL)
8. Dibromochloromethane (122 ng/mL)
9. Tetrachloroethene (81 ng/mL)
10. Tribromomethane (28.9 ng/mL)



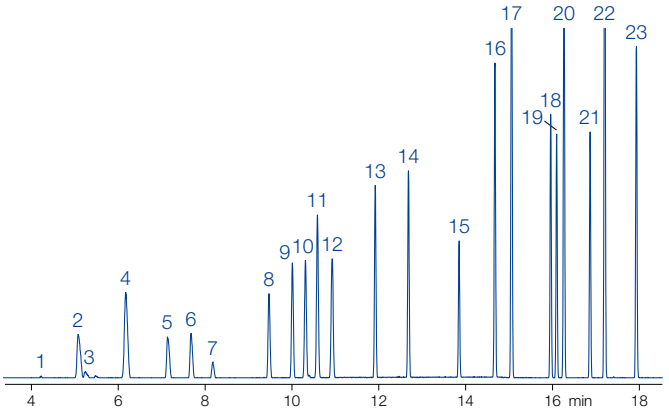
Volatile halogenated hydrocarbons and BTX

MN Appl. No. 200160

Column:	OPTIMA® 624, 50 m x 0.25 mm ID, 1.40 µm film
Injection:	1 µL, split 50 mL/min
Carrier gas:	0.9 mL/min He (constant flow)
Temperature:	40 °C (5 min) → 160 °C, 10 °C/min
Detector:	MSD 5971

Peaks:

1. Vinyl chloride
2. Trichlorofluoromethane (F 11)
3. Pentane
4. 1,1,2-Trichlorotrifluoroethane (F 113)
5. Dichloromethane
6. *trans*-1,2-Dichloroethene
7. Hexane
8. *cis*-1,2-Dichloroethene
9. Trichloromethane
10. 1,1,1-Trichloroethane
11. Tetrachloromethane
12. 1,2-Dichloroethane + benzene
13. Trichloroethene
14. Bromodichloromethane
15. Toluene
16. Tetrachloroethene
17. Dibromochloromethane
18. Chlorobenzene
19. Ethylbenzene
20. *m*- + *p*-Xylene
21. *o*-Xylene
22. Tribromomethane
23. Bromobenzene



Ordering information

PERMABOND® SE-54-HKW

	Length → 25 m	50 m
0.32 mm ID (0.5 mm OD)		
1.80 µm film	723945.25	723945.50

Further applications can be found online in our application database at *www.mn-net.com/apps*

Capillary columns for silane · DEG

PERMABOND® Silane for silane analysis

Recommended application

- Developed especially for the analysis of monomeric silanes and chlorosilanes (not for the separation of trimethylsilyl derivatives)
- Also suited for the separation of dimeric siloxanes and silazanes

Temperature

- 0.32 mm ID: T_{max} 260 °C (long-term temperature), T_{max} 280 °C (short-term max. temperature in a temperature program)
- 0.53 mm ID: T_{max} 240 and 260 °C, resp.

Ordering information

PERMABOND® Silane

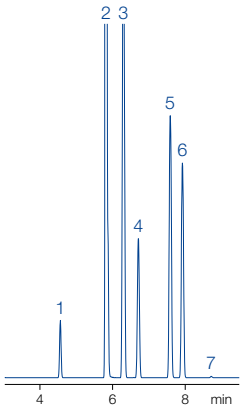
	Length → 25 m	50 m
0.32 mm ID (0.5 mm OD)		723409.50
0.53 mm ID (0.8 mm OD)	723411.25	

Chloromethylsilanes

MN Appl. No. 200090

Column: PERMABOND® Silane, 50 m x 0.32 mm ID
Injection: 0.5 µL gas, split 80 mL/min
Carrier gas: 1 mL/min He (constant flow)
Temperature: 50 °C → 100 °C, 5 °C/min
Detector: MSD 5971

Peaks:
1. Tetramethylsilane
2. Dichloromethane
3. Tetrachlorosilane
4. Chlorotrimethylsilane
5. Methyltrichlorosilane
6. Dichlorodimethylsilane
7. Hexamethyldisiloxane

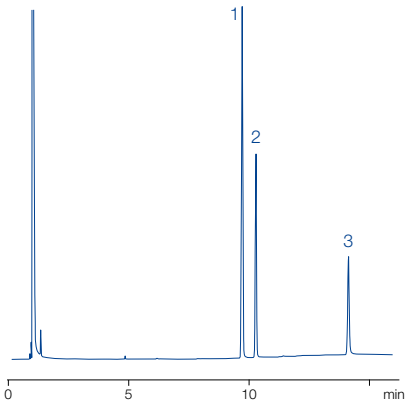


Diethylene glycol standard in wine

MN Appl. No. 201500

Column: PERMABOND® CW 20 M-DEG, 25 m x 0.25 mm ID
Injection: 0.5 µL, split ~1:40
Carrier gas: 1.2 bar N_2
Temperature: 80 °C → 200 °C, 10 °C/min
Detector: FID 260 °C

Peaks:
DEG standard
1. 1,4-Butanediol
2. Diethylene glycol
3. Glycerol



PERMABOND® CW 20 M-DEG for determination of diethylene glycol · USP G16

Key features

- Polyethylene glycol 20 000 (diethylene glycol tested)

Recommended application

- Determination of diethylene glycol (DEG), e.g., for the quality control of wine

Temperature

- T_{max} 220 °C (long-term temperature), T_{max} 240 °C (short-term max. temperature in a temperature program)

Ordering information

PERMABOND® CW 20 M-DEG

	Length → 25 m
0.25 mm ID (0.4 mm OD)	
0.25 µm film	723063.25
0.32 mm ID (0.5 mm OD)	
0.25 µm film	723327.25

Further applications can be found online in our application database at *www.mn-net.com/apps*

Untreated capillaries

Recommended application

- Capillary electrophoresis
- Preparation of capillary columns
- Capillary LC applications

Ordering information

Untreated capillaries

	Length → 1 m Pack of 3	10 m Pack of 1	25 m Pack of 1
Capillaries for electrophoresis			
0.025 mm ID (0.4 mm OD)	723793.1	723793.2	
0.05 mm ID (0.4 mm OD)	723790.1	723790.2	
0.075 mm ID (0.4 mm OD)	723791.1	723791.2	
0.10 mm ID (0.4 mm OD)	723792.1	723792.2	
Untreated capillaries			
0.20 mm ID (0.4 mm OD)		723148.10	723148.25
0.25 mm ID (0.4 mm OD)		723101.10	723101.25
0.32 mm ID (0.5 mm OD)		723151.10	723151.25
0.53 mm ID (0.8 mm OD)		723501.10	723501.25
Untreated capillaries are supplied without cage.			

Deactivated capillary columns precolumns / guard columns

Recommended application

- As precolumns / guard columns, whenever a larger contamination capacity is required
- Preparation of capillary columns

Ordering information

Deactivated capillary columns

	Length → 10 m	25 m
Methyl-Sil deactivated (T_{max} 320 °C)		
0.25 mm ID (0.4 mm OD)	723106.10	723106.25
0.32 mm ID (0.5 mm OD)	723346.10	723346.25
0.53 mm ID (0.8 mm OD)	723558.10	723558.25
Phenyl-Sil deactivated (T_{max} 320 °C)		
0.25 mm ID (0.4 mm OD)	723108.10	723108.25
0.32 mm ID (0.5 mm OD)	723348.10	723348.25
0.53 mm ID (0.8 mm OD)	723560.10	723560.25
CW deactivated (T_{max} 250 °C)		
0.25 mm ID (0.4 mm OD)	723105.10	723105.25
0.32 mm ID (0.5 mm OD)	723349.10	723349.25
0.53 mm ID (0.8 mm OD)	723562.10	723562.25
Untreated capillaries are supplied without cage.		

For a considerably longer lifetime, even for contaminated or matrix-containing samples, MN offers the option of integrated precolumns. All capillary columns are available with a 10 m guard column with matched deactivation. For ordering, please add V1 at the end of the REF number. Guard column combinations with other lengths, IDs or different deactivation are available on request.

Retention gaps

Key features

- The retention gap technique in combination with on-column injection allows to concentrate a large sample volume in the capillary column.
- Choice of the retention gap depends on the solvent used: the flooded zone after injection should be between 20–30 cm/µL
- Me-Sil retention gap: only for use with *n*-hexane and diethyl ether
- Phe-Sil retention gap: for all solvents except methanol and water
- CW retention gap: for all solvents and especially for methanol and water

Temperature

- T_{max} 250 °C (CW retention gaps), T_{max} 320 °C (Me-Sil and Phe-Sil retention gaps)

Note:

- Calculation example: length of flooded zone ~ 20–30 cm/µL, retention gap 10 m x 0.32 mm ID, capillary column: 25 m x 0.32 mm ID, max. injection volume ~ 30–50 µL
- A retention gap must be inert without any noticeable retention: Me-Sil retention gaps are more inert than Phe-Sil, while Phe-Sil is less susceptible to contamination
- Retention gaps can also be used as transfer lines or precolumns (contamination capacity about 5–10 µg).

Ordering information

Retention gaps

	Length →	
	10 m	25 m
Me-Sil retention gaps (T_{max} 320 °C)		
0.25 mm ID (0.4 mm OD)	723706.10	723706.25
0.32 mm ID (0.5 mm OD)	723707.10	723707.25
0.53 mm ID (0.8 mm OD)	723708.10	723708.25
Phe-Sil retention gaps (T_{max} 320 °C)		
0.25 mm ID (0.4 mm OD)	723709.10	723709.25
0.32 mm ID (0.5 mm OD)	723710.10	723710.25
0.53 mm ID (0.8 mm OD)	723711.10	723711.25
CW retention gaps (T_{max} 250 °C)		
0.25 mm ID (0.4 mm OD)	723712.10	723712.25
0.32 mm ID (0.5 mm OD)	723713.10	723713.25
0.53 mm ID (0.8 mm OD)	723714.10	723714.25
Retention gaps are supplied without cage.		

For a considerably longer lifetime, even for contaminated or matrix-containing samples, MN offers the option of integrated precolumns. All capillary columns are available with a 10 m guard column with matched deactivation. For ordering, please add V1 at the end of the REF number. Guard column combinations with other lengths, IDs or different deactivation are available on request.

Derivatization reagents

Key features

- Derivatization reagents:
  To improve volatility, increase thermal stability or to achieve a lower limit of detection in gas chromatography
- Prerequisite: quantitative, rapid and reproducible formation of only one derivative
- Halogen atoms inserted by derivatization, e.g., trifluoro-acetates, allow the specific detection in an ECD with the advantage of high sensitivity.
- Specific derivatizations may influence elution orders and fragmentation patterns in a MS
- We provide reagents for
  - acylation
  - alkylation (methylation)
  - silylation
- For 1 x 10 mL, 1 x 50 mL and 6 x 50 mL also available with screw closure

Ordering information

Derivatization method development kits*

Designation	Contents of the kit	REF
Which type of derivatization is suited best for your sample (alkylation, acylation or silylation)?	2 x 1 mL each of TMSH, MSTFA, MBTFA	701952
Acylation kit		
Which is the proper reagent for acylation?	2 x 1 mL each of MBTFA, TFAA, MBHFBA	701950
Alkylation kit		
Which is the proper reagent for methylation?	3 x 1 mL each of TMSH, DMF-DMA	701951
Silylation kit		
Which is the proper reagent for silylation?	2 x 1 mL each of MSTFA, BSTFA, TSIM, MSHFBA	701953

* These products contain harmful substances which must be specially labeled as hazardous. For detailed information please see SDS.

Selection guide for derivatization of important functional groups in GC

Function	Method	Derivative	Recommended reagents
alcohols, phenols	silylation	R'O – TMS	BSA, MSTFA, MSHFBA, TSIM, SILYL-2110, SILYL-21, SILYL-1139
R'OH	acylation	R'O – CO – R	TFAA, HFBA, MBTFA, MBHFBA
	alkylation	R'O – R	TMSH
sterically hindered	silylation	R'O – TMS	TSIM, BSTFA, SILYL-991
amines	silylation	R' – NR'' – TMS	BSA, MSTFA, MSHFBA, SILYL-991
primary, secondary	acylation	R' – NR'' – CO – R	TFAA, HFBA, MBTFA, MBHFBA
hydrochlorides	silylation	R' – NR'' – TMS	MSTFA
amides	silylation	not stable	
	acylation	R' – CO – NH – CO – R	TFAA, MBTFA, HFBA, MBHFBA
amino acids	silylation	R' – CH(NH – TMS) – CO – O – TMS	BSA, BSTFA, MSTFA, MSHFBA
	alkylation (a)	R' – CH(NH – CO-R) – CO – O – R	a) MeOH/TMCS, TMSH
	+ acylation (b)		b) TFAA, HFBA, MBTFA, MBHFBA
Carboxylic acids (fatty acids)	silylation	R' – CO – O – TMS susceptible to hydrolysis	BSA, MSTFA, MSHFBA, TMCS, TSIM, SILYL-2110, SILYL-21, Silyl-1139
	alkylation	R' – CO – O – R	DMF-DMA, MeOH/TMCS (1 M), TMSH
salts	silylation	R' – CO – O – TMS susceptible to hydrolysis	TMCS
carbohydrates	silylation		MSTFA, TSIM, HMDS, SILYL-1139
	acylation		TFAA, MBTFA
steroids	silylation		BSA, TSIM
	acylation		TFAA, MBTFA, HFBA, MBHFBA

These products contain harmful substances which must be specially labeled as hazardous. For detailed information please see SDS.

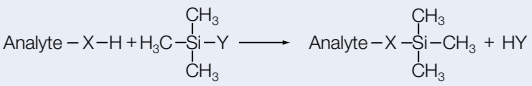
Due to their purpose, derivatization reagents are very reactive substances. For this reason, they should be stored cool and protected from moisture. For easy access with a syringe, our derivatization reagents are supplied in vials with crimp caps (exception DMCS and TMCS with screw closure). Vials with pierced sealing disks have limited stability and should be used soon.

The derivatization procedures can be found on page 367.

Reagents / methods for derivatization

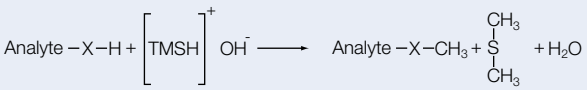
General reaction mechanisms

Silylation



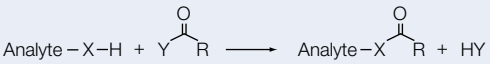
X = e.g., O, S, COO, etc.
Y = rest of silylation reagents

Alkylation (Methylation) · example TMSH



X = e.g., O, S, COO, etc.

Acylation



X = e.g., O, S, NH, etc.
Y = rest of acylation reagents

MACHEREY-NAGEL derivatization reagents for GC

Content of brochure

- Product range for acylation, alkylation and silylation reagents
- Protocols for derivatization
- Diverse tips and hints

Ordner now your derivatization brochure KATEN200144

Acylation reagents

Acyl halides

Key features

- By-product of acylation with acyl halides: corresponding hydrohalic acids excess of reagent and acid have to be removed or trapped by a suitable base (e.g., pyridine)

- Pentafluorobenzoyl chloride
  PFBC: $C_6F_5-CO-Cl$
  M 230.52 g/mol, Bp 158–159 °C (760 mm Hg),
  Density d20°/4° = 1.601

Anhydrides

Key features

- By-products of acylation with anhydrides: corresponding acids excess reagent and the acid formed are to be removed
- Trifluoroacetic acid anhydride TFAA: $CF_3-CO-O-CO-CF_3$
  M 210.04 g/mol, Bp 39.5–40.5 °C (760 mm Hg),
  Density d20°/4° = 1.490

- Heptafluorobutyric acid anhydride
  HFBA: $C_3F_7-CO-O-CO-C_3F_7$
  M 410.06 g/mol, Bp 106–107 °C (760 mm Hg),
  Density d20°/4° = 1.665

Bisacylamides

Key features

- By-products: corresponding neutral acylamides: high volatility
- Easily removed; due to the neutral conditions and their favorable chromatographic characteristics, the removal of surplus bisacylamides and their by-products is often not necessary. Therefore, the sample preparation is much easier.

- *N*-methyl-bis(trifluoroacetamide)
  MBTFA: $CF_3-CO-N(CH_3)-CO-CF_3$
  M 223.08 g/mol, Kp 123–124 °C (760 mm Hg),
  Density d20°/4° = 1.55
- *N*-methyl-bis(heptafluorobutyramide)
  MBHFBA: $C_3F_7-CO-N(CH_3)-CO-C_3F_7$
  M 423.1 g/mol, Kp 165–166 °C (760 mm Hg),
  Density d20°/4° = 1.673

Methods for acylation

Acylation with fluorinated acid anhydrides (TFAA, HFBA)

- Applicable for alcohols, phenols, carboxylic acids, amines, amino acids and steroids, stable derivatives for FID or ECD detection
- Procedure see page 367 or online at *www.mn-net.com/apps*
  TFAA: MN Appl. Nr. 213041
  HFBA: MN Appl. Nr. 213042

Acylation with fluorinated acid amides (MBTFA, MBHFBA)

- Recommended for alcohols, primary and secondary amines as well as for thiols under mild, neutral conditions
- MBTFA also forms very volatile derivatives with carbohydrates [17].
- Procedure see page 367 or online at *www.mn-net.com/apps*
  MBTFA: MN Appl. Nr. 213051
  MBHFBA: MN Appl. Nr. 21305

Ordering information

Acylation reagents*

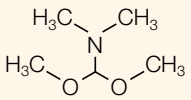
Substance	Packing unit			
	10 x 1 mL	20 x 1 mL	1 x 10 mL	5 x 10 mL
HFBA				
		701110.201	701110.110	701110.510
MBTFA				
		701410.201	701410.110	701410.510
MBHFBA				
	701420.101	701420.201		
PFBC				
	701120.101			
TFAA				
			701130.110	701130.510

* These products contain harmful substances which must be specially labeled as hazardous. For detailed information please see SDS.

On request for 1 x 10 mL, 1 x 50 mL and 6 x 50 mL also available with screw closure.

Alkylation / methylation reagents

DMF-DMA *N*,*N*-dimethylformamide dimethylacetal

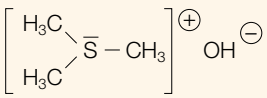


- M 119.17 g/mol,
  Kp 106–107 °C (760 mm Hg),
  Density d20°/4° = 0.897

Key features

- Methylation reagents

TMSH (0.2 mol/L in methanol) Trimethylsulfonium hydroxide



- M 94.06 g/mol

Key features

- Methylation reagents

Methods for alkylation / methylation

Methylation with TMSH

- Suited for free acids, chlorophenoxycarboxylic acids, their salts and derivatives as well as for phenols and chlorophenols [18]
- The great advantage is the simplification of the sample preparation. Lipids or triglycerides can be converted to the corresponding fatty acid methyl esters (FAMEs) by simple transesterification.
- This reaction is very elegant and convenient, because it is only necessary to add the reagent (0.2 mol/L in methanol) to the sample solution. Removal of surplus reagent is not required, since at 250 °C inside the injector of the gas chromatograph, TMSH will pyrolyze solely to volatile methanol and dimethylsulfide. Due to high reactivity, a complete conversion is usually obtained at ambient temperature. Heating (e.g., 10 min at 100 °C) in a closed sample vial may be necessary, however.
- Procedure see page 367 or online at *www.mn-net.com/apps*
  MN Appl. Nr. 213060

Methylation with DMF-DMA

- Applicable for fatty acids, primary amines and (partially) amino acids, under formation of *N*-dimethyl-aminomethylene amino acid methyl esters [19]
- Since DMF-DMA is a poor solvent, it is essential to use a mixture of DMF-DMA with pyridine, THF, acetone (barbiturates) or another solvent.
- Procedure see page 367 or online at *www.mn-net.com/apps*
  MN Appl. Nr. 213070

Methylation with methanol – TMCS (1 M)

- Suited for the esterification of free carboxylic acids and the transesterification of glycerides. Formation of HCl catalyzes the reaction. TMCS, resp. silyl ethers remove the water and thus drive the reaction to completion. The mixture should be freshly prepared.
- Procedure see page 367 or online at *www.mn-net.com/apps*
  MN Appl. Nr. 213080

For GC separation of FAMEs from natural butter fat after derivatization with TMSH see Appl. 201680 at *www.mn-net.com/apps*

Ordering information

Alkylation reagents*

Substance	Packing unit 10 x 1 mL	20 x 1 mL	1 x 10 mL	5 x 10 mL
DMF-DMA				
		701430.201	701430.110	
TMSH				
	701520.101	701520.201	701520.110	701520.510

* These products contain harmful substances which must be specially labeled as hazardous. For detailed information please see SDS.

On request for 1 x 10 mL, 1 x 50 mL and 6 x 50 mL also available with screw closure.

Reagents / methods for silylation

Silylation reagents

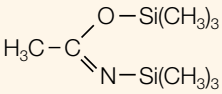
The most common form of silylation in GC is the replacing of active hydrogen atoms with a trimethylsilyl group (TMS derivative). Less frequently, trialkylsilyl groups or dimethylsilyl groups with longer alkyl chains are also in use. The alkylsilyl group increases volatility and enhances thermal stability of the sample.

Silylation can be catalyzed either acidic by addition of TMCS or basic by addition of pyridine or TSIM (e.g., for sterically hindered functionalities like tert. alcohols).

Reactivity of silylation reagents (acc. to M. Donike): TMS amide (e.g., BSA, MSTFA) > TMS amine = TSIM > Enol-O-TMS ether > S-TMS ether > O-TMS ether > TMS-O-TMS

Stability of the TMS derivatives: O-TMS ether > S-TMS ether > Enol-O-TMS ether > TMS amine > TMS amide

BSA *N,O*-bis-trimethylsilyl-acetamide



- M 203.4 g/mol,
  Bp 71–73 °C (35 mm Hg),
  Density d20°/4° = 0.832

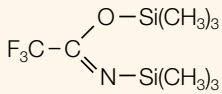
Key features

- Strong silylation reagent
- Not recommended for use with carbohydrates or very low molecular weight compounds
- Good solvent for polar compounds, but frequently used in combination with a solvent (pyridine, DMF etc.) or with other silylation reagents. Dissolved in DMF, BSA is the prime derivatization reagent for phenols.

Recommended application

- Alcohols, amines, carboxylic acids, phenols, steroids, biogenic amines and alkaloids are derivatized to stable TMS derivatives

BSTFA *N,O*-bis-trimethylsilyl-trifluoroacetamide



- M 257.4 g/mol,
  Bp 40 °C (12 mm Hg),
  Density d20°/4° = 0.961

Key features

- Powerful trimethylsilyl donor with approx. the same donor strength as the nonfluorinated analog BSA
- Advantage of BSTFA over BSA: greater volatility of its reaction products, particularly useful for GC analysis of low boiling TMS amino acids
- BSTFA is nonpolar (less polar than MSTFA) and can be mixed with acetonitrile for improved solubility. For the silylation of fatty acid amides, hindered hydroxyl groups and other difficult to silylize compounds, e.g., secondary alcohols and amines, we recommend BSTFA + 1 % trimethylchlorosilane (TMCS), available under the designation SILYL-991 (see page 366).

Silylation with BSA, BSTFA or SILYL-991 (BSTFA + 1 % TMCS)

- Procedure see page 367 or online at *www.mn-net.com/apps*

BSA	MN Appl. Nr. 213091
BSTFA	MN Appl. Nr. 213092
SILYL-991	MN Appl. Nr. 213093

Silylation with BSA in combination with other silylation reagents

- Procedure see page 367 or online at *www.mn-net.com/apps*
  MN Appl. Nr. 213100

Ordering information

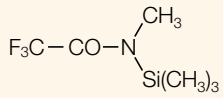
Silylation reagents*

Substance	Packing unit 20 x 1 mL	1 x 10 mL	5 x 10 mL	1 x 50 mL	1 x 100 mL
BSA					
		701210.110	701210.510	701210.150	
BSTFA					
	701220.201	701220.110	701220.510		
SILYL-991 –(BSTFA – TMCS (99:1))					
	701490.201			701490.150	701490.1100

* These products contain harmful substances which must be specially labeled as hazardous. For detailed information please see SDS.

On request for 1 x 10 mL, 1 x 50 mL and 6 x 50 mL also available with screw closure.

MSTFA *N*-methyl-*N*-trimethylsilyl-trifluoroacetamide



- M 199.1 g/mol, Bp 70 °C (75 mm Hg), Density d20°/4° = 1.11

Key features

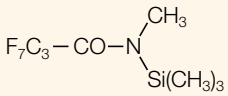
- The most volatile trimethylsilyl amide available, very strong TMS donor which does not cause noticeable FID fouling even during long-time measuring series

Recommended application

- Carboxylic acids, hydroxy and ketocarboxylic acids, amino acids, amines, alcohols, polyalcohols, sugars, mercaptans and similar compounds with active hydrogen atoms. Even amine hydrochlorides can be silylated directly.

- The addition of protic solvents in submolar quantities, e.g., TFA for extremely polar compounds (hydrochlorides) or pyridine for carbohydrates), can improve the already good dissolving power of MSTFA.
- Advantages: complete conversion with high reaction rates, even without a catalyst (1–2 % TMCS or TSIM); the by-product of the reaction (*N*-methyltrifluoroacetamide) shows a high volatility and a short retention time

MSHFBA *N*-methyl-*N*-trimethylsilyl-heptafluorobutyramide



- M 299.1 g/mol, Bp 148 °C (760 mm Hg)

Key features

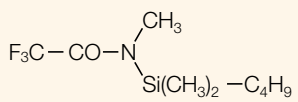
- Similar to MSTFA in reactivity and chromatography
- Either applied alone or in combination with a catalyst (TMCS, TSIM) or another silylation reagent with or without solvent; the by-product *N*-methylheptafluorobutyric amide has a lower retention time than the silylating reagent

Recommended application

- Carboxylic acids, alcohols, phenols, primary and secondary amines and amino acids

- Especially useful for flame ionization detection due to the large ratio of fluorine to silicon of 7:1, since degradation of the surplus MSHFBA does not produce SiO_2 but volatile, non-corrosive silicon compounds

MBDSTFA *N*-methyl-*N*-*tert*-butyldimethylsilyl-trifluoroacetamide



- M 241.3 g/mol, Bp 170 °C (760 mm Hg), Density d20°/4° = 1.121

Key features

- Silylation reagent that donates a *tert*-butyldimethylsilyl group (TBDMS) for derivatizing active hydrogen atoms in hydroxyl, carboxyl and thiol groups as well as primary and secondary amines
- Fast reactions (typically 5–20 min) with high yields (> 96 %), by-products are neutral volatiles
- TBDMS ethers are 10^4 times more stable than the corresponding TMS ethers
- Due to the large protecting group, chromatographic retention times are longer. This may have a beneficial impact on some separations. The high concentration of M^+-57 ions is an interesting topic for GC/MS.

Reagents / methods for silylation

Silylation with MSTFA, MSHFBA or MBDSTFA

- Procedure see page 367 or online at *www.mn-net.com/apps*
  MSTFA MN Appl. Nr. 213111 · MSHFBA MN Appl. Nr. 213112 · MBDSTFA MN Appl. Nr. 213113

Ordering information

Silylation reagents*

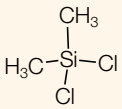
	Packing unit							
Substance	10 x 1 mL	20 x 1 mL	1 x 10 mL	5 x 10 mL	1 x 100 mL	6 x 50 mL	6 x 100 mL	12 x 100 mL
MSTFA								
		701270.201	701270.110	701270.510	701270.1100	701270.650	701270.6100	701270.12100
MSHFBA								
		701260.201	701260.110	701260.510	701260.1100		701260.6100	
MBDSTFA								
	701440.101	701440.201						

* These products contain harmful substances which must be specially labeled as hazardous. For detailed information please see SDS.

On request for 1 x 10 mL, 1 x 50 mL and 6 x 50 mL also available with screw closure.

Ultrapure derivatization reagents for acylation, alkylation and silylation.

DMCS Dimethyldichlorosilane

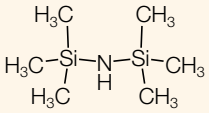


- M 129.06 g/mol,
  Bp 70 °C (760 mm Hg),
  Density d20°/4° = 1.07

Key features

- Used to form dimethylsilyl (DMS) derivatives
- DMS derivatives are much more susceptible to hydrolysis than TMS derivatives, it is therefore vital to have strictly anhydrous conditions during the conversion.

HMDS Hexamethyldisilazane

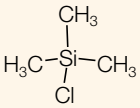


- M 161.4 g/mol,
  Bp 126 °C (760 mm Hg),
  Density d20°/4° = 0.7742

Key features

- Weak TMS donor; used as a sole reagent, it is slow and not very effective.
- Aprotic solvents like acetonitrile, pyridine, dimethylformamide, carbon disulfide and dimethylacetamide recommend themselves for use with HMDS.
- With catalytic quantities, e.g., 1 % of, or as a mixture with TMCS (2:1, v/v; SILYL-21 and SILYL-2110) it is perfectly suited for a quick and quantitative trimethylsilylation of organic compounds.

TMCS Trimethylchlorosilane

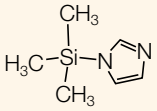


- M 108.7 g/mol,
  Bp 57 °C (760 mm Hg),
  Density d20°/4° = 0.8580

Key features

- Often used as a catalyst with other trimethylsilyl reagents
- As a sole reagent, it can be used to prepare TMS derivatives of organic acids.

TSIM *N*-trimethylsilyl-imidazole



- M 140.3 g/mol,
  Bp 94–96 °C (760 mm Hg),
  Density d20°/4° = 0.961

Key features

- Strongest hydroxyl silylator
- It is remarkable that TSIM reacts quickly and smooth with hydroxyl (even tert. OH) and carboxyl groups, but not with amines. Hence it is especially suited for multiple derivatizations, when compounds with various functional groups are to be derivatized in different ways (e.g., -O-TMS, -*N*-HFB derivatives of catecholamines).

Recommended application

- Alcohols, phenols, organic acids, steroids, hormones, glycols, nucleotides, narcotics
- Reagent of choice for carbohydrates and most steroids (even strongly hindered steroids)

Silylation with TSIM or SILYL-1139 (TSIM – pyridine 11:39)

- Procedure see page 367 or online at *www.mn-net.com/apps*

TSIM: MN Appl. Nr. 213121
SILYL-1139: MN Appl. Nr. 213122

Reagents / methods for silylation

Ordering information

Silylation reagents*

	Packing unit			
Substance	20 x 1 mL	1 x 10 mL	5 x 10 mL	6 x 50 mL
DMCS				
				701230.650
HMDS				
			701240.510	701240.650
TMCS				
	701280.201			701280.650
TSIM				
	701310.201	701310.110	701310.510	

* These products contain harmful substances which must be specially labeled as hazardous. For detailed information please see SDS.

On request for 1 x 10 mL, 1 x 50 mL and 6 x 50 mL also available with screw closure.

Ordering information

Reagent mixtures for silylation*

		Packing unit				
Mixture	Composition	20 x 1 mL	1 x 10 mL	5 x 10 mL	1 x 50 mL	1 x 100 mL
SILYL-271						
	BSA - HMDS - TSIM (2:7:1)	701450.201	701450.110	701450.510		
SILYL-1139						
	TSIM - Pyridine (11:39)	701460.201				
SILYL-21						
	HMDS - TMCS (2:1)	701470.201				
SILYL-2110						
	HMDS - TMCS - Pyridine (2:1:10)	701480.201				
SILYL-991						
	BSTFA - TMCS (99:1)	701490.201			701490.150	701490.1100

* These products contain harmful substances which must be specially labeled as hazardous. For detailed information please see SDS.

On request for 1 x 10 mL, 1 x 50 mL and 6 x 50 mL also available with screw closure.

Due to their purpose, derivatization reagents are very reactive substances. For this reason, they should be stored cool and protected from moisture. For easy access with a syringe, our derivatization reagents are supplied in vials with crimp caps (exception DMCS and TMCS with screw closure). Vials with pierced sealing disks have limited stability and should be used soon.

Silylation with SILYL-21 or SILYL-2110

- Recommended applications: sugars, glycols, sterically unhindered alcohols, carboxylic acids, acids in urine, hydroxy fatty acids, nucleotides, steroids, vitamin D, xanthone derivatives
- Procedure see page 367 or online at *www.mn-net.com/apps*
  SILYL-21 MN Appl. Nr. 213131
  SILYL-2110 MN Appl. Nr. 213132

O-trimethylsilylation with MSTFA followed by *N*-trifluoroacetylation with MBTF

- Procedure see page 367 or online at *www.mn-net.com/apps*
  MSTFA / MBTFA MN Appl. Nr. 213140

Acylation

with fluorinated acid anhydrides · TFAA MN Appl. No. 213041 · HFBA MN Appl. No. 213042

Dissolve 0.1 to 1 mg sample in 0.1 mL solvent, add 0.1 mL of the anhydride and heat to 60–70 °C for 1–2 h. If the sample needs not be concentrated prior to the analysis and if there is no danger of catalytically induced side reactions, pyridine is used as solvent. The reaction solution can be injected directly into the gas chromatograph. Otherwise, use a volatile solvent and evaporate solvent, excess reagent and free acid in a stream of nitrogen. Dissolve residue in 50 µL hexane, chloroform etc. and inject aliquot portions.

with fluorinated acid amides · MBTFA MN Appl. No. 213051 · MBHFBA MN Appl. No. 213052

Add 0.5 mL MBTFA or MBHFBA to about 2 mg sample. If there is no reaction at ambient temperature, heat the reaction mixture to 120 °C. Compounds difficult to dissolve, can be trifluoroacetylated in suitable solvent mixtures. It is recommended to use a ratio of solvent to MBTFA or MBHFBA of 4:1. The reaction mixture is chromatographed directly.

Alkylation (Methylation)

with TMSH · MN Appl. No. 213060

Dissolve 100 mg sample (e.g., butter) in 5 mL of a solvent (e.g., *tert*.-butyl methyl ether). Add 50 µL reagent to 100 µL of this solution. The mixture is injected directly. The temperature of the injector must be at least 250 °C.

with DMF-DMA · MN Appl. No. 213070

Add 1 mL of a mixture of DMF-DMA and pyridine (1:1) to 1–50 mg fatty acids. The sample can be injected as soon as a clear solution has formed. It is recommended, however, to heat the solution to 60–100 °C for 10–15 min.

with methanol – TMCS · MN Appl. No. 213080

Add 1 mL methanol – TMCS to about 50 mg carboxylic acid or glyceride and heat. Then evaporate in a stream of nitrogen and dissolve again for injection in, e.g., *n*-heptane.

Silylation

with BSA, BSTFA oder SILYL-991 (BSTFA + 1 % TMCS)
BSA MN Appl. No. 213091 · BSTFA MN Appl. No. 213092 SILYL-991 MN Appl. No. 213093

Add 0.5 mL of the silylation reagent to 1–10 mg sample; if necessary, add some solvent (normally pyridine or DMF [dimethylformamide]). Heat to 60–80 °C for 20 min to increase the reaction rate. 1–2 drops of TMCS (trimethylchlorosilane) or TSIM will also speed up the reaction.

with BSA in combination with other silylation reagents · MN Appl. No. 213100

BSA alone silylates all sterically unhindered hydroxyl groups of the steroid skeleton; addition of TMCS will enable reaction of moderately hindered OH groups (reaction time 3–6 h at 60 °C). After addition of TSIM even strongly hindered hydroxyl groups will react (reaction time 6–24 h at 60 °C).

with MSTFA, MSHFBA or MBDSTFA
MSTFA MN Appl. No. 213111 · MSHFBA MN Appl. No. 213112 · MBDSTFA MN Appl. No. 213113

Dissolve 10–15 mg sample in 0.8 mL solvent, then add 0.2 mL of the silylation reagent. The reaction mixture can be heated to 60–70 °C for up to 1 h and can be analyzed directly. If TFA is used as a solvent, proceed as follows [20]: dissolve 1–2 mg sample in 100 µL TFA. Dropwise add 0.9 mL of the silylating reagent. After cooling the sample can be chromatographed directly.

with TSIM or SILYL-1139 (TSIM – pyridine 11:39) · TSIM MN Appl. No. 213121 · SILYL-1139 MN Appl. No. 213122

Dissolve 10–15 mg sample in 0.8 mL solvent, then add 0.2 mL of the silylation reagent. The reaction mixture can be heated to 60–70 °C for up to 1 hour and can be analyzed directly. Recommended solvent pyridine. When using SILYL-1139, the presence of water does not interfere.

with SILYL-21 or SILYL-2110 · SILYL-21 MN Appl. No. 213131 · SILYL-2110 MN Appl. No. 213132

Carefully add SILYL-21 or SILYL-2110 to 1–10 mg of the sample. Precipitated ammonium chloride does not interfere. If the sample should not dissolve within 5 min, heat to 75–85 °C. If no mutarotation is to be expected, you may dissolve the sugar in warm pyridine first and then add the silylation reagent. In some cases it may be advantageous to use a different solvent instead of pyridine. For derivatization of 3-ketosteroids we recommend to use DMF (dimethylformamide)

O-trimethylsilylation with MSTFA followed by *N*-trifluoroacetylation with MBTFA · MN Appl. No. 213140

Completely silylate 2 mg of the sample with 0.3 mL MSTFA, e.g., as described on page 363. After addition of 0.3 mL MBTFA the *N*-trimethylsilyl group is replaced by the *N*-trifluoroacetyl group. The mixture can be analyzed directly.

Test mixtures for GC capillary columns

Test mixtures

Key features

- Test mixtures for GC capillary columns to control the performance of fused silica capillary columns and the GC system

Ordering information

Test mixtures*

Designation		Pack of	REF
Activity test mixture (FA-TMS test according to Donike) in MSTFA/*n*-hexane (1 + 4)	1 mg/mL each of TMS capric acid (C10), TMS myristic acid (C14), TMS stearic acid (C18), TMS behenic acid (C22), hexadecane (C16), eicosane (C20), tetracosane (C24), octacosane (C28)	1 mL	722307
Grob test mixture (modified) in *n*-hexane	(in mg/mL) *n*-decane (~ 2.8), *n*-undecane (~ 2.9), *n*-octanol (~ 3.6), 2,6-dimethylphenol (~ 3.2), 2,6-dimethylaniline (~ 3.2), methyl decanoate (~ 4.2), dicyclohexylamine (~ 3.1), methyl undecanoate (~ 4.2), methyl dodecanoate (~ 4.1)	1 mL	722310
MN OPTIMA® test mixture in pentane	0.1 % each of undecane, dodecane, octanol, dimethylaniline, decylamine, methyl decanoate, methyl undecanoate, henicosane, docosane, tricosane (chromatograms see page 305)	1 mL	722316
MN OPTIMA® amine test mixture in ethanol	0.2 % diisobutylamine, 1 % diethanolamine, 0.2 % 2,6-dimethylaniline, 0.2 % *o*-propanol-pyridine, 0.2 % dicyclohexylamine, 0.2 % dibenzylamine	1 mL	722317
FAME test mixture in hexane	0.1 % each of FAMEs C4, C6, C8, C10, C12, C14, C16, C18, C18:1 cis, C18:1 trans, C18:2, C18:3, C20, C22, C22:1, C24 (chromatogram see page 334)	1 mL	722320

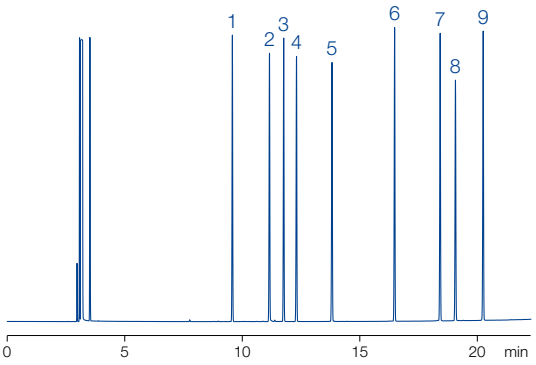
* These products contain harmful substances which must be specially labeled as hazardous. For detailed information please see SDS.

Grob test mixture (modified) (REF 722310)

MN Appl. No. 211250

Column: OPTIMA® 5, 50 m x 0.25 mm ID, 1.0 µm film
Injection: 1 µL, split 1:40, 280 °C
Carrier gas: 1.5 bar H_2
Temperature: 80 °C → 280 °C (10 min), 8 °C/min
Detector: FID 280 °C

Peaks:
1. *n*-Decane
2. 1-Octanol
3. *n*-Undecane
4. 2,6-Dimethylphenol
5. 2,6-Dimethylaniline
6. Methyl decanoate
7. Methyl undecanoate
8. Dicyclohexylamine
9. Methyl dodecanoate



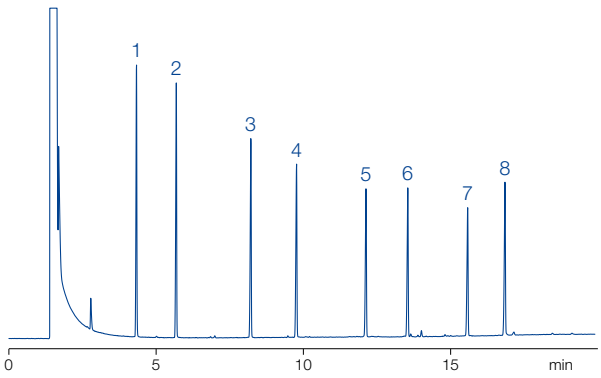
Test mixtures for GC capillary columns

Activity test mixture (REF 722307)

MN Appl. No. 211240

Column:	OPTIMA® 5, 25 m x 0.32 mm ID, 1.0 µm film
Injection:	1 µL, split 1:40, 300 °C
Carrier gas:	0.6 bar H_2
Temperature:	150 °C → 300 °C (8 min), 10 °C/min
Detector:	FID 300 °C

Peaks:
1. TMS capric acid (C_{10})
2. Hexadecane (C_{16})
3. TMS myristic acid (C_{14})
4. Eicosane (C_{20})
5. TMS stearic acid (C_{18})
6. Tetracosane (C_{24})
7. TMS behenic aicd (C_{22})
8. Octacosane (C_{28})

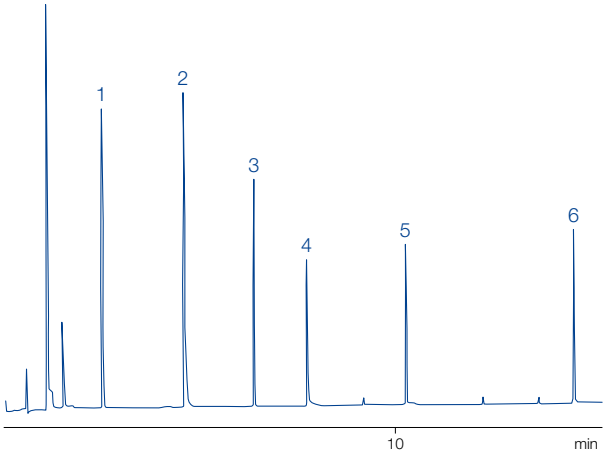


OPTIMA® Amine test mixture (REF 722317)

MN Appl. No. 250020

Column:	OPTIMA® 5 Amine, 30 m x 0.32 mm ID, 1.5 µm film
Injection:	1 µL, split 1:40
Carrier gas:	0.6 bar H_2
Temperature:	100 °C → 280 °C, 10 °C/min
Detector:	FID 280 °C

Peaks:
1. Diisobutylamine
2. Diethanolamine
3. 2,6-Dimethylaniline
4. o-Propanol-pyridine
5. Dicyclohexylamine
6. Dibenzylamine



Ferrules for capillary columns

Ferrules

Key features

- Graphite ferrules provide the highest temperature stability (up to 450 °C). They are reusable, if handled with care. We also offer 1/16” graphite ferrules specially designed for Carlo Erba / Fisons or for Agilent gas chromatographs.
- Vespel ferrules with 40 % graphite. Temperature-stable up to 400 °C and reusable.

Ordering information

Ferrules

Bore (= column OD)	Graphite	Vespel +40 % Graphite
T_{max} →	450 °C	400 °C
1/16” ferrules		
0.4 mm		706246
0.5 mm	708308	
1/16” ferrules for Carlo Erba (Fisons) instruments		
0.8 mm	708340	
1/16” ferrules for Hewlett-Packard (Agilent) instruments		
0.4 mm	708353	
0.5 mm	708354	
0.8 mm	708355	
1/8” ferrules		
no bore	708341	
1/4” ferrules		
no bore	708344	
0.4 mm	708345	
0.5 mm	708346	

Injection Port Septa blister pack for cleanliness and easily handling

Key features

- BTO septa
  for highest demands in GC and GC-MS
  – pierced, soft – CenterGuide™
- AG3 septa
  with higher durability than BTO
  – pierced, hard – CenterGuide™
- Marathon Septa
  with extreme durability for > 400 injections
  – pierced – CenterGuide™

Ordering information

Injection port septa

Septum grade		BTO septa	AG3 septa	Marathon septa
				
OD	T_{max}			
9 mm	400 °C	702646	702656	702660
11 mm	400 °C	702647	702657	702661
11.5 mm	400 °C	702648	702658	702662
Shimadzu®	300 °C	702649	702659	702663
	Pack of	25	25	25

Standard Septa in classical plastic container

Key features

- Standard septa (ST)
  beige silicone, 60° shore A, 4 mm
- High temperature septa (HT)
  red non-bleeding silicone, 60° shore A, 3 mm
  (320 °C max.)
- Silicone septa
  soft, transparent
- Silicone / PTFE septa
  white silicone, one side coated with grey PTFE, 3 mm

Ordering information

Classical septa

Septum grade		Standard septa (ST)	High temperature septa (HT)	Silicone septa	Silicone septa / PTFE
					
OD					
9 mm		702609	702619	702602	
10 mm		702610	702620		702625
11 mm		702611	702621	702604	702626
12 mm		702612	702622	702605	702627
13 mm		702613	702623	702606	702628
17 mm			702632		
	Pack of	50	50	50	50

Connectors for capillary GC columns

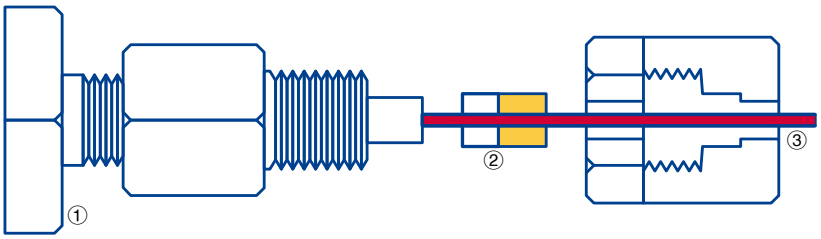
Key features

- Glass connectors for fused silica capillary columns from 0.2 to 0.53 mm ID:
  manufactured from deactivated glass with slightly tapered inner diameter; used to join two fused silica capillaries of equal or different diameters. Advantages compared to stainless steel fittings are easy connection without tools, optical control during connection, negligible heat capacity and no dead volume.
- Graphseal ferrules for capillary columns: a stainless steel ferrule filled with graphite – the ideal sealing material for capillaries. The capillary is mounted on a 1/16" exit (detector, injector etc.), with the appropriate ferrule, a nut (with slit) and an adapter (see table below).

Ordering information

Connectors for capillary GC columns

Description	Pack of	REF
Graphseal ferrules for capillary columns		
0.4 mm bore	10 ferrules	708337
0.5 mm bore	10 ferrules	708318
0.8 mm bore	10 ferrules	708319
Universal capillary glass connectors		
linear	5 connectors	707971
linear	10 connectors	707972
Y splitter	1 connector	707973



① 1/16" exit
② Graphseal ferrule
③ Capillary

Tools and general accessories for GC

Key features

- Magnifying lens with scale: an essential tool for any laboratory. In capillary GC it is often important to inspect column integrity or check cut ends of capillaries. When closing a column by melting the magnifying lens can be used to check whether the column is really closed or whether an open channel has been formed in the sealed end. Our lens provides 8fold magnification and is supplied with a scale as pictured in the figure below. The space between lines is equivalent to 1/10 mm.
- Diamond file: a useful tool for cutting capillaries and smoothing ends of capillaries. Square capillary ends are especially important for butt connections (e.g., in Valco unions).
- Glass wool, quartz wool and glass fiber wadding are used for, e.g., GC liners, packed GC columns etc.

Ordering information

Tools and general accessories

Description		Pack of	REF
Tools for capillary GC			
Diamond file	for cutting capillaries and straightening capillary ends	1	708300
Magnifying lens with scale	magnification 8x	1	706296
PTFE tape for sealing, reels 12 m long, 12 mm wide, 0.1 mm thick		1 reel	706512
Glass wool			
Glass wool, long fibers, DMCS treated, for packed GC columns		50 g	706201
Glass fiber wadding silanized, very fine fibers		25 g	718002
Quartz wool, very fine fibers		25 g	718587

Contents

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Glass plates

ALUGRAM® Xtra aluminum sheets
ALUGRAM® aluminum sheets

POLYGRAM® polyester sheets

Thin layer chromatography (TLC) and high performance thin layer chromatography (HPTLC), also called planar chromatography, are, like all chromatographic techniques, based on a multistage distribution process involving

- Suitable adsorbents (the stationary phase) coated as a thin layer onto a suitable support (e.g., glass plate, polyester or aluminum sheet; also see page 272)
- Solvents or solvent mixtures (the mobile phase or eluent)
- Sample molecules

The principle of TLC is known for more than 100 years [11]. The real break-through as an analytical method, however, came about 50 years ago as a consequence of the pioneering work of Egon Stahl [12].

Today TLC has gained increasing importance as an analytical separation technique, which is probably due to effects of instrumentation and automation [13]. At the same time the applicability of thin layer chromatography was enhanced by development of new adsorbents and supports.

Today MACHEREY-NAGEL offers a versatile range of ready-to-use layers, which are the result of 50 years of continuous research and development.

Features of modern TLC / HPTLC

The success of thin layer chromatography as a highly efficient microanalytical separation method is based on a large number of advantageous properties:

- High sample throughput in a short time
- Suitable for screening tests
- Pilot procedure for HPLC and Flash chromatography
- After separation the analytical information can be stored for a longer period of time (the TLC ready-to-use layer acts as storage medium for data)
- Separated substances can be subjected to subsequent analytical procedures (e.g., IR, MS) at a later date
- Rapid and cost-efficient optimization of the separation due to easy change of mobile and stationary phase

Principle steps of a TLC separation

Sample preparation

For separation the sample must meet several requirements to obtain good results. Since the TLC plate is a disposable product, sample preparation in general is not as demanding as for other chromatographic methods. However, eventually several steps for sample pretreatment may be necessary. These include sampling, mechanical crushing, extraction steps, filtration and sometimes enrichment of interesting components or clean-up, i.e. removal of undesired impurities.

Our TLC micro-sets introduce some simple methods of sample pretreatment. The dyes or dye mixtures of the beginner's set do not require complicated procedures. The advanced sets require the user to carry out some additional steps for preparing a sample, thus introducing the user to techniques often performed in industrial laboratories.

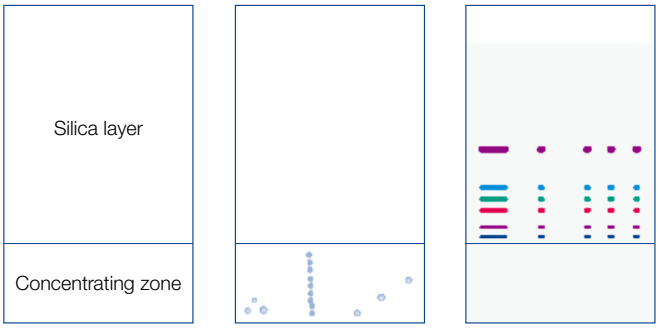
Thorough preparation of samples is an important prerequisite for the success of a TLC separation. For our range of products for more demanding sample pretreatment please see the chapter "SPE" from page 10.

Sample application

The most frequent technique is application with a glass capillary as spot or short streak.

Application as streak will yield better results especially for instrumental quantification. For both types of application some manual skill is required to obtain reproducible results. Substance zones which are too large from the beginning will cause poor separation since during chromatography they will become even larger and more diffuse.

A valuable aid for manual application especially of large volumes of very dilute samples is the concentrating zone (e.g., SILGUR-25 UV_{254}), which consists of a chromatographically inactive adsorbent (kieselguhr). The substances to be separated are concentrated to a small band in the concentrating zone and the separation starts at the beginning of the chromatographically active adsorbent silica.



Another method for sample concentration is a short pre-elution (few mm) with a solvent, in which all substances have a high R_f value.

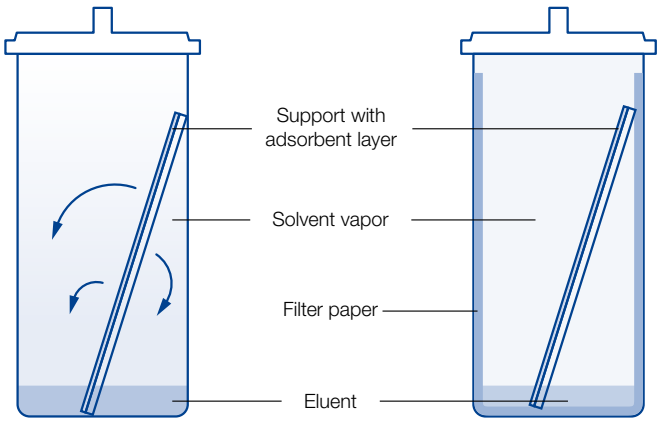
If a quantitative evaluation with a TLC scanner is to follow the separation we recommend to use commercially available sample applicators for spotting. These range from simple spotting guides via nanoapplicators to completely automated spotting devices. Application as streak can be performed automatically by spraying of the sample without touching the layer of the TLC plate. Application as band over the whole width of the TLC plate is especially important for preparative TLC. After application allow the solvent of the samples to evaporate completely (about 10 min) or blow with cold or hot air. Development of a chromatogram should never start before the solvent of the applied samples is evaporated completely.

Developing a chromatogram – separation techniques

The most frequently used separation technique is ascending TLC in a trough chamber (standard method, linear development). Usually it is applied as single development. However, multiple development, with or without change of eluent (step technique) can improve separation results. For 2-dimensional development only 1 spot of the sample is applied in one edge of a plate. After chromatography in the first direction the plate is dried, turned by 90° and developed in the 2nd dimension with another eluent. Thus complicated mixtures give 2-dimensional chromatograms taking advantage of the different separating properties of two eluents.

For selection and optimization of the eluent numerous publications are available. A generally applicable standardized optimization method is described by H. Keuker et al. [14].

It is important to pay attention to the atmosphere in the developing chamber. If reproducible migration distances are required, saturation of the chamber atmosphere with eluent vapor is necessary. For this purpose the developing chamber is lined with well absorbing chromatography paper (e.g., MN 260) and charged with a correspondingly larger volume of eluent.



A) Normal saturation, arrows show evaporation of the eluent from the layer

B) Chamber lined with filter paper, saturated with eluent vapor

Evaluation of a thin layer chromatogram

Evaluation depends on the purpose of the chromatographic analysis. For qualitative determination often localization of substances is sufficient. This can be easily achieved by parallel runs with reference substances.

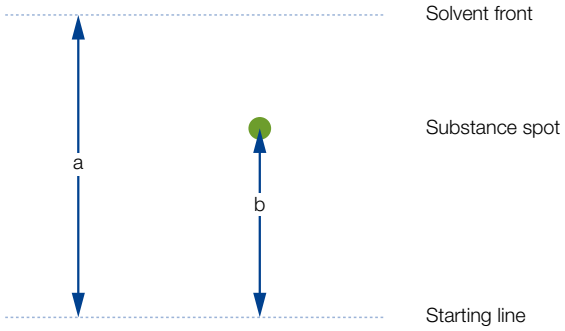
A parameter often used for qualitative evaluation is the R_f value (retention factor) or the 100-fold value hR_f. The R_f value is defined as follows:

$$R_f = \frac{\text{distance starting line – middle of spot}}{\text{distance starting line – solvent front}} = \frac{b}{a}$$

i.e. the R_f values are between 0 and 1, best between 0.1 and 0.8 (i.e. 10–80 for hR_f). If reproducible R_f values are to be obtained, it is essential that several parameters such as chamber saturation, composition of solvent mixtures, temperature etc. are strictly controlled.

Quantitative evaluation is possible by suitable calibration measurements. For this purpose either the area of a substance spot is measured or a photometric evaluation is performed directly on the layer. The latter procedure, however, requires a higher instrumental expense.

The following paragraphs describe the most frequently used methods for evaluation in TLC.



Qualitative detection

Qualitative evaluation is generally made directly on the TLC plate via characteristic R_f values of substances, i.e. the ratio of distance start – substance zone to distance start – solvent front and specific chemical reactions.

Visualization of separated substances

First of all it is necessary to recognize the position of a substance spot. Only in very few cases the sample is a dye which can be seen with the naked eye. Much more often for unspecific visualization substances can be viewed under UV light, since many substances show a UV absorption. If a fluorescent indicator is added to the layer, all substances absorbing in the respective region of wave length cause a quenching of the fluorescence, i.e. they appear as dark spots on the fluorescent layer. Customary fluorescent indicators are excited at 254 nm or (less frequently) at 366 nm with a mercury lamp. For our program of fluorescent indicators for TLC please see page 296.

Quenching of the fluorescence

Identification of separated substances is possible via the R_f value compared to the pure compound, which is often applied simultaneously on the same plate.

For a number of compounds their native fluorescence can be used for visualization, which is excited by UV light (mostly long-wave UV) (e.g., aflatoxins). This allows not only determination of the R_f value, but often enables a further qualitative assignment.

If these methods do not allow localization or characterization of a substance, post-chromatographic detection methods can be applied, chemical reactions on the TLC plate [15]. Quite unspecific reactions are iodine adsorption and the charring technique (spraying with sulfuric acid and heat treatment).

More reliable results are possible with specific reagents for spraying or dipping, which form colored or fluorescent compounds with the substances to be detected. Depending on the sensitivity of these reactions they are not only used for group or substance specific characterization (in addition to the R_f value) but also for quantification down to trace levels. As example take the ninhydrin reaction. Formation of a (usually red) zone with this detection method yields the information, that a certain group of substances, e.g., α-amino acids, are present. The R_f value allows further assignment to one or several single compounds.

For identification of a substance a combination of different detection methods can be useful. Thus almost all lipids can be converted to products with light green fluorescence by reaction with 2',7'-dichlorofluorescein. Adsorption of iodine vapor enables a differentiation between saturated and unsaturated lipids or lipids containing nitrogen. And finally the R_f value is a third means of identification.

Here are some general remarks concerning spraying: use all spray reagents under a fume hood. The developed, dried TLC plate or sheet is placed on a sheet of filter paper for spraying. Usually it is sufficient to fill the sprayer with about 5–10 mL solution. Spray from a distance of about 15 cm with the aid of a rubber ball or – if available – with pressurized air. It is always better to spray a layer twice very thinly and evenly (with intermediate drying), than to saturate the layer with excessive spray reagent. In the latter case spots tend to become diffuse. After visualization mark outlines of zones with a lead pencil, because some spots tend to fade after a while.

Especially for quantitative evaluation short dipping of the layer in the respective reagent solution is recommended. For this purpose automatic instruments are commercially available, which allow reproducible dipping.

When a substance is localized on the TLC plate (e.g., under UV), but not yet identified, TLC scanners allow recording of UV spectra of individual substance zones directly on the layer, or the zone is removed by scratching or cutting (for sheets), eluted and further analyzed, e.g., by FT-IR, RAMAN, NMR or mass spectroscopy.

Quantitative evaluation

Often TLC is considered to be only a semiquantitative analytical procedure. This is true for visual evaluation of spots, since the eye can only compare but not measure absolute values. If, however, a direct optical evaluation ("in situ" measurement) is performed on the TLC plate with a thin layer scanner, after measurement of calibration functions, exact quantitative results are possible. Commercial scanners offer many features such as evaluation in absorption and fluorescence, unattended programmed scanning of lanes, multi-wave length measurement, background correction, selectable base line for integration, recording of spectra, evaluation of circular or anti-circular chromatograms with very high ease of operation. In addition to manual operation control by a computer is possible with respective data collection and storage. Usually wavelengths from 200 to 700 nm are available (visible and UV), e.g., all post-chromatographic (and of course all pre-chromatographic) visualization procedures are evaluated with the proper wavelength, which is determined with the instrument. Time requirements for all these possibilities are extremely low. Interlaboratory experiments with standard deviations of 2 % show how excellent results are obtainable [16].

TLC micro-sets introductory kits for science education

Beginner's set

- Features separations with simple developing solvents; samples are colored thus eliminating the need for visualization.
- All equipment needed is contained in the set.

Advanced sets F1, F2 and F3

- Require some experience and skill from the user: some of the samples have to be pretreated before separation, and for identification of substances spray reagents have to be used

TLC micro-set A for beginners

This kit contains all chemicals and accessories for the following separations:

- Separation of the fat-soluble (lipophilic)
  Test dye mixture 1: butter yellow, indophenol, sudan blue II, sudan red G
- Separation of a mixture of anthraquinone dyes
  Test dye mixture 2: blue 1, blue 3, green, green blue, red, violet 1, violet 2
- Separation of a mixture of food dyes
  Test dye mixture 3: brilliant black BN (E151), fast red E, erythrosine (E127), yellow orange S (sunset yellow CFC, E110), naphthol red S, ponceau 4 R (E124), tartrazine (E102)
- Separation of dyes from felt tip pens

Contents of TLC micro-set A for beginners

1 manual

3 developing chambers

50 glass capillaries 1 µL

1 spotting guide

2 felt tip pens

1 measuring cylinder 10 mL

50 polyester sheets 4 x 8 cm each of POLYGRAM®: SIL G/UV_{254}, Alox N/UV_{254} and CEL 300

8 mL each of test dye mixture 1 (4 lipophilic dyes), test dyes sudan red G, and sudan blue II

8 mL each of test dye mixture 2 (7 anthraquinone dyes), test dyes blue 1 and violet 2

8 mL each of test dye mixture 3 (7 food dyes), test dyes yellow orange S, and brilliant black BN

100 mL each of toluene, toluene – cyclohexane (2:1, v/v), ethanol, 2.5 % sodium citrate solution, 25 % ammonia solution – 2-propanol (5:3, v/v)

Ordering information

Designation	Pack of	REF
TLC micro-set A for beginners*	1 kit	814000
Replacement parts for TLC micro-set A		
Test dye mixture 1*, solution of 4 lipophilic dyes in toluene (components see above)	8 mL	814001
Test dye mixture 2*, solution of 7 anthraquinone dyes in toluene – cyclohexane (2:1, v/v) (components see above)	8 mL	814002
Test dye mixture 3, aqueous solution of 7 food dyes (components see above)	8 mL	814003
Collection of 4 individual components of test dye mixture 1*	4 x 8 mL	814011
Collection of 7 individual components of test dye mixture 2*	7 x 8 mL	814012
Collection of 7 individual components of test dye mixture 3	7 x 8 mL	814013
Sodium citrate, 2.5 g in 100 mL bottle to fill up with distilled water	2.5 g	814029

* These products contain harmful substances which must be specially labeled as hazardous. For detailed information please see SDS.

Information about the advanced sets F1, F2 and F3 can be found on page 270 and page 271.

Introductory kits

TLC micro-set F1

This kit contains all chemicals required for the separation of

- Amino acids (test mixture, consisting of alanine, arginine, tryptophan and valine)
- Amino acids in urine
- The heavy metal cations copper(II) and manganese(II)

Contents of TLC micro-set F1

1 manual, 50 glass capillaries 1 µL

50 polyester sheets 4 x 8 cm each of POLYGRAM®: SIL G/UV_{254} and CEL 300

100 mL each of *n*-butanol, ninhydrin spray reagent (0.2 % in ethanol), acetone, 25 % ammonia solution, rubeanic acid spray reagent

50 mL each of 50 % acetic acid, 18 % hydrochloric acid

8 mL each of the amino acid test mixture (see left), tryptophan and arginine reference solutions

8 mL each of the heavy metal cation test mixture (see left), Cu^{2+} and Mn^{2+} reference solutions

TLC micro-set F2

This kit contains all chemicals required

- For analysis of edible fats
- For analysis of fats and cholesterol in blood

Contents of TLC micro-set F2

1 manual, 50 glass capillaries 1 µL

50 polyester sheets 4 x 8 cm POLYGRAM®: SIL G/UV_{254}

5 disposable pipettes 25 µL

5 sample vials N 11 (1.5 mL) with PE caps and seals

3 sample vials 30 mL (for butter, margarine and edible oil)

100 mL each of cyclohexane and molybdatophosphoric acid spray reagent

2 x 50 mL acetone with calibrated pipette

25 mL butan-2-one

8 mL cholesterol reference solution

TLC micro-set F3

This kit contains all chemicals required

- For separation of analgetics (pain relievers)
- For drug analysis as shown for cinchona bark

Contents of TLC micro-set F3

1 manual, 50 glass capillaries 1 µL

50 polyester sheets 4 x 8 cm POLYGRAM®: SIL G/UV_{254}

5 Aspirin® tablets, 5 Thomapyrin® tablets

20 folded filters MN 615 1/4, 11 cm diameter

3 sample vials 8 mL (for Aspirin® sample, Thomapyrin® sample, cinchona bark extract), 5 g cinchona bark

100 mL each of ethanol, 2-propanol, toluene – diethyl ether je 100 mL Ethanol, 2-Propanol, Toluol – Diethylether (61:39, v/v), spray reagent for caffeine and spray reagent according to Dragendorff-Munier

50 mL each of iron(III) chloride solution and potassium hexacyanoferrate(III) solution, 30 mL ethyl acetate

25 mL each of 12.5 % ammonia solution and diethylamine

8 mL each of caffeine, paracetamol, quinine reference solutions

All experiments with TLC micro-sets F1–F3 require the materials kit (see TLC micro-set M on page 271).

Ordering information

Designation	Pack of	REF
TLC micro-set F1*	1 kit	814200
Refill reagents for TLC micro-set F1		
Amino acid test mixtures (components see previous page)	8 mL	814201
Collection of 4 individual components of the amino acid test mixture	4 x 8 mL	814202
Cation test mixture (components see previous page)	8 mL	814204
Collection of 2 individual components of the cation test mixture (Cu^{2+}, Mn^{2+})	2 x 8 mL	814205
TLC micro-set F2*	1 kit	814300
Refill reagents for TLC micro-set F2		
Cholesterol reference solution*	8 mL	814301
TLC micro-set F3*	1 kit	814400
Refill reagents for TLC micro-set F3		
Quinine reference solution*	8 mL	814405
Paracetamol reference solution*	8 mL	814406
Caffeine reference solution*	8 mL	814407
Refill packs TLC sheets for all TLC micro-sets		
TLC polyester sheets POLYGRAM® SIL G/UV_{254}, 4 x 8 cm	4 x 50	814025
TLC polyester sheets POLYGRAM® Alox N/UV_{254}, 4 x 8 cm	4 x 50	814026
TLC polyester sheets POLYGRAM® CEL 300, 4 x 8 cm	4 x 50	814027
TLC polyester sheets POLYGRAM® 4 x 8 cm: 100 x SIL G/UV_{254}; 50 x Alox N/UV_{254}; 50 x CEL 300	1 kit	814028

* These products contain harmful substances which must be specially labeled as hazardous. For detailed information please see SDS.

Accessories for TLC micro-sets can be found under TLC accessories on page 295.
Spray reagents can be found on page 296.

TLC micro-set M

This kit is prerequisite for the separations with kits F1 to F3. In addition, it serves as basic equipment for the individual study of further thin layer chromatographic experiments.

Contents of TLC micro-set M (materials kit)

2 x 50 glass capillaries 1 µL, 2 spotting guides
1 rubber cap for capillaries
1 measuring cylinder 10 mL
1 beaker 25 mL
2 developing chambers
1 glass laboratory sprayer with rubber bulb
1 plastic syringe 1 mL
20 sheets filter paper MN 713 (15 x 21 cm)
50 polyester sheets 4 x 8 cm each of POLYGRAM®: SIL G/UV_{254}, Alox N/UV_{254} and CEL 300

Ordering information

Designation	Pack of	REF
TLC micro-set M (materials kit)	1 kit	814100

Summary of MN ready-to-use layers

Advantages of MN plates and sheets for TLC

Continuous high quality

- Guaranteed by stringent production control including standardized lot tests, surface checks for roughness or cracks as well as hardness and adherence checks

Comprehensive range of phases for TLC / HPTLC

- There is no universal TLC plate which meets all possible types of analyses
- Our versatile range of TLC ready-to-use layers covers many different types of applications

Immediately ready for chromatographic separation

- Coatings or impregnations are not necessary

Homogeneous, smooth, well adhering layers

- An important criterion especially for reproducible quantitative evaluation

Adsorbents for MN plates and sheets for TLC

Classical adsorbents

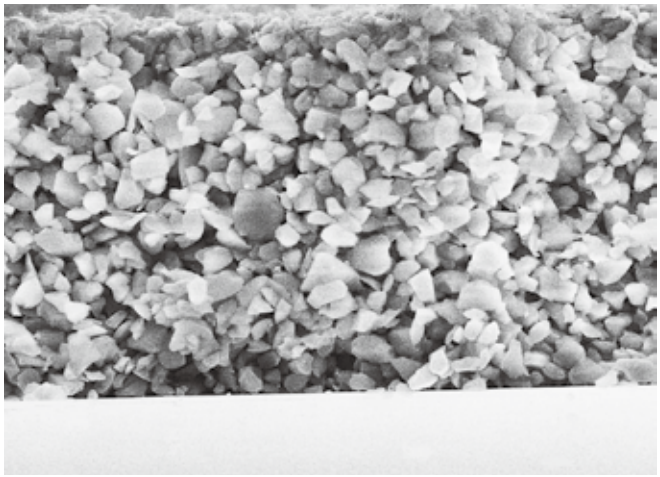
- For ~ 80 % of all TLC separations silica 60 (mean pore diameter 60 Å = 6 nm) is used
- Other classical adsorbents are aluminum oxide, cellulose, kieselguhr, ion exchangers and polyamide

Special phases

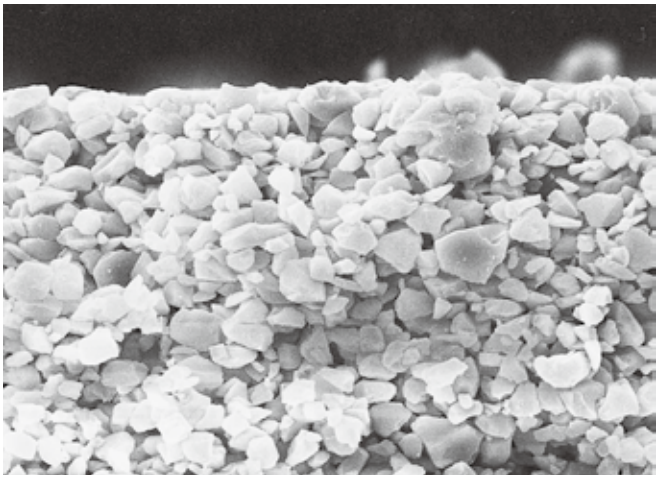
- Modified silica, like C_{18} (octadecyl-) cyano-, amino-, diol-, RP-2
- Special layers for specific separations, like PAH- or enantiomer separation

Particle size distribution and thickness of layer

- Are chosen to fit the given type of application (e.g., HPTLC, standard or preparative separations)
- Most MN ready-to-use layers are available with or without fluorescent indicator



Electron microscope photograph of a cross section through a glass plate with silica layer (magnification x 500)



Electron microscope photograph of a cross section through an aluminum sheet with silica layer (magnification x 500)

Supports for ready-to-use layers for TLC

	Glass plates G	POLYGRAM® P	ALUGRAM® A / ALUGRAM® Xtra Ax
Physical properties of support materials			
Material	glass	polyester	aluminum
Thickness (approx.)	1.3 mm	0.2 mm	0.15 mm
Weight, packaging and storage requirements	high	low	low
Torsional strength	ideal	low	relatively high
Temperature stability	high	max. 185 °C	high
Susceptible to breakage	yes	no	no
Can be cut with scissors	no	yes	yes
Chemical resistance of support materials			
Against solvents	high	high	high
Against mineral acids and conc. ammonia	high	high	low
Stability of the binder system of NP plates in water			
Suitability for aqueous detection reagents	depending on phase	very suitable	ALUGRAM®: limited suitability; ALUGRAM® Xtra: very suitable

Summary of MN ready-to-use layers

Summary

Phase	Support*	Layer	Page
Standard silica particle size 5–17 µm			
ADAMANT	G	silica 60, improved binder system, optimized particle size distribution	274
SIL G	G P A Ax	silica 60, standard grade	276
DURASIL	G	silica 60, special binder system	277
SILGUR	G Ax	silica 60 with kieselguhr concentrating zone	279
Unmodified silica for HPTLC particle size 2–10 µm			
Nano-SILGUR	G Ax	nano silica 60 with kieselguhr concentrating zone	279
Nano-ADAMANT	G	nano silica 60, improved binder system, optimized particle size distribution	281
Nano-SIL	G A Ax	nano silica 60, standard grade	281
Nano-DURASIL	G	nano silica 60, special binder system	282
Modified silica for HPTLC particle size 2–10 µm			
Nano-SIL C18-50 / Nano-SIL C18-100	G	nano silica with partial or complete C_{18} modification	283
RP-18 W/UV_{254}	G A	nano silica with partial octadecyl modification, wettable with water	284
RP-2/UV_{254}	G A	silanized silica = dimethyl-modified nano silica 60	284
Nano-SIL CN	G A	cyano-modified nano silica	285
Nano-SIL NH_2	G A	amino-modified nano silica	286
Nano-SIL DIOL	G	diol-modified nano silica	287
Aluminum oxide			
Alox-25 / Alox N	G P A	aluminum oxide	288
Cellulose, unmodified and modified			
CEL 300	G P A	native fibrous cellulose MN 300	289
CEL 400	G P	microcrystalline cellulose MN 400 (AVICEL®)	289
CEL 300 PEI	P	polyethyleneimine-impregnated cellulose ion exchanger	290
CEL 300 AC	P	acetylated cellulose MN 300	290
POLYAMID-6			
POLYAMID-6	P	perlon = ε-polycaprolactame	290
Layers for special separations			
CHIRALPLATE	G	RP silica with Cu^{2+} ions and chiral reagent, for enantiomer separation of amino acids	291
SIL N-HR	P	high purity silica 60, special binder system, higher gypsum content	291
SIL G-25 HR	G	high purity silica 60 with gypsum, recommended for aflatoxin analysis	292
SIL G-25 Tenside	G	silica G with ammonium sulfate for separation of surfactants	292
Nano-SIL PAH	G	nano silica with special impregnation for PAH analysis	292
IONEX-25 SA-Na	P	mixed layer of strongly acidic cation exchanger and silica	293
IONEX-25 SB-AC	P	mixed layer of strongly basic anion exchanger and silica	293
Alox / CEL-AC-Mix	G	mixed layer of aluminum oxide and acetylated cellulose	293
SILCEL-Mix	G	mixed layer of cellulose and silica	293

* G = Glass plates P = POLYGRAM® polyester sheets A = ALUGRAM® aluminum sheets Ax = ALUGRAM® Xtra aluminum sheets

Unmodified TLC silica layers

ADAMANT G unmodified standard silica layers

Key features

- Outstanding hardness and abrasion resistance due to an optimized binder system
- Increased separation efficiency due to an optimized particle size distribution
- High suitability for trace analysis resulting from a UV indicator with increased brilliance and a lownoise background of the layer

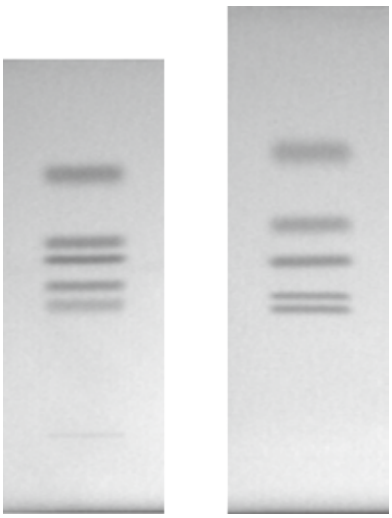
Technical characteristics

- Silica 60, mean pore size 60 Å, specific surface (BET) ~ 500 m^2/g, specific pore volume 0.75 mL/g, particle size 5–17 µm

Separation of steroids

MN Appl. No. 402930

Layers:	ADAMANT UV_{254}, SIL G/UV_{254}
Sample:	0.1 % solution in $CHCl_3$
Eluent:	chloroform – methanol (97:3, v/v)
Migration distance:	ADAMANT 50 mm in 10 min, SIL G 57 mm in 10 min
Detection:	UV



ADAMANT UV_{254} SIL G/UV_{254}

Substance	R_f ADAMANT	R_f SIL G
Cortisone	0.37	0.27
Corticosterone	0.43	0.30
Testosterone	0.50	0.39
Deoxycorticosterone	0.55	0.46
Progesterone	0.73	0.62

Separation of barbiturates

MN Appl. No. 402950

Layer:	ADAMANT UV_{254}
Sample volume:	1 µL
Eluent:	chloroform – acetone (95:5, v/v)
Migration distance:	70 mm in 20 min
Detection:	UV

ADAMANT UV_{254}

Substance	R_f
Thiamylal (0.5 %)	0.69
Thiopental (1.0 %)	0.65
Hexobarbital (5.0 %)	0.41
Pentobarbital (1.0 %)	0.26
Phenobarbital (1.0 %)	0.18

Ordering information

Plate size [cm]	2.5 x 7.5	5 x 10	5 x 10	5 x 20	10 x 10	10 x 20	20 x 20	Thickness of layer	Fluorescent indicator
Pack of [plates]	100	50	200	100	25	50	25		
Glass plates									
ADAMANT		821040	821040.200		821050		821060	0.25 mm	–
ADAMANT UV_{254}	821005	821010	821010.200	821015	821020	821025	821030	0.25 mm	UV_{254}

ALUGRAM® Xtra SIL G Ax unmodified standard silica layers on aluminum

Key features

- Outstanding wettability for precise colorization results, even with 100 % aqueous detection reagents
- Excellent separation efficiency and reproducibility from lot to lot
- Easy and reliable cutting due to an optimized binder system, no flaking of silica

Technical characteristics

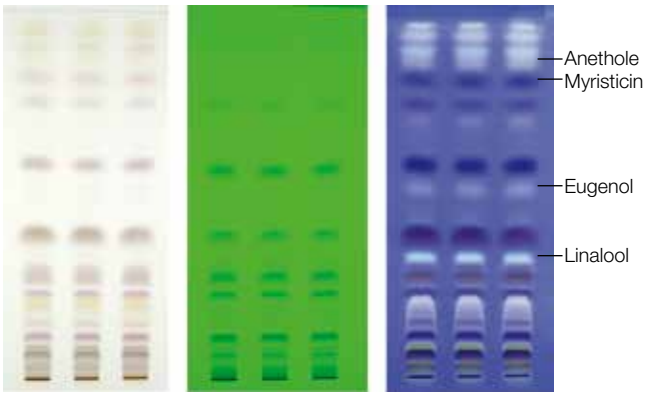
- Silica 60, mean pore size 60 Å, specific surface (BET) ~ 500 m^2/g, specific pore volume 0.75 mL/g, particle size 5–17 µm
- Binder: highly polymeric product, which is stable in almost all organic solvents and resistant towards aggressive visualization reagents, also completely stable in purely aqueous eluents

Separation of nutmeg ingredients

MN Appl. No. 403590

Layer:	ALUGRAM® Xtra SIL G UV_{254}
Sample:	shake 1.0 g freshly powdered drug for 3 min with 4 mL methanol and filter; apply 10 µL
Eluent:	toluene – ethyl acetate (95:5, v/v)
Migration distance:	15 cm
Detection:	254 nm: underivatized daylight and 366 nm: spray with 5 % ethanolic sulfuric acid, 1 % vanillic acid and heat to 105 °C

The chromatograms show the following zones with increasing R_f values: linalool (bluish grey), eugenol (yellowish brown), myristicin (reddish brown), and anethole (pink-violet). Other colored zones may appear.



Ordering information

Plate size [cm]	2.5 x 7.5	4 x 8	5 x 7.5	5 x 10	5 x 20	10 x 20	20 x 20	Thickness of layer	Fluorescent indicator
Pack of [plates]	200	50	20	50	50	20	25		
ALUGRAM® Xtra aluminum sheets									
SIL G			818230.20	818261	818232		818233	0.20 mm	–
SIL G/UV_{254}	818329	818331	818330.20	818360	818332	818362	818333	0.20 mm	UV_{254}

Further application examples can be found online in our application database at *www.mn-net.com/apps*

Unmodified TLC silica layers

SIL G G P A unmodified standard silica layers

Technical characteristics

- Silica 60, mean pore size 60 Å, specific surface (BET) ~ 500 m^2/g, specific pore volume 0.75 mL/g, particle size 5–17 µm
- Thickness of layer for analytical plates 0.25 mm, for preparative plates 0.5 and 1 mm; for 2 mm preparative layers a slightly coarser material is used
- Indicators: manganese activated zinc silicate with green fluorescence for short-wave UV (254 nm); special inorganic fluorescent pigment with blue fluorescence for long-wave UV (366 nm)
- Binders: highly polymeric products, which are stable in almost all organic solvents and resistant towards aggressive visualization reagents; binder system for POLYGRAM® sheets is also completely stable in purely aqueous eluents

Ordering information

Glass plates

Plate size [cm]	2.5 x 7.5	5 x 10	5 x 10	5 x 20	10 x 10	10 x 20	20 x 20	Thickness of layer
Pack of [plates]	100	50	200	100	25	50	25	
SIL G-25		809017	809017.200	809011		809012	809013	0.25 mm
SIL G-25 UV_{254}	809028.100	809027	809027.200	809021	809020	809022	809023	0.25 mm
SIL G-25 $UV_{254+366}$				809121		809122	809123	0.25 mm

Glass plates

Pack of [plates]	(preparative TLC)	20		
SIL G-50		809051	0.50 mm	
SIL G-50 UV_{254}		809053	0.50 mm	

Glass plates

Pack of [plates]	(preparative TLC)	15		
SIL G-100		809061	1.00 mm	
SIL G-100 UV_{254}		809063	1.00 mm	

Glass plates

Pack of [plates]	(preparative TLC)	12		
SIL G-200		809073	2.00 mm	
SIL G-200 UV_{254}		809083	2.00 mm	

POLYGRAM® polyester sheets

Plate size [cm]	2.5 x 7.5	4 x 8	5 x 20	20 x 20	40 x 20	
Pack of [plates]	200	50	50	25	25	
SIL G	805902	805032	805012	805013	805014	0.20 mm
SIL G/UV_{254}	805901	805021	805022	805023	805024	0.20 mm
SIL G/UV_{254}			roll 500 x 20 cm 805017			0.20 mm

ALUGRAM® aluminum sheets

Plate size [cm]	2.5 x 7.5	4 x 8	5 x 7.5	5 x 10	5 x 20	10 x 20	20 x 20	
Pack of [plates]	200	50	20	50	50	20	25	
SIL G			818030.20	818161	818032	818163	818033	0.20 mm
SIL G/UV_{254}	818129	818131	818130.20	818160	818132	818162	818133	0.20 mm

Further application examples can be found online in our application database at *www.mn-net.com/apps*

DURASIL G unmodified standard silica layers

Technical characteristics

- Silica 60, mean pore size 60 Å, specific surface (BET) ~ 500 m^2/g, specific pore volume 0.75 mL/g, particle size 5–17 µm
- Hard, water-resistant and wettable layers due to a special binder system

Ordering information

Plate size [cm]	5 x 10	5 x 10	5 x 20	10 x 20	20 x 20	Thickness of layer	Fluorescent indicator
Pack of [plates]	50	200	100	50	25		
Glass plates							
DURASIL-25				812003	812004	0.25 mm	–
DURASIL-25 UV_{254}	812005	812005.200	812006	812007	812008	0.25 mm	UV_{254}

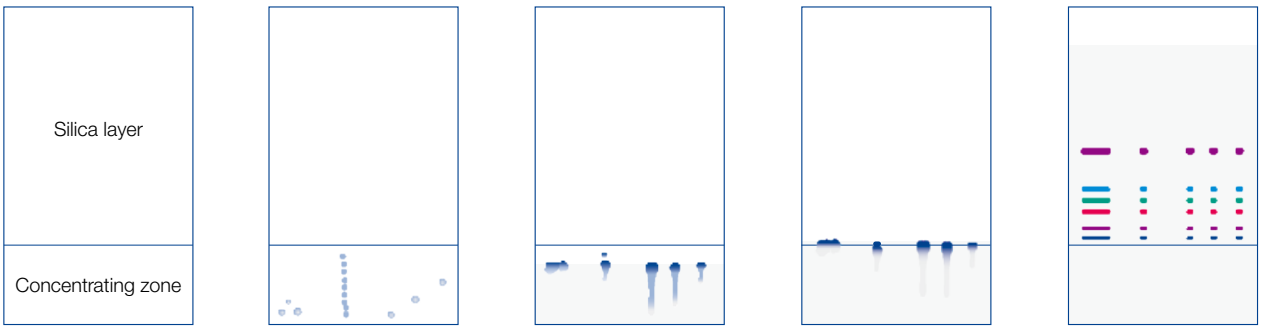
The most TLC layers are available as glass plate, polyester- or aluminum sheet (also see page 272 and 273).

Silica layers with concentrating zone

MN TLC pre-coated layers – qualitative and individual tailored

Kieselguhr zone

- For rapid sample application
- Because kieselguhr is completely inert towards a large number of compounds, the samples always form a narrow band at the interface of the two adsorbents, irrespective of shape, size or position of the spots in the concentrating zone. Separation then takes place in the silica layer.



Arbitrary application in the concentrating zone

Applied substances are concentrated

Applied substances at the phase boundary

Developed chromatogram

SILGUR G Ax unmodified standard silica layers with concentrating zone

Technical characteristics

- Silica 60, mean pore size 60 Å, specific surface (BET) ~ 500 m^2/g, specific pore volume 0.75 mL/g, particle size 5–17 µm
- Kieselguhr zone for rapid sample application (see page 278)
- Channel-plate with 19 channels help to prevent cross contamination by separating several samples
- More samples can be separated on a plate, and spot areas can be more easily determined

Ordering information

Plate size [cm]	10 x 20	20 x 20	Thickness of layer	Fluorescent indicator
Glass plates				
Pack of [plates]	50	25		
SILGUR-25	810012	810013	0.25 mm	–
SILGUR-25 UV_{254}	810022	810023	0.25 mm	UV_{254}
Channel-Plates				
Pack of [plates]		25		
SILGUR-25-C UV_{254}		810123	0.25 mm	UV_{254}
ALUGRAM® Xtra aluminum sheets				
Pack of [plates]	20	25		
SILGUR	818412	818413	0.20 mm	–
SILGUR UV_{254}	818422	818423	0.20 mm	UV_{254}

Nano-SILGUR G Ax unmodified HPTLC silica layers with concentrating zone

Technical characteristics

- Nano silica 60, pore size 60 Å, specific surface (BET) ~ 500 m^2/g, mean specific pore volume 0.75 mL/g, particle size 2–10 µm
- Kieselguhr zone for rapid sample application (see page 278)

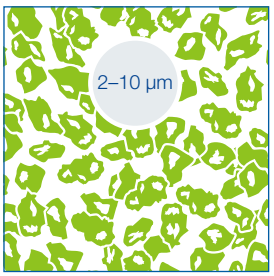
Ordering information

Plate size [cm]	10 x 10	Thickness of layer	Fluorescent indicator
Pack of [plates]	25		
Glass plates			
Nano-SILGUR-20	811032	0.20 mm	–
Nano-SILGUR-20 UV_{254}	811042	0.20 mm	UV_{254}
ALUGRAM® Xtra aluminum sheets			
Nano-SILGUR	818432	0.20 mm	–
Nano-SILGUR UV_{254}	818442	0.20 mm	UV_{254}

Sharper separation by nano silica

Nano silica for HPTLC

- Narrow fractionation of the silica particles allows theoretical plate heights, which are one order of magnitude smaller than on standard silica layers.



Nano silica

Standard silica

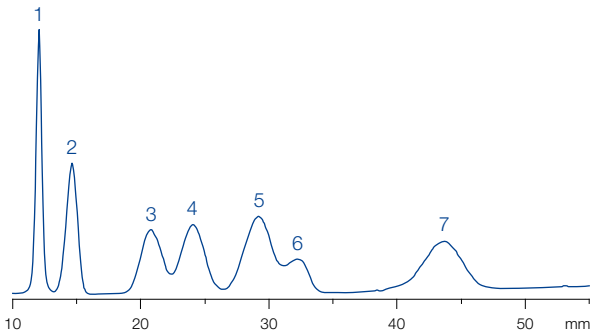
Advantages

- Shorter migration distances
- Lower amount of samples required
- Increased detection sensitivity with equal selectivity
- Less developing time

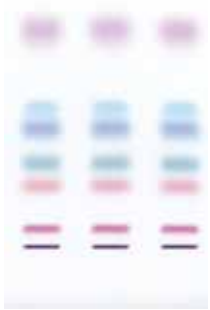
Comparison of ADAMANT and Nano-ADAMANT plates for separation of anthraquinone dyes

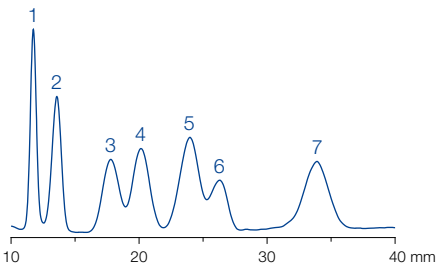
Layers: A) ADAMANT
B) Nano-ADAMANT
Sample: 1 µL, about 0.1 %
Eluent: toluene – cyclohexane (4:3, v/v)
Migration time: A) 30 min, B) 15 min

Peaks:
1. Blue 3
2. Violet 2
3. Red
4. Green
5. Blue 1
6. Greenish blue
7. Violet 1



A)





B)

Nano-ADAMANT G unmodified HPTLC silica layers

Key features

- Outstanding hardness and abrasion resistance due to an optimized binder system
- Increased separation efficiency due to an optimized particle size distribution
- High suitability for trace analyses resulting from a UV indicator with increased brilliance and a lownoise background of the layer

Technical characteristics

- Nano silica 60, mean pore size 60 Å, specific surface (BET) ~ 500 m^2/g, specific pore volume 0.75 mL/g, particle size 2–10 µm

Ordering information

Plate size [cm]	10 x 10	10 x 20	Thickness of layer	Fluorescent indicator
Pack of [plates]	25	50		
Glass plates				
Nano-ADAMANT	821140	821150	0.20 mm	–
Nano-ADAMANT UV_{254}	821110	821120	0.20 mm	UV_{254}

Nano-SIL G Ax A unmodified HPTLC silica layers

Technical characteristics

- Nano silica 60, mean pore size 60 Å, specific surface (BET) ~ 500 m^2/g, specific pore volume 0.75 mL/g, particle size 2–10 µm
- Indicator: manganese activated zinc silicate with green fluorescence for short-wave UV (254 nm)
- Binder: highly polymeric product, which is stable in almost all organic solvents and resistant towards aggressive visualization reagents

Ordering information

Plate size [cm]	5 x 5	5 x 20	10 x 10	10 x 20	20 x 20	Thickness of layer	Fluorescent indicator
Pack of [plates]	100	50	25	50	25		
Glass plates							
Nano-SIL-20	811011		811012	811013		0.20 mm	–
Nano-SIL-20 UV_{254}	811021		811022	811023		0.20 mm	UV_{254}
ALUGRAM® Xtra aluminum sheets							
Nano-SIL G		818240			818241	0.20 mm	–
Nano-SIL G/UV_{254}		818342			818343	0.20 mm	UV_{254}
ALUGRAM® aluminum sheets							
Nano-SIL G					818141	0.20 mm	–
Nano-SIL G/UV_{254}					818143	0.20 mm	UV_{254}

Unmodified HPTLC silica layers

Nano-DURASIL G unmodified HPTLC silica layers

Technical characteristics

- Nano silica 60, mean pore size 60 Å, specific surface (BET) ~ 500 m^2/g, specific pore volume 0.75 mL/g, particle size 2–10 µm
- Indicator: manganese activated zinc silicate with green fluorescence for short-wave UV (254 nm)
- Hard, water-resistant and wettable layers due to a special binder system
- Different selectivity compared to ADAMANT and SIL-G plates no reversed phase tendency, more polar than Nano-SIL

Ordering information

Plate size [cm] Pack of [plates]	10 x 10 25	10 x 20 50	Thickness of layer	Fluorescent indicator
Glass plates				
Nano-DURASIL-20	812010	812011	0.20 mm	–
Nano-DURASIL-20 UV_{254}	812013	812014	0.20 mm	UV_{254}

Nano-SIL C18 G octadecyl-modified HPTLC silica layers

Technical characteristics

- Nano silica 60, mean pore size 60 Å, specific surface (BET) ~ 500 m^2/g, specific pore volume 0.75 mL/g, pH stability 2–10, particle size 2–10 µm
- Indicator: acid-resistant product with a pale blue fluorescence for short-wave UV (254 nm), UV-absorbing substances appear as dark-blue to black spots on a light-blue background

Modification

- Partial (50 %) or complete (100 %) octadecyl modification, carbon content 7.5 and 14 %, respectively
- Order of polarity: silica > DIOL > NH_2 > CN > RP-2 > C18-50 > RP-18 W > C18-100

Recommended application

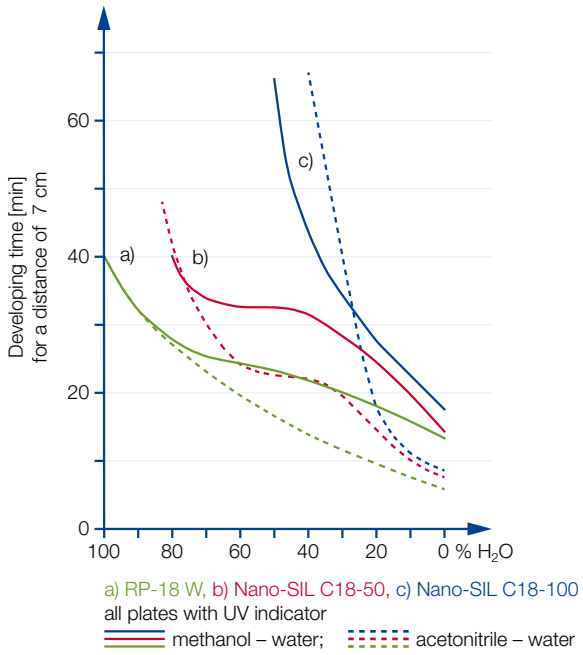
- Reversed phase separation mode with eluents from anhydrous solvents to mixtures with high concentrations of water (see table and figure below)
- Alkaloids, amino acids, preservatives, optical brighteners, barbiturates, polycyclic aromatic hydrocarbons (PAH), drugs, peptides, flavonoids, phenols, indole derivatives, steroids

Ordering information

Plate size [cm]		10 x 10	Thickness of layer	Fluorescent indicator
Pack of [plates]		25		
Glass plates				
Nano-SIL C18-50	50 % silanized	811054	0.20 mm	–
Nano-SIL C18-50 UV_{254}	50 % silanized	811064	0.20 mm	UV_{254}
Nano-SIL C18-100	100 % silanized	811052	0.20 mm	–
Nano-SIL C18-100 UV_{254}	100 % silanized	811062	0.20 mm	UV_{254}

Eluent	v/v	Migration distances [mm/15 min]		
		C18-50	C18-100	RP-18 W
Methanol – H_2O	2:1	57	45	44
	1:1	52	21	40
	1:2	50	0	43
	1:3	40	0	45
	1:4	30	0	46
	0:1	0	0	54
Acetonitrile – H_2O	2:1	62	46	66
	1:1	52	30	54
	1:2	51	27	46
	1:3	48	15	44
	1:9	20	0	42
Trichloromethane		68	64	71

Migration of C18-50 and C18-100 silica layers as compared to RP-18 W plates



a) RP-18 W, b) Nano-SIL C18-50, c) Nano-SIL C18-100
all plates with UV indicator
methanol – water; acetonitrile – water

Elution properties of MN RP plates in mixtures of methanol – water and acetonitrile – water

Further application examples can be found online in our application database at *www.mn-net.com/apps*

Modified silica layers

RP-18 W/UV_{254} G A octadecyl-modified HPTLC silica layers

Technical characteristics

- Nano silica 60, mean pore size 60 Å, specific surface (BET) ~ 500 m^2/g, specific pore volume 0.75 mL/g, particle size 2–10 µm, for preparative plates (1 mm thickness of layer) standard silica 60, pH stability 2–10, particle size 5–17 µm
- Indicator: acid-resistant product with a pale blue fluorescence for short-wave UV (254 nm), UV-absorbing substances appear as dark-blue to black spots on a light-blue background

Modification

- Partial octadecyl (C_{18}) modification, wettable with water, carbon content 14 %
- Order of polarity: silica > DIOL > NH_2 > CN > RP-2 > C18-50 > RP-18 W > C18-100

Recommended application

- NP or RP separation with eluents from anhydrous solvents to mixtures with high concentrations of water (see table and figure on previous page), relative polarity of the eluent determines the polarity of the layer
- Aminophenols, barbiturates, preservatives, nucleobases, polycyclic aromatic hydrocarbons, steroids, tetracyclines, plasticizers (phthalates)

Ordering information

Plate size [cm]	4 x 8	5 x 10	5 x 20	10 x 10	10 x 20	20 x 20	Thickness of layer	Fluorescent indicator
Glass plates								
Pack of [plates]			50	25	50	25		
RP-18 W/UV_{254}			811073	811075	811072	811071	0.25 mm	UV_{254}
Pack of [plates] (preparative TLC)						15		
RP-18 W/UV_{254}						811074	1.00 mm	UV_{254}
ALUGRAM® aluminum sheets								
Pack of [plates]	50	50	50	25		25		
RP-18 W/UV_{254}	818144	818152	818145	818147		818146	0.15 mm	UV_{254}

RP-2/UV_{254} G A “silanized silica” = dimethyl-modified standard silica layers

Technical characteristics

- Silica 60, mean pore size 60 Å, specific surface (BET) ~ 500 m^2/g, specific pore volume 0.75 mL/g, pH stability 2–10, particle size 5–17 µm
- Indicator: acid-resistant product with a pale blue fluorescence for short-wave UV (254 nm), UV-absorbing substances appear as dark-blue to black spots on a light-blue background

Modification

- Silanized silica with dimethyl modification, carbon content 4 %
- Order of polarity: silica > DIOL > NH_2 > CN > RP-2 > C18-50 > RP-18 W > C18-100

Recommended application

- Normal phase or reversed phase separation modes with purely organic, organic - aqueous or purely aqueous eluents
- Active plant constituents, steroids

Ordering information

Plate size [cm]	10 x 20	20 x 20	Thickness of layer	Fluorescent indicator
Pack of [plates]	50	25		
Glass plates				
RP-2/UV_{254}	811081	811082	0.25 mm	UV_{254}
ALUGRAM® aluminum sheets				
RP-2/UV_{254}		818171	0.15 mm	UV_{254}

Nano-SIL CN G A cyano-modified HPTLC silica layers

Technical characteristics

- Nano silica 60, mean pore size 60 Å, specific surface (BET) ~ 500 m^2/g, specific pore volume 0.75 mL/g, pH stability 2–8, particle size 2–10 µm
- Indicator: acid-resistant product with a pale blue fluorescence for short-wave UV (254 nm), UV-absorbing substances appear as dark-blue to black spots on a light-blue background

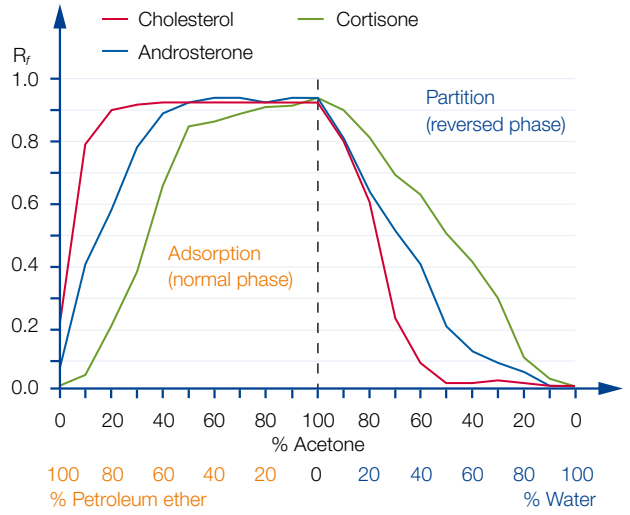
Modification

- Cyanopropyl modification, carbon content 5.5 %
- Order of polarity: silica > DIOL > NH_2 > CN > RP-2 > C18-50 > RP-18 W > C18-100

Recommended application

- NP or RP separation modes depending on the polarity of the developing solvent (see figure below)
- Steroid hormones, phenols, preservatives

R_f values of different steroids as a function of eluent composition



Layer: Nano-SIL CN/UV

Polarity of the eluent governs the type of separation mechanism:

Eluent system petroleum ether (PE) – acetone (NP mode)

the higher the concentration of PE, the stronger are the adsorptive interactions of the steroids with the stationary phase

Eluent system acetone – water (RP mode)

the sequence of elution of the steroids is reversed, the most nonpolar compounds are most strongly retained

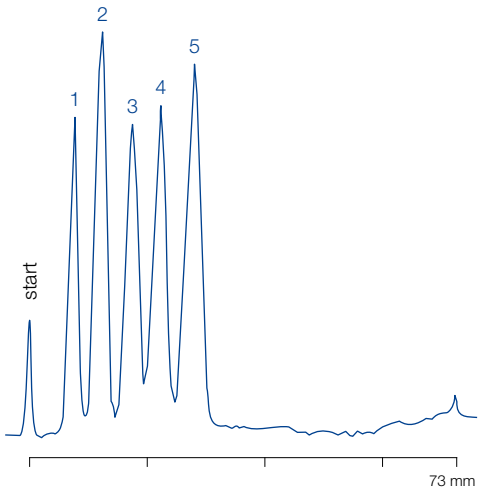
Separation of preservatives

MN Appl. No. 401440

Layer:	Nano-SIL CN/UV
Sample volume:	400 nL
Eluent:	ethanol – water – glacial acetic acid (20:80:0.2) with 0.1 mol/L tetraethylammonium chloride
Migration distance:	73 mm in 30 min
Detection:	TLC scanner, UV 254 nm

Peaks:

1. Propyl p-hydroxybenzoate
2. Ethyl p-hydroxybenzoate
3. Methyl p-hydroxybenzoate
4. Benzoic acid
5. Sorbic acid



Ordering information

Plate size [cm]	4 x 8	10 x 10	10 x 20	Thickness of layer	Fluorescent indicator
Pack of [plates]	50	25	25		
Glass plates					
Nano-SIL CN/UV		811115	811116	0.20 mm	UV_{254}
ALUGRAM® aluminum sheets					
Nano-SIL CN/UV	818184			0.15 mm	UV_{254}

Modified silica layers

Nano-SIL NH_2 G A amino-modified HPTLC silica layers

Technical characteristics

- Nano silica 60, mean pore size 60 Å, specific surface (BET) ~ 500 m^2/g, specific pore volume 0.75 mL/g, pH stability 2–8, particle size 2–10 µm
- Indicator: acid-resistant product with a pale blue fluorescence for short-wave UV (254 nm), UV-absorbing substances appear as dark-blue to black spots on a light-blue background

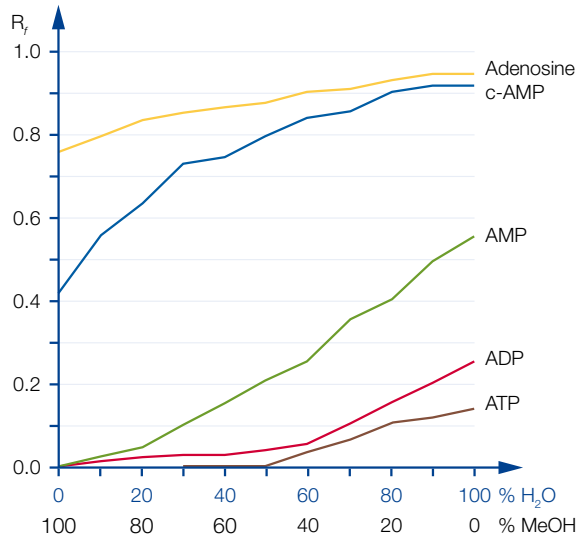
Modification

- Aminopropyl modification, carbon content 3.5 %
- Order of polarity: silica > DIOL > NH_2 > CN > RP-2 > C18-50 > RP-18 W > C18-100
- Layer can be wetted equally well with pure water as with organic solvents

Recommended application

- Vitamins, sugars, steroids, purine derivatives, xanthines, phenols, nucleotides and pesticides

Influence of eluent composition on the separation of nucleotides



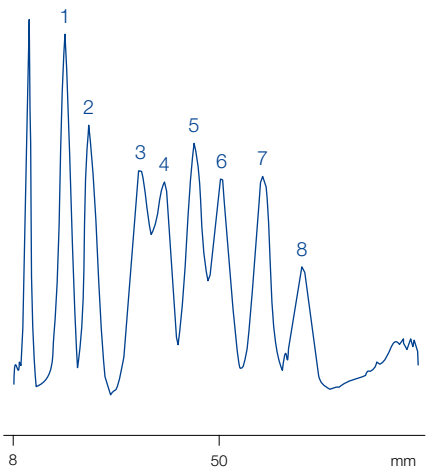
Layer:	Nano-SIL NH_2/UV
Eluent:	MeOH – H_2O according to fig. + 0.18 mol/L NaCl
Migration distance:	7 cm
c-AMP, AMP:	adenosine monophosphate
ADP:	adenosine diphosphate
ATP:	adenosine triphosphate

Separation of sugars

MN Appl. No. 401590

Layer:	Nano-SIL NH_2/UV
Sample volume:	0.5 µL
Eluent:	ethyl acetate – pyridine – water – glacial acetic acid (60:30:10:5, v/v/v/v)
Migration distance:	80 mm in 45 min, double development
Detection:	dry layer at 160 °C for 5 min, TLC scanner, UV 254 nm

Peaks (0.1 % each):
1. Lactose
2. Saccharose
3. Galactose
4. Glucose
5. Fructose
6. Arabinose
7. Xylose
8. Ribose



Ordering information

Plate size [cm]	4 x 8	10 x 10	10 x 20	Thickness of layer	Fluorescent indicator
Pack of [plates]	50	25	25		
Glass plates					
Nano-SIL NH_2/UV		811111	811112	0.20 mm	UV_{254}
ALUGRAM® aluminum sheets					
Nano-SIL NH_2/UV	818182			0.15 mm	UV_{254}

Further application examples can be found online in our application database at *www.mn-net.com/apps*

Nano-SIL DIOL G diol-modified HPTLC silica layers

Technical characteristics

- Nano silica 60, mean pore size 60 Å, specific surface (BET) ~ 500 m^2/g, specific pore volume 0.75 mL/g, pH stability 2–8, particle size 2–10 µm
- Indicator: acid-resistant product with a pale blue fluorescence for short-wave UV (254 nm), UV-absorbing substances appear as dark-blue to black spots on a light-blue background

Modification

- Diol modification, carbon content 5.5 %
- Order of polarity: silica > DIOL > NH_2 > CN > RP-2 > C18-50 > RP-18 W > C18-100
- Layer can be wetted equally well with pure water as with organic solvents

Recommended application

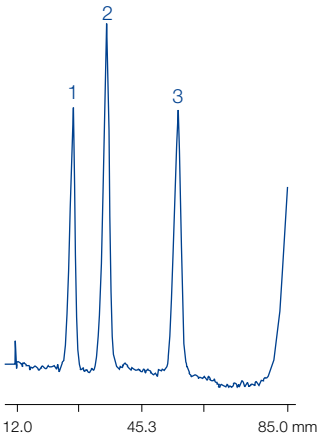
- Steroids, pesticides and plant constituents
- For critical separations an alternative to silica
- Since it is less sensitive to the water content of the environment, leads to more reproducible results compared to silica

Separation of herbicides

MN Appl. No. 401950

Layer:	Nano-SIL DIOL/UV
Sample volume:	2 µL
Eluent:	petroleum ether (40–60 °C) – acetone (80:20, v/v)
Migration distance:	70 mm
Detection:	TLC scanner, 230 nm

Peaks:
(0.07 % each in methanol)
1. Metoxuron
2. Monuron
3. Metobromuron



Ordering information

Plate size [cm]	10 x 10	Thickness of layer	Fluorescent indicator
Pack of [plates]	25		
Glass plates			
Nano-SIL DIOL/UV	811120	0.20 mm	UV_{254}

Further layers

Alox G P A aluminum oxide layers

Technical characteristics

- Aluminum oxide, mean pore size 60 Å, specific surface (BET) ~ 200 m^2/g
- Inert organic binder
- Indicator: manganese-activated zinc silicate

Recommended application

- Terpenes, alkaloids, steroids, aliphatic and aromatic compounds
- We recommend to activate aluminum oxide layers before use by heating 10 minutes at 120 °C

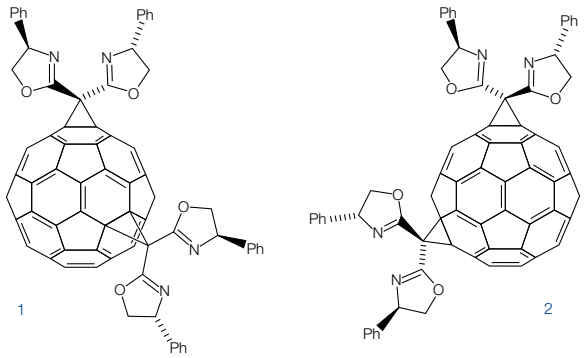
Separation of bisadducts of fullerenes

MN Appl. No. 401930

F. Djojo, A. Hirsch, Chem. Eur. J. 4 (1998), 344–356

Layer: ALUGRAM® Alox N/UV_{254}
Eluent: toluene – ethyl acetate (95:5, v/v)
Detection: UV, 254 nm

Compound	R_f values
Bis[bis(4-phenyloxazolin)methane]fullerene 1	0.14
Bis[bis(4-phenyloxazolin)methane]fullerene 2	0.26

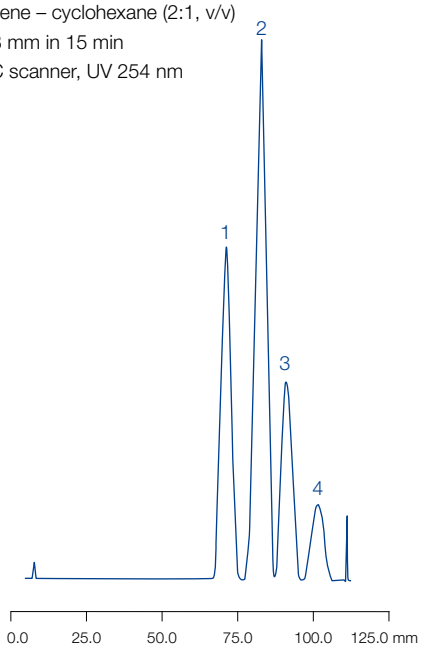


Separation of lipophilic dyes

MN Appl. No. 403010

Layer: Alox-25 UV_{254}
Sample volume: 1000 nL
Eluent: toluene – cyclohexane (2:1, v/v)
Migration distance: 108 mm in 15 min
Detection: TLC scanner, UV 254 nm

Peaks:
1. Indophenol
2. Sudan red G
3. Sudan blue II
4. Butter yellow



Ordering information

Plate size [cm]	4 x 8	5 x 20	20 x 20	Thickness of layer	Fluorescent indicator
Glass plates					
Pack of [plates]		100	25		
Alox-25 UV_{254}		807021	807023	0.25 mm	UV_{254}
Pack of [plates] (preparative TLC)			15		
Alox-100 UV_{254}			807033	1.00 mm	UV_{254}
POLYGRAM® polyester sheets					
Pack of [plates]	50	50	25		
Alox N/UV_{254}	802021	802022	802023	0.20 mm	UV_{254}
ALUGRAM® aluminum sheets					
Pack of [plates]		50	25		
Alox N/UV_{254}		818024	818023	0.20 mm	UV_{254}

Further application examples can be found online in our application database at *www.mn-net.com/apps*

Cellulose MN 300 G P A native fibrous cellulose layers

Technical characteristics

- Fiber length (95 %) 2–20 µm, average degree of polymerization 400–500, specific surface acc. to Blaine 15 000 cm^2/g, ≤ 20 ppm Fe, 6 ppm Cu, 7 ppm P; CH_2Cl_2- extract ≤ 0.25 %; residue on ignition at 850 °C ≤ 1500 ppm

Recommended application

- Partition chromatography of polar substances such as amino acids, carboxylic acids or carbohydrates

Ordering information

Plate size [cm]	4 x 8	5 x 20	20 x 20	Thickness of layer	Fluorescent indicator
Glass plates					
Pack of [plates]			25		
CEL 300-10			808013	0.10 mm	–
CEL 300-10 UV_{254}			808023	0.10 mm	UV_{254}
CEL 300-25			808033	0.25 mm	–
CEL 300-25 UV_{254}			808043	0.25 mm	UV_{254}
Pack of [plates] (preparative TLC)			20		
CEL 300-50			808053	0.50 mm	–
CEL 300-50 UV_{254}			808063	0.50 mm	UV_{254}
POLYGRAM® polyester sheets					
Pack of [plates]	50	50	25		
CEL 300	801011		801013	0.10 mm	–
CEL 300 UV_{254}		801022	801023	0.10 mm	UV_{254}
ALUGRAM® aluminum sheets					
Pack of [plates]	50	50	25		
CEL 300	818155		818153	0.10 mm	–
CEL 300 UV_{254}		818157	818156	0.10 mm	UV_{254}

Cellulose MN 400 (AVICEL®) G P microcrystalline cellulose layers

Technical characteristics

- Prepared by hydrolysis of high purity cellulose with HCl, average degree of polymerization 40–200

Recommended application

- Carboxylic acids, lower alcohols, urea and purine derivatives

Ordering information

Plate size [cm]	10 x 20	20 x 20	Thickness of layer	Fluorescent indicator
Pack of [plates]	50	25		
Glass plates				
CEL 400-10	808072	808073	0.10 mm	–
POLYGRAM® polyester sheets				
CEL 400		801113	0.10 mm	–
CEL 400 UV_{254}		801123	0.10 mm	UV_{254}

Further layers

Cellulose MN 300 PEI P PEI-impregnated cellulose ion exchange layers

Technical characteristics

- Fibrous cellulose impregnated with polyethyleneimine

Recommended application

- Analysis of nucleic acids, and of mutagenic substances with the ^{32}P postlabelling procedure

Ordering information

Plate size [cm]	20 x 20	Thickness of layer	Fluorescent indicator
Pack of [plates]	25		
POLYGRAM® polyester sheets			
CEL 300 PEI	801053	0.10 mm	–
CEL 300 PEI/UV_{254}	801063	0.10 mm	UV_{254}

Cellulose MN 300 AC P acetylated cellulose layers

Technical characteristics

- Fibrous cellulose with 10 % content of acetylated cellulose for reversed phase chromatography

Recommended application

- Reversed phase chromatography

Ordering information

Plate size [cm]	Acetyl content	20 x 20	Thickness of layer	Fluorescent indicator
Pack of [plates]		25		
POLYGRAM® polyester sheets				
CEL 300 AC-10 %	10 %	801033	0.10 mm	–

Polyamid-6 P ε-polycaprolactame layers

Technical characteristics

- Polyamide 6 = nylon 6 = perlon = ε-aminopolycaprolactame
- Separation mechanism based on hydrogen bonds to amide groups of the polymer matrix as well as on ionic, dipole and electron donor-acceptor interactions

Recommended application

- Natural compounds, phenols, carboxylic acids, aromatic nitro compounds and especially amino acids

Ordering information

Plate size [cm]	5 x 20	20 x 20	Thickness of layer	Fluorescent indicator
Pack of [plates]	50	25		
POLYGRAM® polyester sheets				
POLYAMID-6	803012	803013	0.10 mm	–
POLYAMID-6 UV_{254}	803022	803023	0.10 mm	UV_{254}

Further application examples can be found online in our application database at *www.mn-net.com/apps*

CHIRALPLATE G special layer enantiomer separation

Technical characteristics

- Reversed phase nano silica impregnated with Cu^{2+} ions and a chiral selector (proline derivative)
- Separation based on ligand exchange, i.e. formation of ternary mixed-ligand complexes with the Cu(II) ions, differences in the stability of the diastereomeric complexes cause chromatographic separation

Recommended application

- Enantiomer separation of amino acids, *N*-methylamino acids, *N*-formylamino acids, α-alkylamino acids, thiazolidine derivatives, dipeptides, lactones, α-hydroxycarboxylic acids

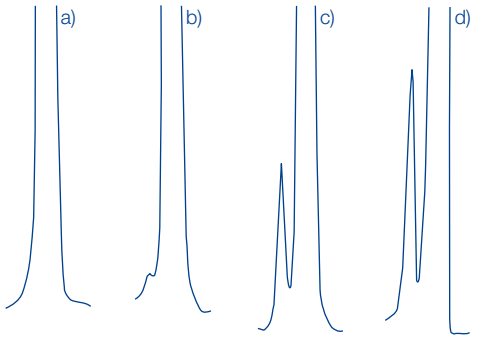
Enantiomer separation of amino acids

MN Appl. No. 400520

Quantitative determination (remission location curves) of TLC-separated enantiomers of *tert*.-leucine:

Layer:	CHIRALPLATE
Eluent:	methanol – water (10:80, v/v)
Detection:	dip in 0.3 % ninhydrin solution quantification with scanner, 520 nm

a) L-*tert*.-leucine
b) L-*tert*.-leucine + 0.1 % D-*tert*.-leucine
c) L-*tert*.-leucine + 1 % D-*tert*.-leucine
d) external reference sample



Ordering information

Plate size [cm]	5 x 20	10 x 10	10 x 20	20 x 20	Thickness of layer	Fluorescent indicator
Glass plates						
Pack of [plates]			4			
CHIRALPLATE			811056		0.25 mm	UV_{254}
Pack of [plates]	50	25	25	25		
CHIRALPLATE	811057	811059	811055	811058	0.25 mm	UV_{254}

SIL N-HR P unmodified standard silica layers

Technical characteristics

- High purity silica 60, mean pore size 60 Å, specific surface (BET) ~ 500 m^2/g, specific pore volume 0.75 mL/g, particle size 5–17 µm, different binder system compared to SIL G results in different separation characteristics
- A special feature of the POLYGRAM® SIL N-HR is a higher gypsum content

Ordering information

Plate size [cm]	5 x 20	20 x 20	Thickness of layer	Fluorescent indicator
Pack of [plates]	50	25		
POLYGRAM® polyester sheets				
SIL N-HR/UV_{254}	804022	804023	0.20 mm	UV_{254}

Layers for special TLC separations

SIL G-25 HR G special layer for aflatoxin separation

Technical characteristics

- High purity silica 60 with gypsum and a very small quantity of a polymeric organic binder; softer than the standard silica layer, i.e. spots can be scratched and the layer absorbs faster

Recommended application

- Aflatoxins

Ordering information

Plate size [cm]	20 x 20	Thickness of layer	Fluorescent indicator
Pack of [plates]	25		
Glass plates			
SIL G-25 HR	809033	0.25 mm	–
SIL G-25 HR/UV_{254}	809043	0.25 mm	UV_{254}

SIL G-25 Tenside G special layer for separation of surfactants

Technical characteristics

- Silica G impregnated with ammonium sulfate

Recommended application

- Detergents, alkanesulfonates, polyglycols

Ordering information

Plate size [cm]	20 x 20	Thickness of layer	Fluorescent indicator
Pack of [plates]	25		
Glass plates			
SIL G-25 Tenside	810063	0.25 mm	–

Nano-SIL PAH G special HPTLC silica layer for PAH analysis

Technical characteristics

- Nano silica 60, mean pore size 60 Å, specific surface (BET) ~ 500 m^2/g, specific pore volume 0.75 mL/g, particle size 2–10 µm
- Impregnated with caffeine, an electron acceptor for PAH analysis based on charge-transfer complexes

Recommended application

- 6 PAHs according to German drinking water specifications (TVO) in accordance with German standard DIN 38407 part 7

Ordering information

Plate size [cm]	10 x 20	Thickness of layer	Fluorescent indicator
Pack of [plates]	50		
Glass plates			
Nano-SIL PAH	811051	0.20 mm	–

Further application examples can be found online in our application database at *www.mn-net.com/apps*

IONEX P special mixed layers of silica with ion exchange resins

IONEX-25 SA-Na:

- Mixture of silica and a strongly acidic cation exchanger coated to polyester sheets

IONEX-25 SB-AC:

- Mixture of silica and a strongly basic anion exchanger coated to polyester sheets
- Both layers contain an inert organic binder

Recommended application

- Amino acids, e.g., in protein and peptide hydrolyzates, in seeds and fodder, in biological fluids; for racemate separation in peptide syntheses, for the separation of nucleic acid hydrolyzates, aminosugars, amino acids, antibiotics, inorganic phosphates, cations and other compounds with ionic groups

Ordering information

Plate size [cm]		20 x 20	Thickness of layer	Fluorescent indicator
Pack of [plates]		25		
POLYGRAM® polyester sheets				
IONEX-25 SA-Na	strongly acidic cation exchanger	806013	0.20 mm	–
IONEX-25 SB-AC	strongly basic anion exchanger	806023	0.20 mm	–

Mixed layers for TLC G

Alox/CEL-AC-Mix-25:

- Mixed layer of aluminum oxide G and acetylated cellulose, recommended for separation of PAH

SILCEL-Mix-25:

- Mixed layer of cellulose and silica, recommended for separation of preservatives and other antimicrobial compounds

Ordering information

Plate size [cm]	20 x 20	Thickness of layer	Fluorescent indicator
Pack of [plates]	25		
Glass plates			
Alox/CEL-AC-Mix-25	810053	0.25 mm	–
SILCEL-Mix-25 UV_{254}	810043	0.25 mm	UV_{254}

Further application examples can be found online in our application database at *www.mn-net.com/apps*

Chromatography papers

Chromatography papers

Chromatography papers

- Paper chromatography is the oldest chromatographic technique separation due to partition of the analytes between special paper grades and the mobile phase, which penetrates the paper by capillary action ascending.
- Descending and circular techniques are possible

Please note

- Always treat chromatography papers with care
- Never touch them with fingers, because this will contaminate the surface
- Do not bend them sharply, because this will decrease the capillary action (preferably store them flat)

Direction

- Chromatography papers possess a preferred direction of the fibers with higher absorption properties (with our sheets 58 x 60 cm, the longer edge)
- We recommend to use them in the direction of higher absorption

Ordering information

Code	Weight [g/m^2]	Thickness [mm]	Description	Flow rate	Size [cm]	Pack of	REF
MN 214	140	0.28	smooth	90–100 mm/30 min	58 x 60	100 sheets	817001
MN 218	180	0.36	smooth	90–100 mm/30 min	58 x 60	100 sheets	817002
MN 260	90	0.20	smooth	120–130 mm/30 min	58 x 60	100 sheets	817003
MN 261	90	0.18	smooth	90–100 mm/30 min	58 x 60	100 sheets	817004
MN 827	270	0.70	soft carton	130–140 mm/10 min	58 x 60	100 sheets	817005
MN 866	650	1.70	soft carton	100–120 mm/10 min	38 x 38	100 sheets	817006
MN 866	650	1.70	soft carton	100–120 mm/10 min	80 x 80	100 sheets	817007
MN 214 ff	140	0.28	MN 214 defatted *	90–100 mm/30 min	56 x 58	100 sheets	817008

* This paper is extracted with organic solvents.

For further papers, filters and membranes, feel free to ask for our catalog “Filtration”.

Accessories

- Beside ready-to-use layers for thin layer chromatography also accessories are required
- Selection of accessories for reliable separation in TLC

Ordering information

Designation	Pack of	REF
Simultaneous developing chamber for TLC, 20 x 20 cm	1	814019
Simultaneous developing chamber for TLC, 10 x 10 cm	1	814018
Developing chambers for TLC micro-sets	4	814021
Glass laboratory sprayer with rubber bulb	1	814101
Glass capillaries 1 µL	3 x 50	814022
Rubber caps for capillaries	2	814102
Plastic syringe, 1 mL content with graduation	1	814104
Spotting guides	2	814023
Measuring cylinders, glass, 10 mL content	2	814024
MN ALUGRAM® scissors, ground blade, black handle	1	818666
Filter paper MN 713, 15 x 21 cm	100	814103
Folded filters MN 615 1/4, 11 cm diameter	100	531011
Chromatography paper MN 260, 7.5 x 17 cm (for chamber saturation)	100	814030

Visualization reagents

- Small selection of frequently used spray reagents for post chromatographic detection reactions in TLC suited for spraying or dipping TLC plates
- A detailed description of many more detection procedures for TLC is available on request

Ordering information

Spray reagent	Solvent	Detection of	Pack of	REF
Aniline phthalate	2-propanol – ethanol (1:1)	reducing sugars, oxohalic acids	100 mL	814919
Bromocresol green	2-propanol	organic acids	100 mL	814920
Reagent for caffeine detection	water – acetone	caffeine	100 mL	814401
2',7'-Dichlorofluorescein	2-propanol	lipids (saturated, unsaturated)	100 mL	814921
4-(Dimethylamino)-benzaldehyde	2-propanol	terpenes, sugars, steroids	100 mL	814922
Reagent according to Dragendorff-Munier	water	alkaloids and other nitrogen compounds	100 mL	814402
Iron(III) chloride	water	phenolic compounds e.g., acetylsalicylic acid, paracetamol	100 mL	814403
Potassium hexacyanoferrate(III)	water		100 mL	814404
Molybdatophosphoric acid	ethanol	lipids, sterols, steroids, reducing compounds	100 mL	814302
Ninhydrin	ethanol	amino acids, amines and amino sugars	100 mL	814203
Rhodamine B	ethanol	lipids	100 mL	814923
Rubeanic acid	ethanol	heavy metal cations	100 mL	814206

These products contain harmful substances which must be specially labeled as hazardous. For detailed information please see SDS.

Fluorescent indicators

UV indicators with efficient radiation for short-wave as well as long-wave UV ranges

- UV_{254}: manganese-activated zinc silicate with absorption maximum at 254 nm, green fluorescence, relatively susceptible towards acids: its fluorescence can be completely quenched by acidic solvents
- UV_{366}: inorganic fluorescent pigment with absorption maximum at 366 nm, blue fluorescence

Ordering information

	Composition	Absorption maximum	Color of fluorescence	Pack of 100 g
Fluorescent indicator UV_{254}	manganese-activated zinc silicate	254 nm	green	816710.01
Fluorescent indicator UV_{366}	inorganic fluorescent pigment	366 nm	blue	816720.01

Silica adsorbent for TLC

Pore size 60 Å, pore volume 0.75 mL/g, specific surface (BET) ~ 500 m^2/g, pH 7 for a 10 % aqueous suspension

- Silica G: standard grade, particle size 2–20 µm, Fe < 0.02 %, Cl < 0.02 %, 13 % gypsum as binder
- Silica N: standard grade, particle size 2–20 µm, Fe < 0.02 %, Cl < 0.02 %, no binder
- Silica G-HR: high purity grade, particle size 3–20 µm, Fe < 0.002 %, Cl < 0.008 %, gypsum as binder
- Silica P: preparative grade, particle size 5–50 µm, Fe < 0.02 %, Cl < 0.02 %, organic binder
- Silica P with gypsum: preparative grade, particle size 5–50 µm, Fe < 0.02 %, Cl < 0.02 %, gypsum as binder

Ordering information

Designation	Fluorescent indicator	1 kg	5 kg
Silica G	–	816310.1	816310.5
Silica G/UV_{254}	UV_{254}	816320.1	816320.5
Silica N	–	816330.1	816330.5
Silica N/UV_{254}	UV_{254}	816340.1	816340.5
Silica G-HR	–	816410.1	816410.5
Silica P/UV_{254}	UV_{254}	816380.1	816380.5
Silica P/UV_{254} with gypsums	UV_{254}	816400.1	816400.5

Polyamid adsorbent for TLC

Polyamide 6 = nylon 6 = perlon = ε-polycaprolactame

Ordering information

Designation	Fluorescent indicator	1 kg
Polyamid-DC 6	–	816610.1
Polyamid-DC 6 UV_{254}	UV_{254}	816620.1

Cellulose MN 301 native fibrous cellulose

- Standard grade, fiber length (95 %) 2–20 µm
- Average degree of polymerization 400–500, specific surface acc. to Blaine 15 000 cm^2/g
- ≤ 20 ppm Fe, 6 ppm Cu, 7 ppm P, CH_2Cl_2 extract ≤ 0.25 %, residue on ignition at 850 °C ≤ 1500 ppm

Ordering information

Designation	1 kg	5 kg
Cellulose MN 301	816250.1	816250.5

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본 사

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